

# Research on the origins of life: How to handle this problem? Complex systems

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FLinT Center, SDU

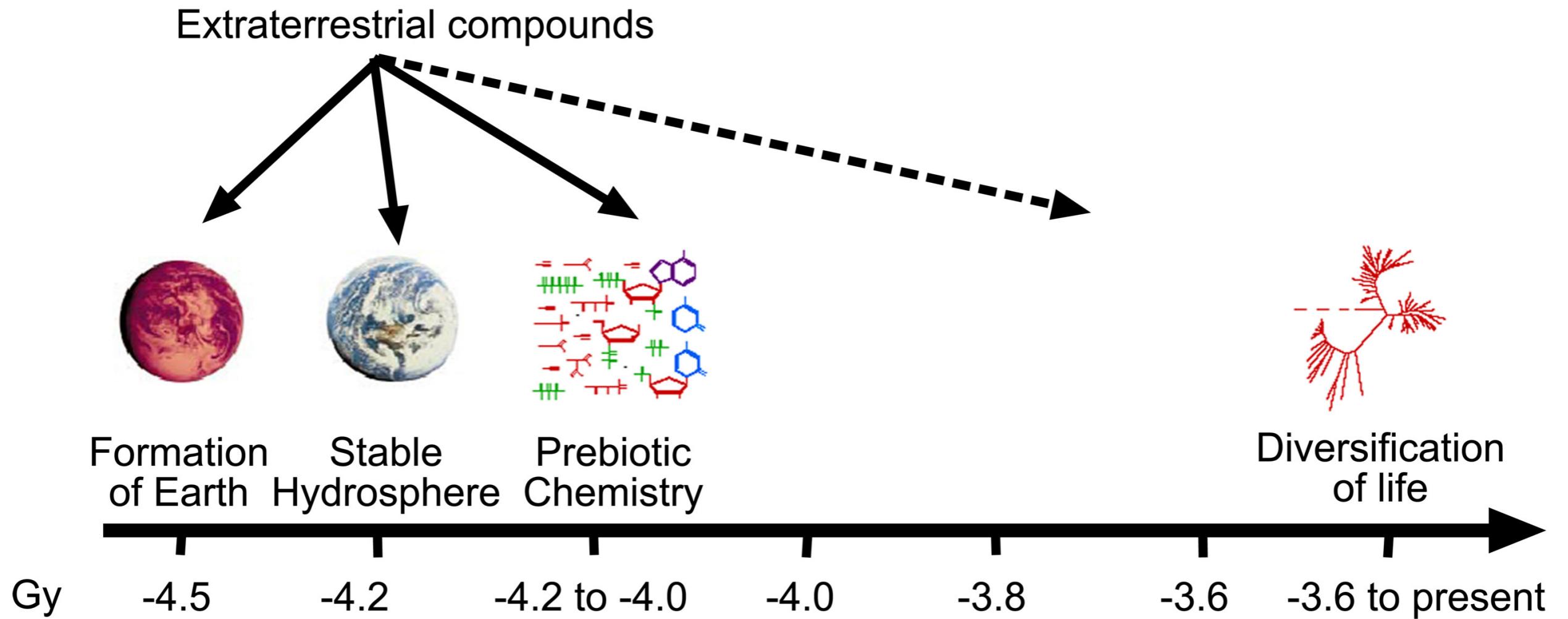
[monnard@ifk.sdu.dk](mailto:monnard@ifk.sdu.dk)

CERN, Geneva, May 20, 2011



UNIVERSITY OF SOUTHERN DENMARK

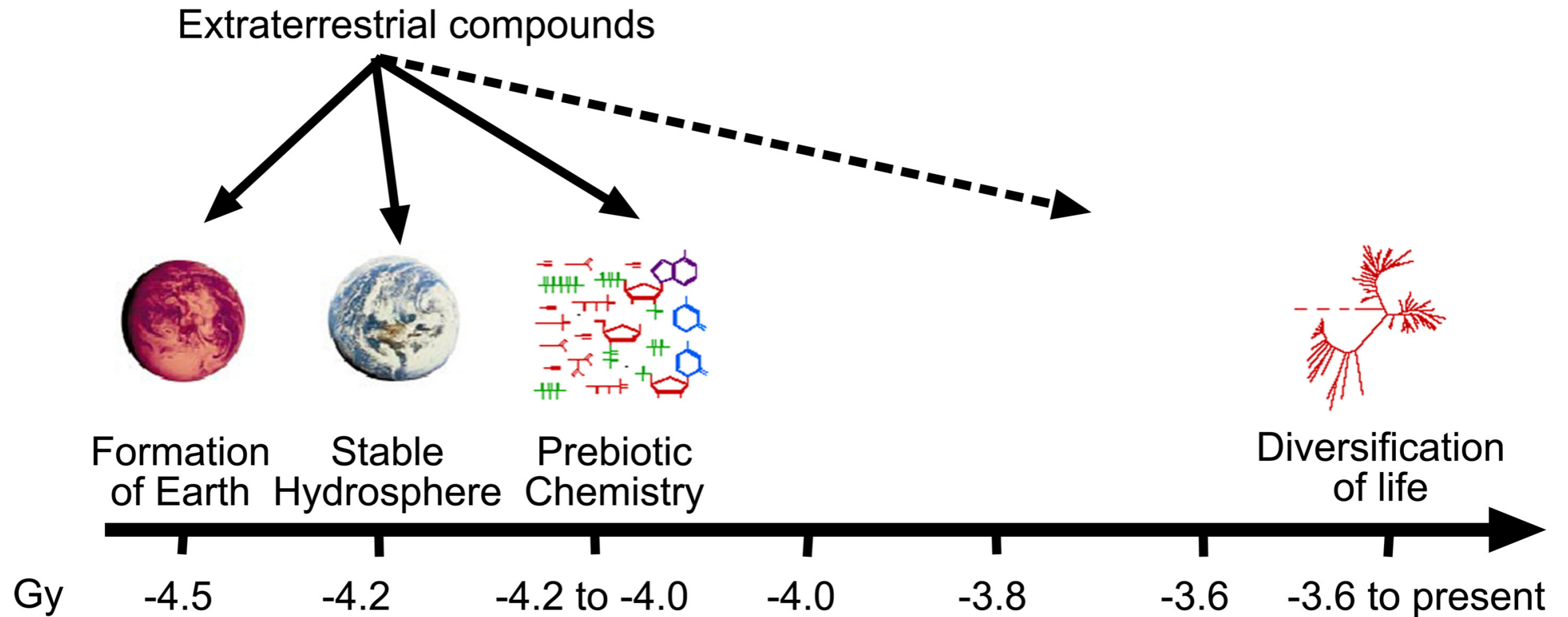
# Origins of Life: Scenario



*Adapted from G.F. Joyce, 2002 Nature, 418, 214.*



# Origins of Life: Scenario

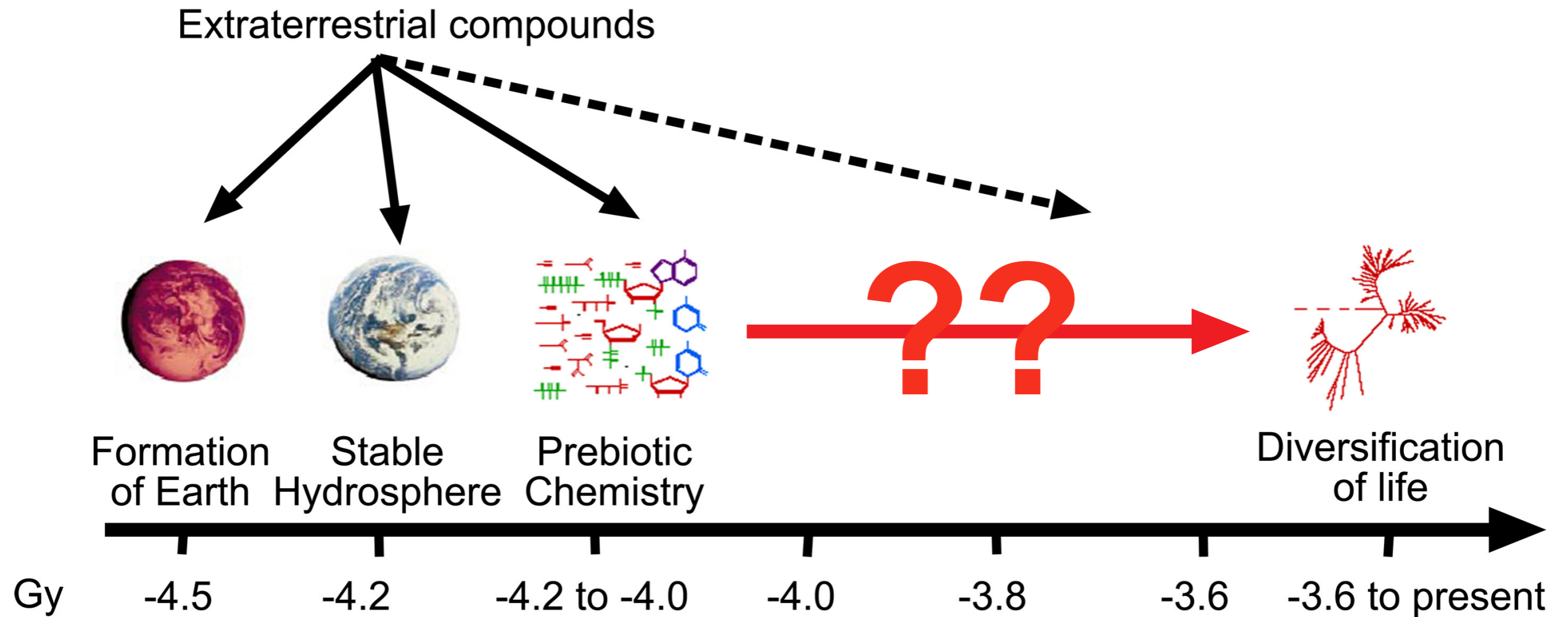


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LCA set was based on membranous container and the triad DNA-RNA-protein



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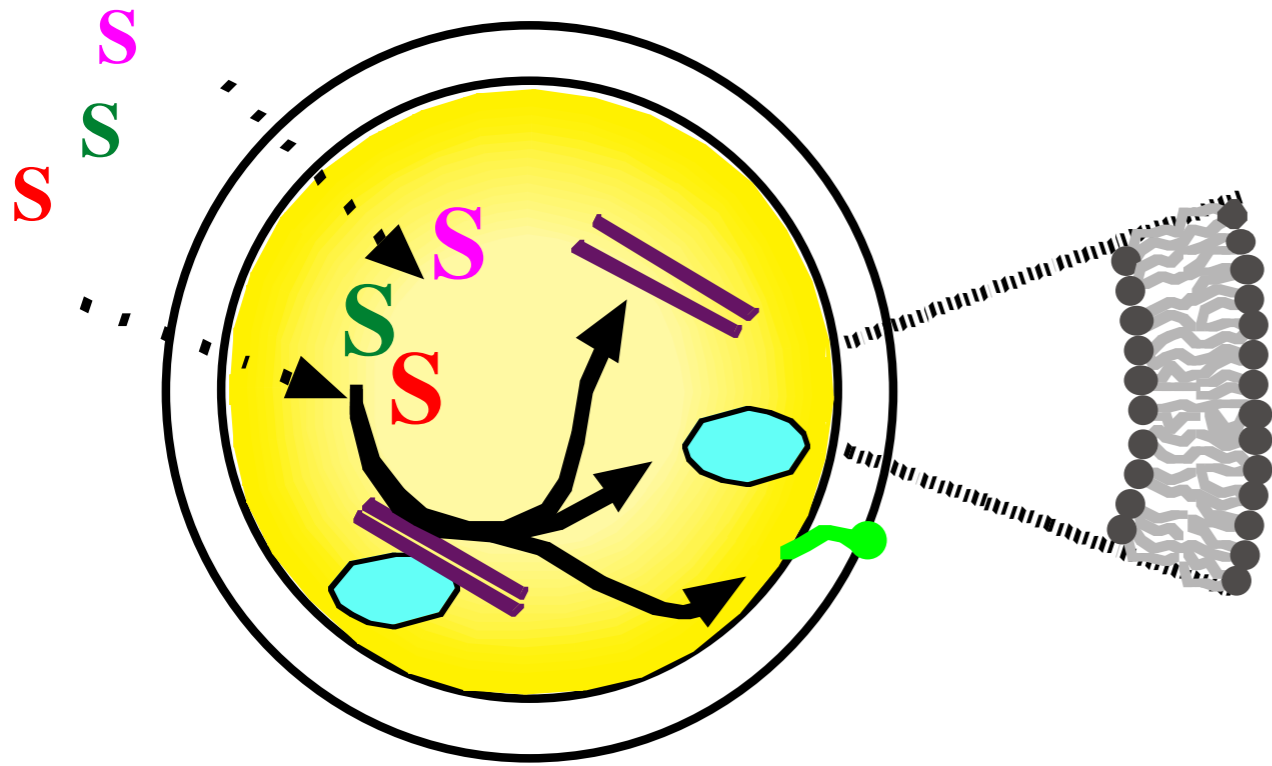


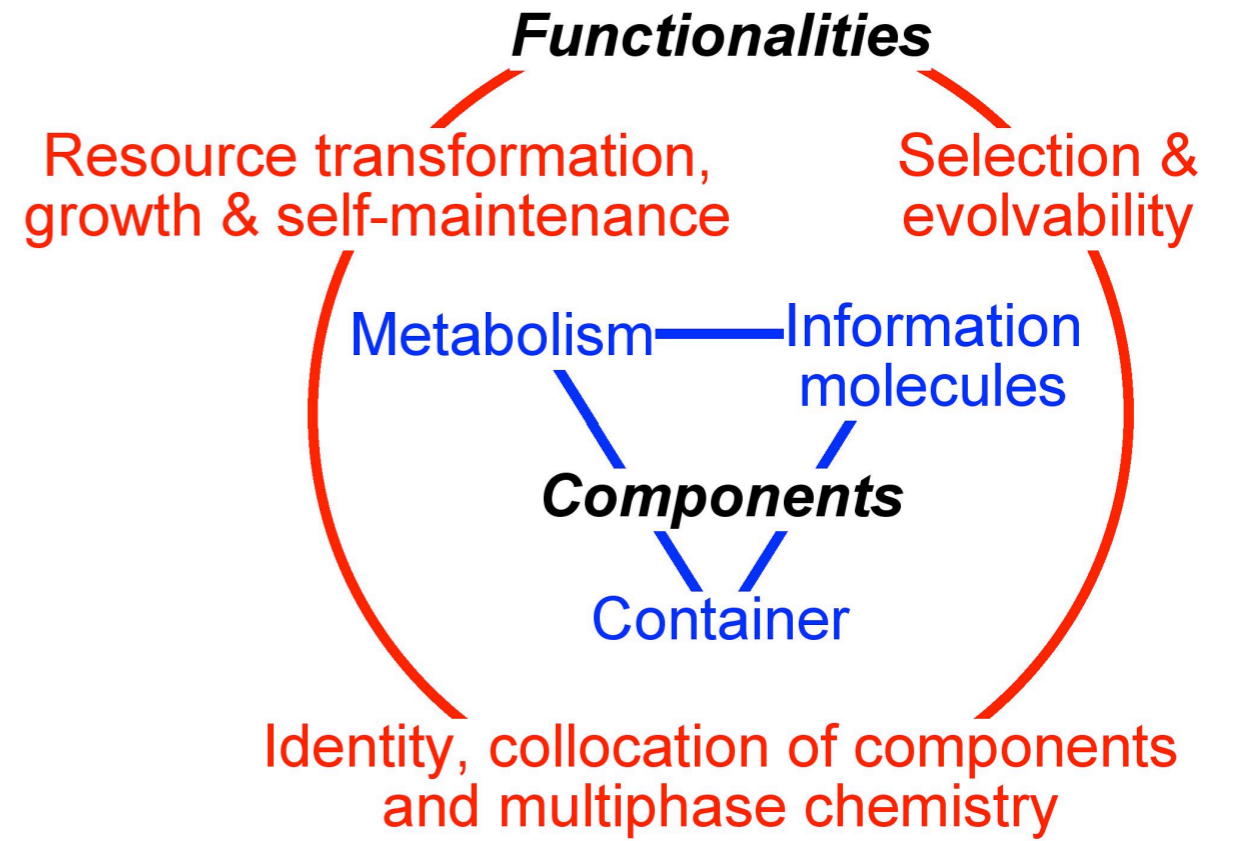
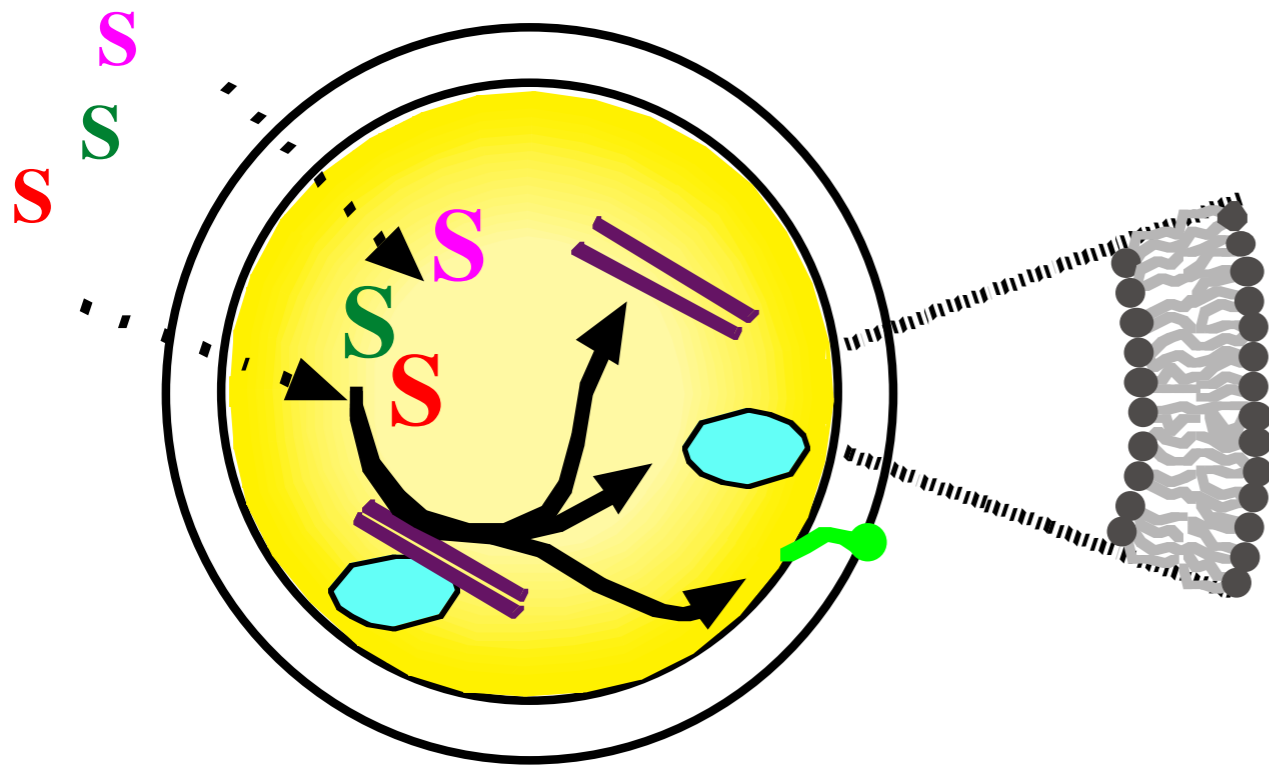
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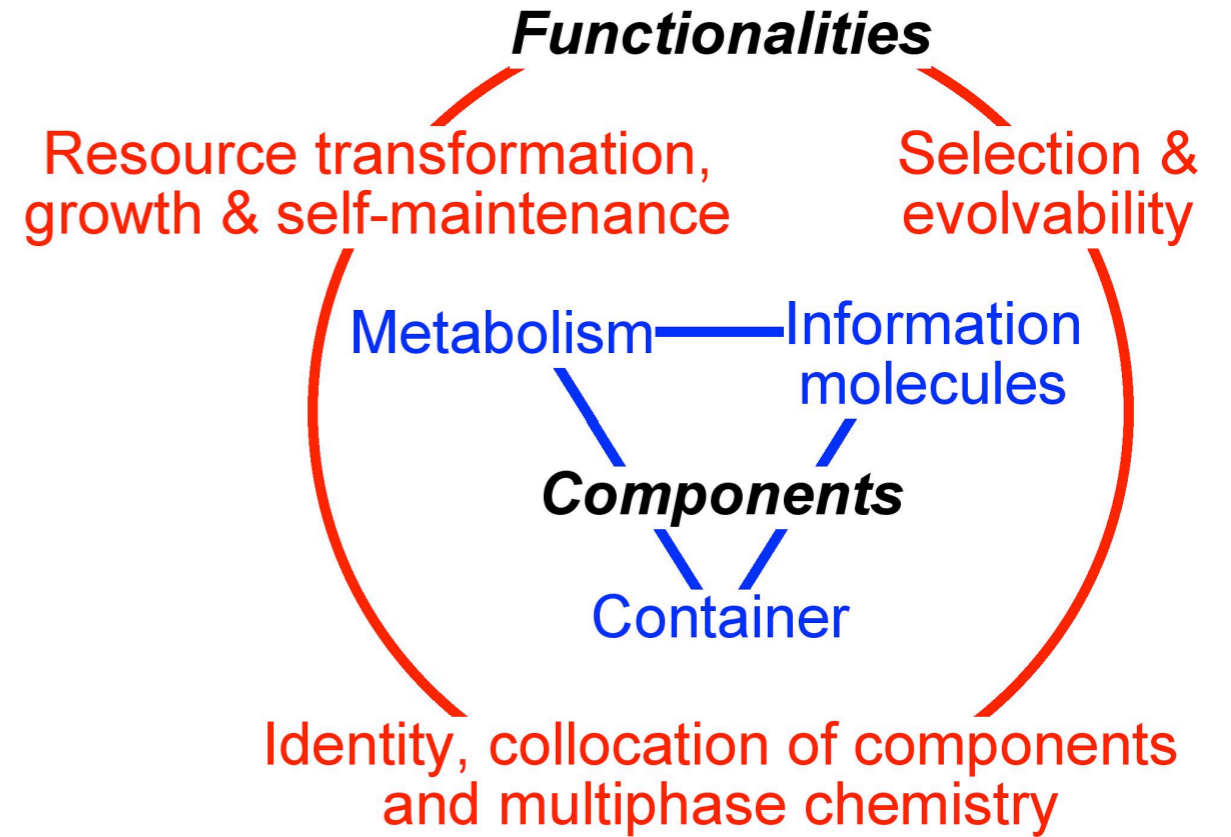
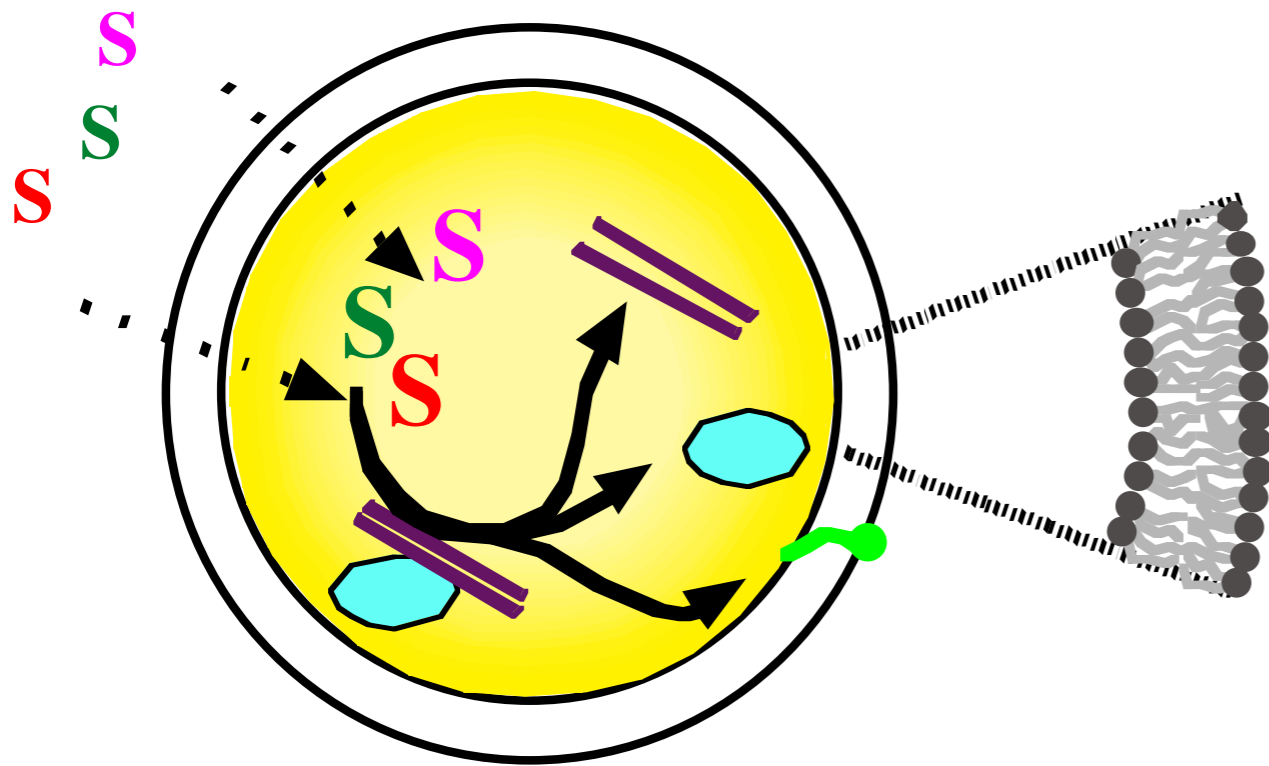
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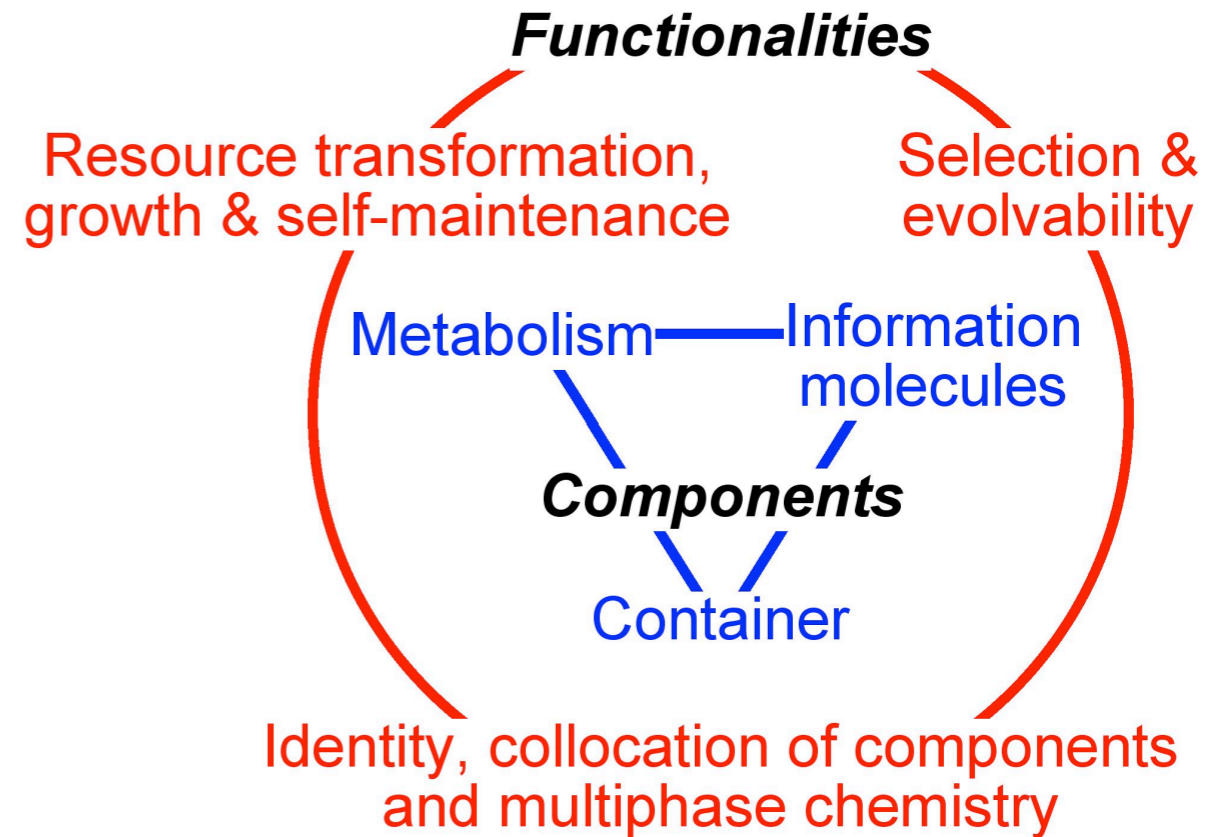
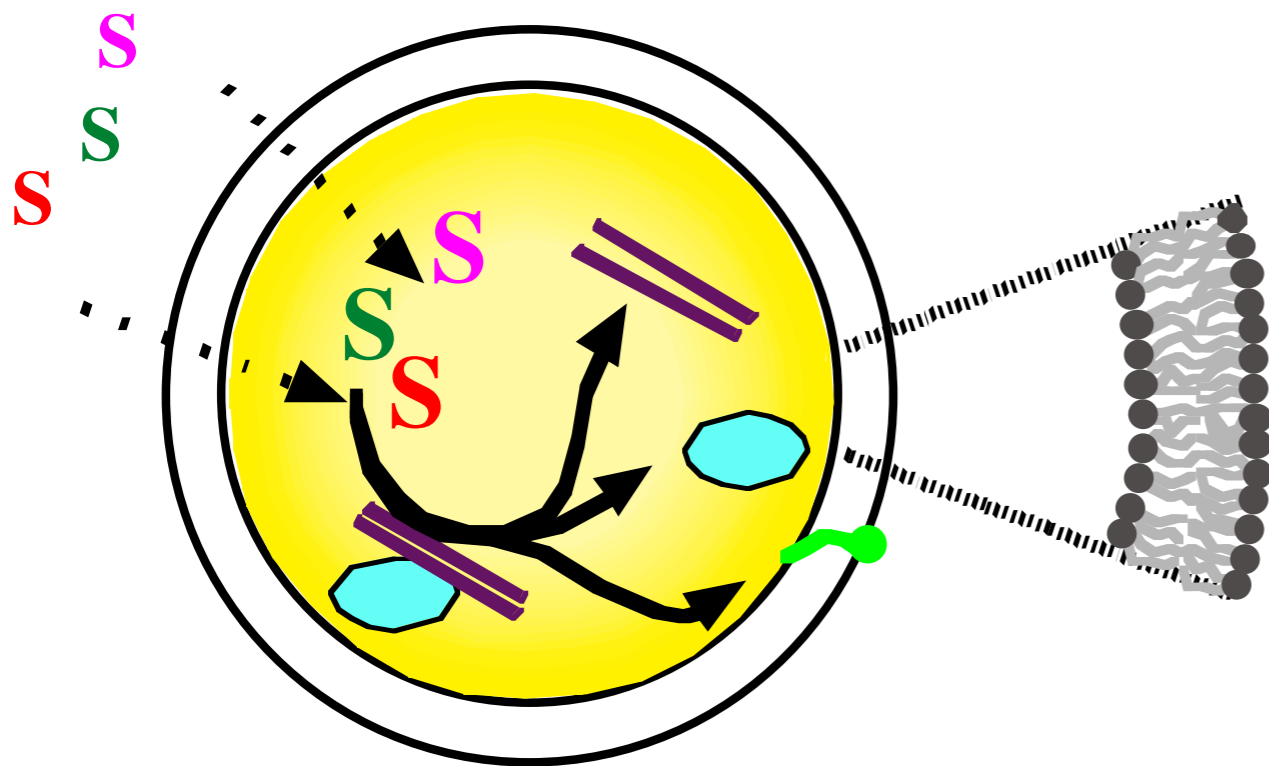
**Self-maintenance** (*Resource transformation, identity preservation*)

**Self-replication** (*Growth and division*)

**Evolution** (*Trait selection*)







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In the systemic approach to building a minimal self-replicating chemical system, only a system composed of these three components can mimics the systemic and functional properties of living systems



# Procell concept: Components (bottom-up approach)

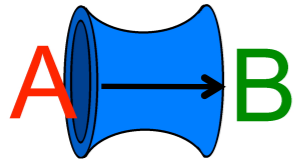
Metabolism

Container

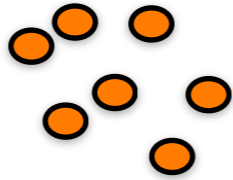
Information

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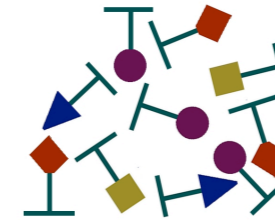
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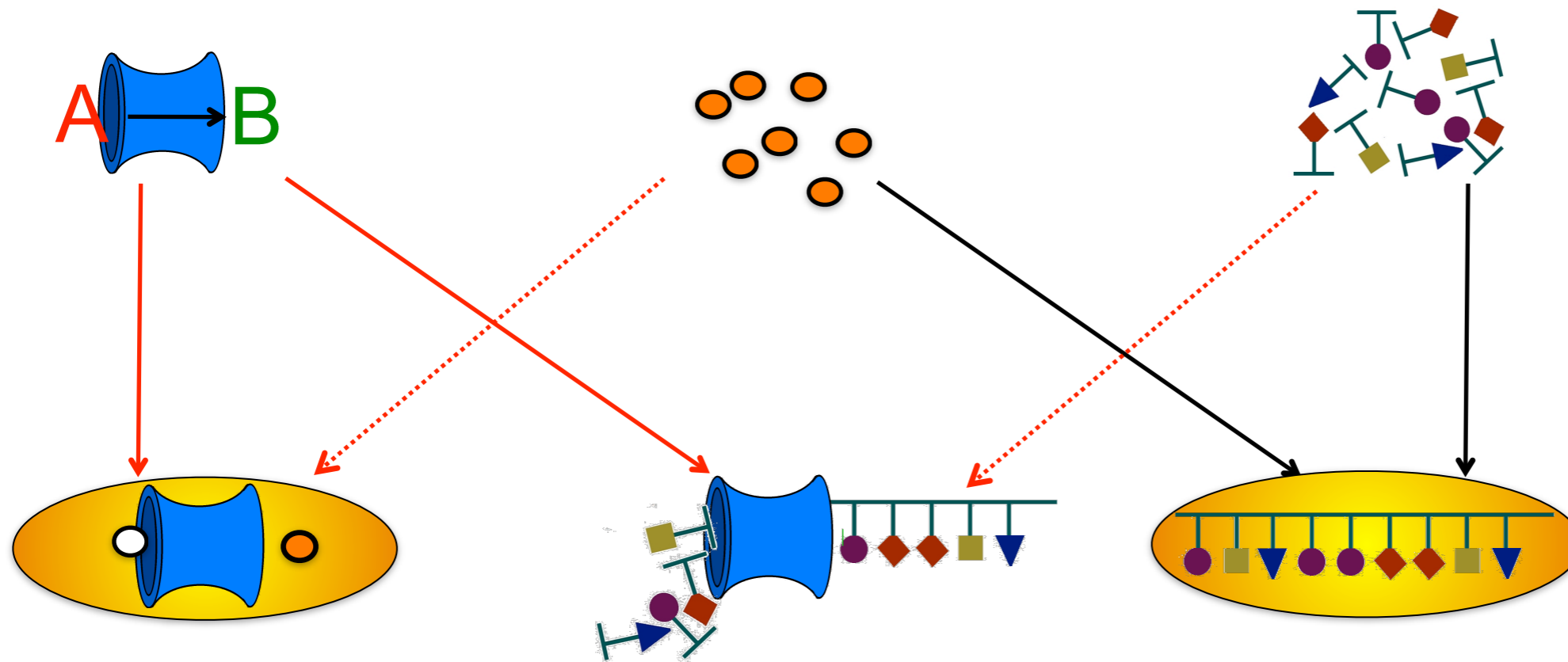


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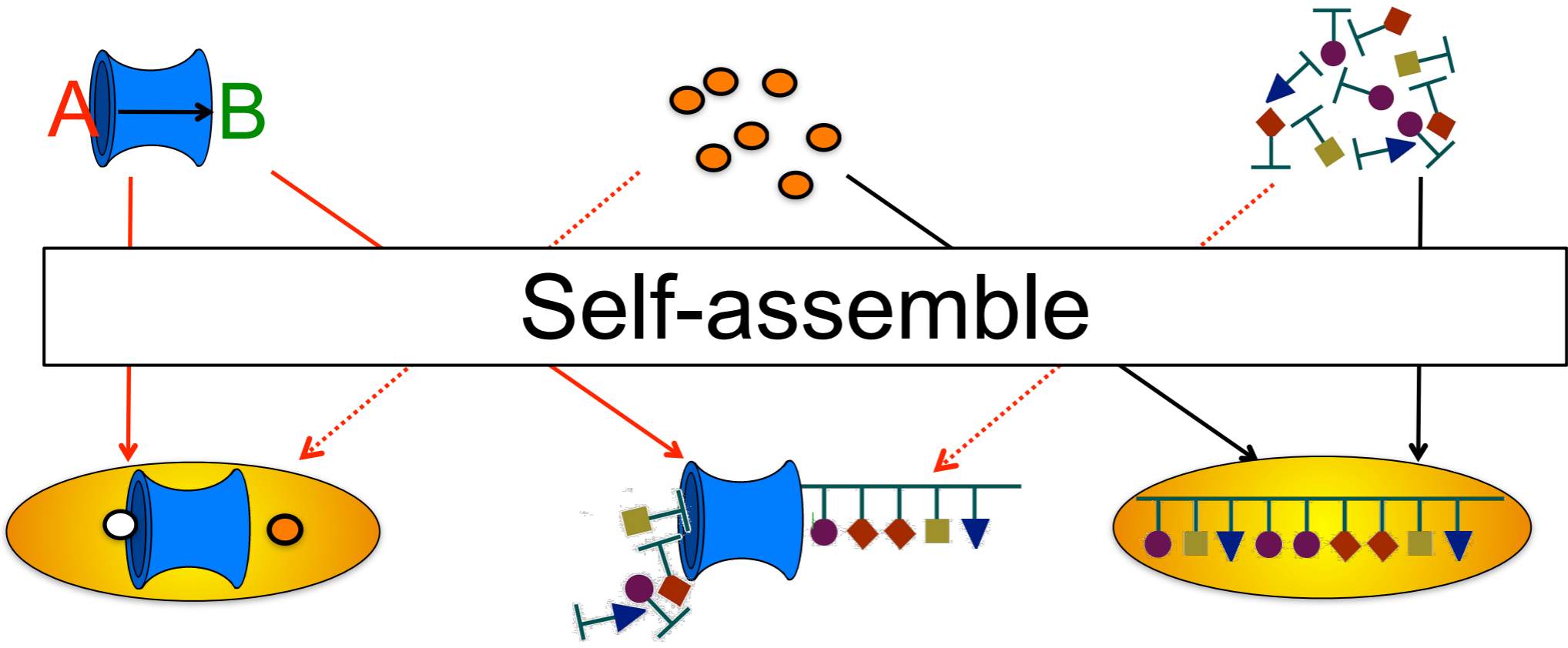


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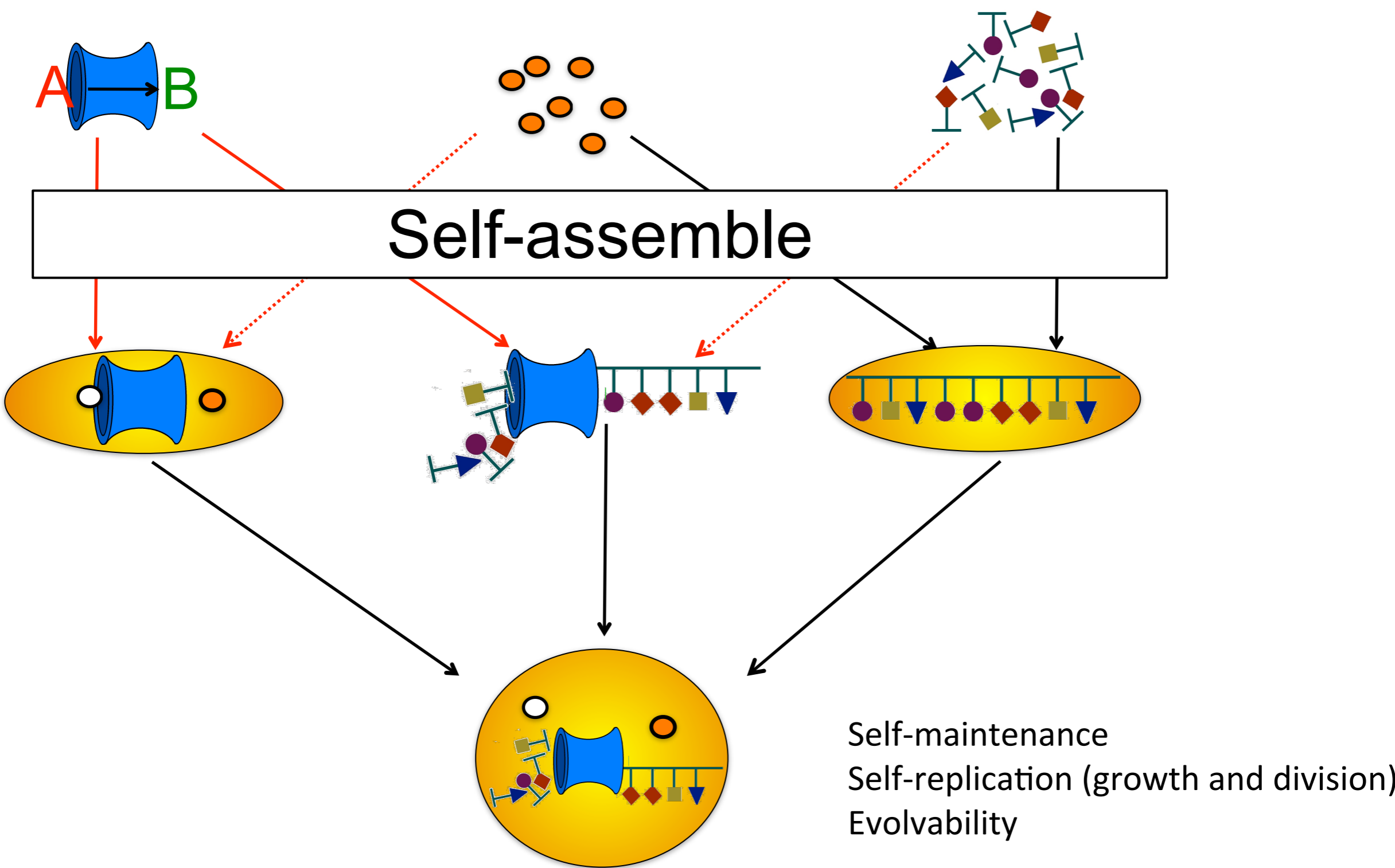


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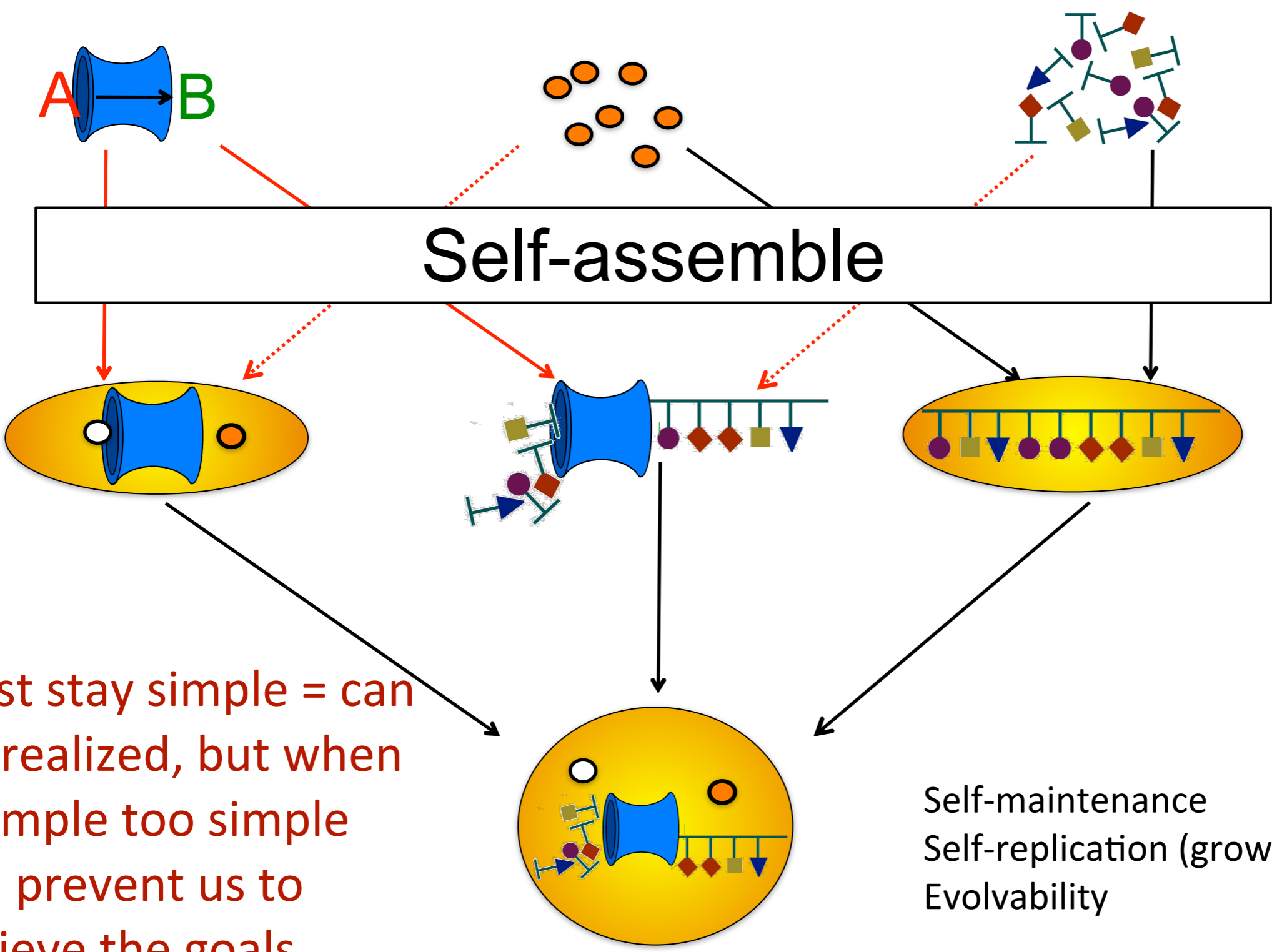


# Protocell concept: Components (bottom-up approach)

Metabolism

Container

Information



Must stay simple = can be realized, but when is simple too simple and prevent us to achieve the goals.

Self-maintenance  
Self-replication (growth and division)  
Evolvability

# Primitive membranous compartments: Self-assembly of prebiotically plausible amphiphiles

**Mixture of various single chain amphiphiles  
(chain length, headgroup functions and numbers)**





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## Environmental impacts on single-chain amphiphile membranes

- **pH** (*Apel et al 2002 Biochim. Biophys. Acta*)
- **Dilution (CVC)** (*Maurer et al 2009 Astrobiology*)
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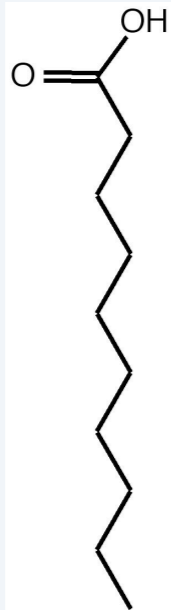
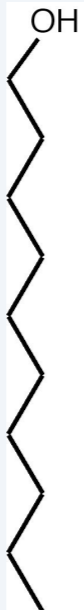
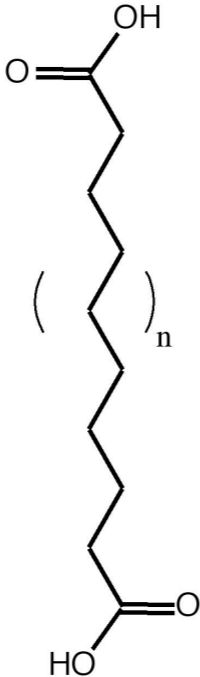
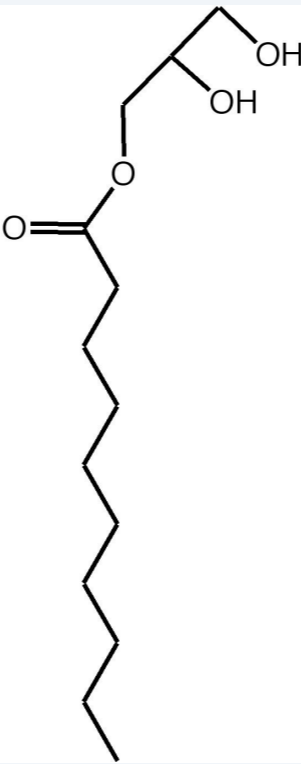
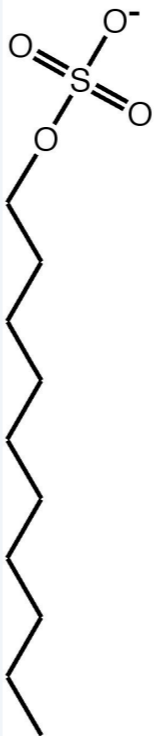
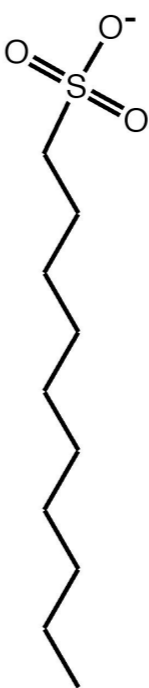
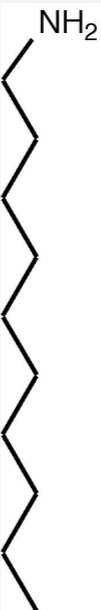
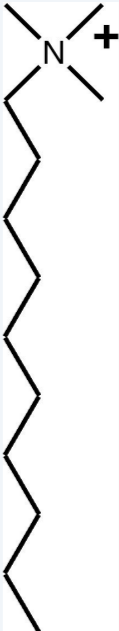
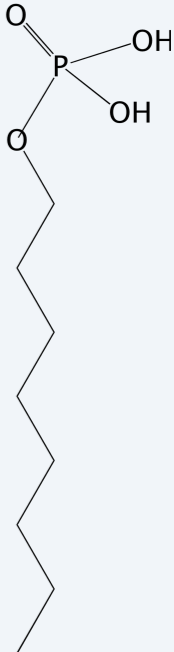
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**Fluctuations in these parameters occurred frequently**

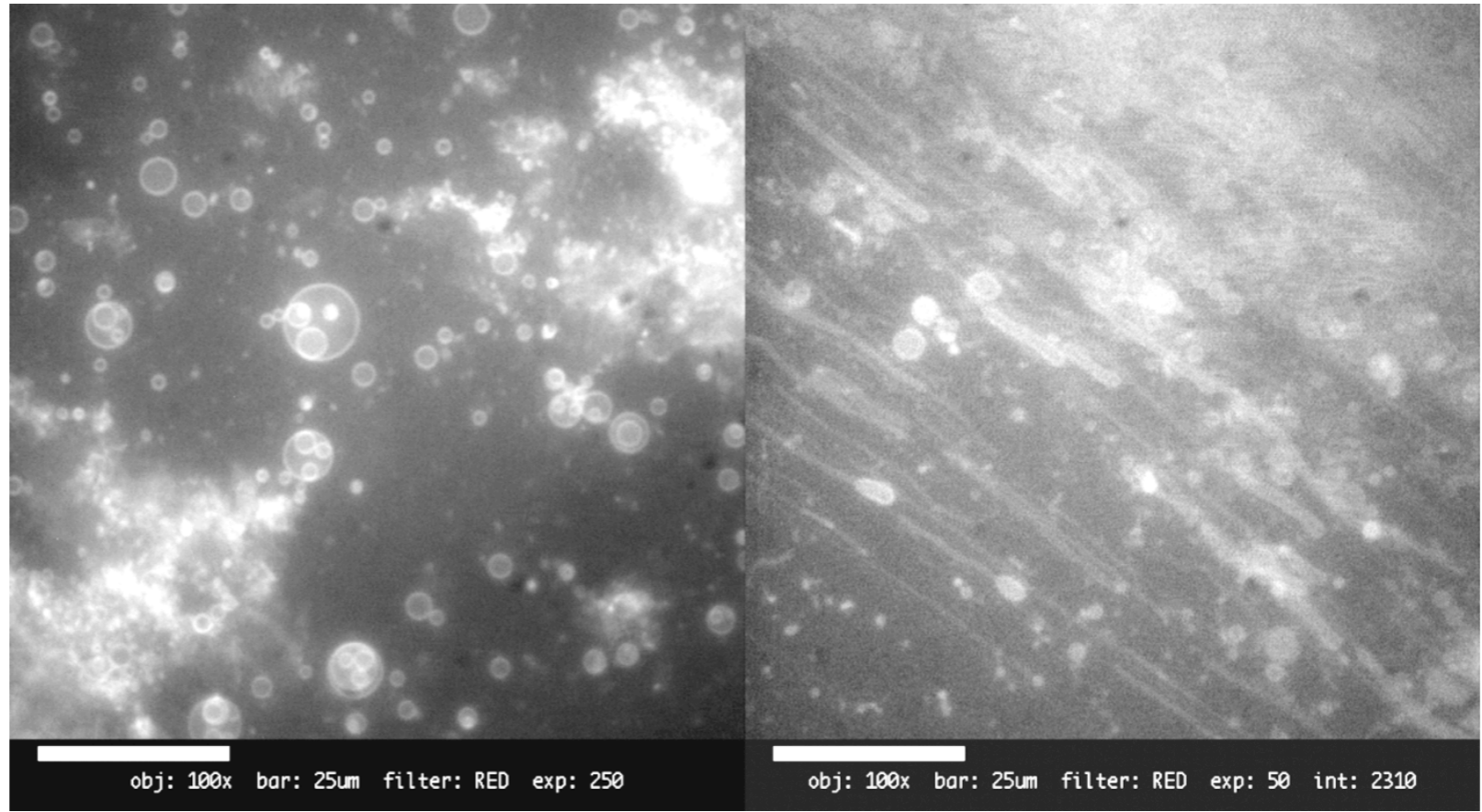
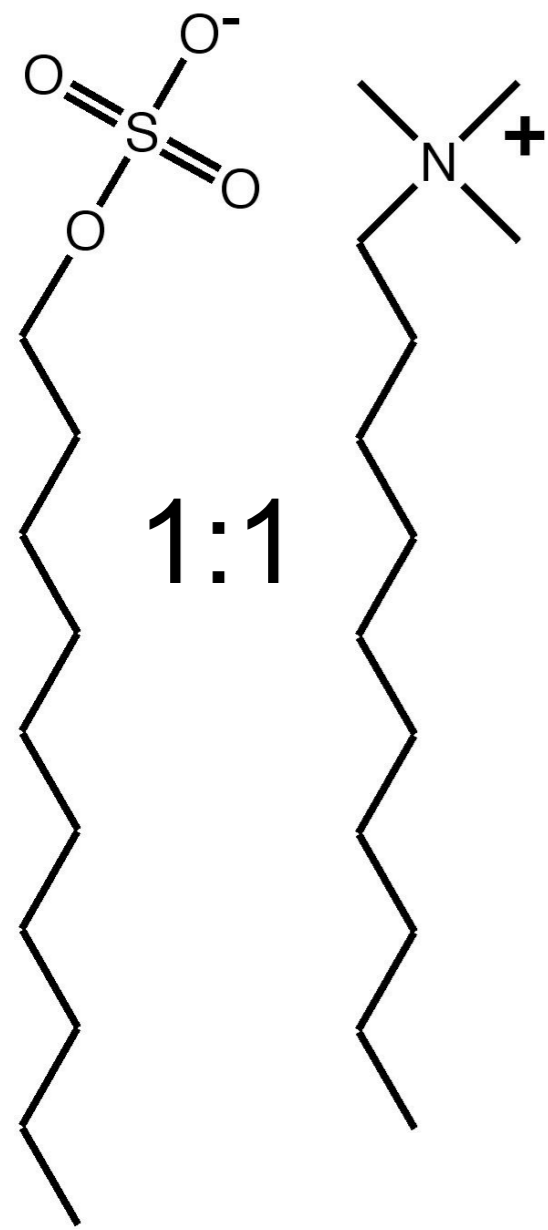


# Chemical structures

									
Headgroup	COOH	OH	Di COOH (bola)	Glycerol- ester	SO <sub>4</sub> <sup>-</sup>	SO <sub>3</sub> <sup>-</sup>	NH <sub>2</sub>	N(CH <sub>3</sub> ) <sub>3</sub> <sup>+</sup>	PO <sub>4</sub> <sup>2-</sup>
In Chondrites	☺	☺	☺	?	?	?	☹	☹	?
Prebiotic synthesis	☺	☺	?	☺	?	?	?	?	☺



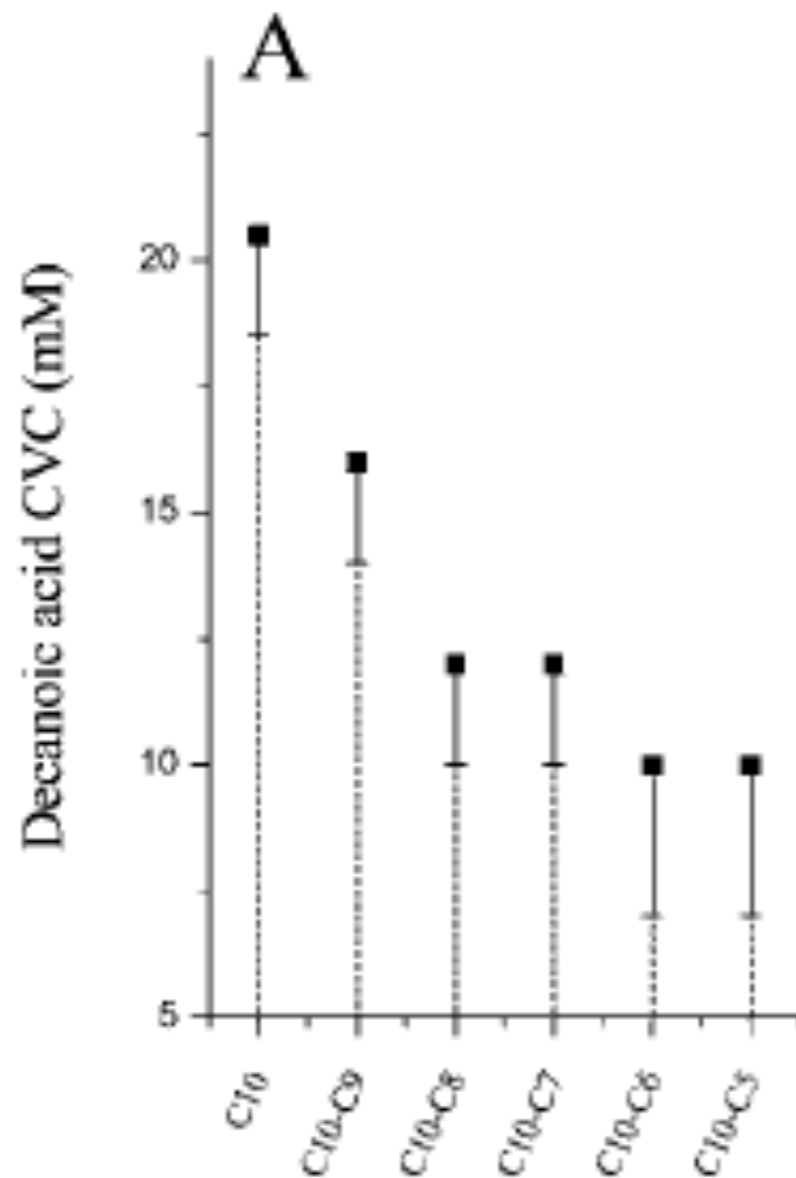
# Mixtures of single chain amphiphiles that do not form membranous structures on their own



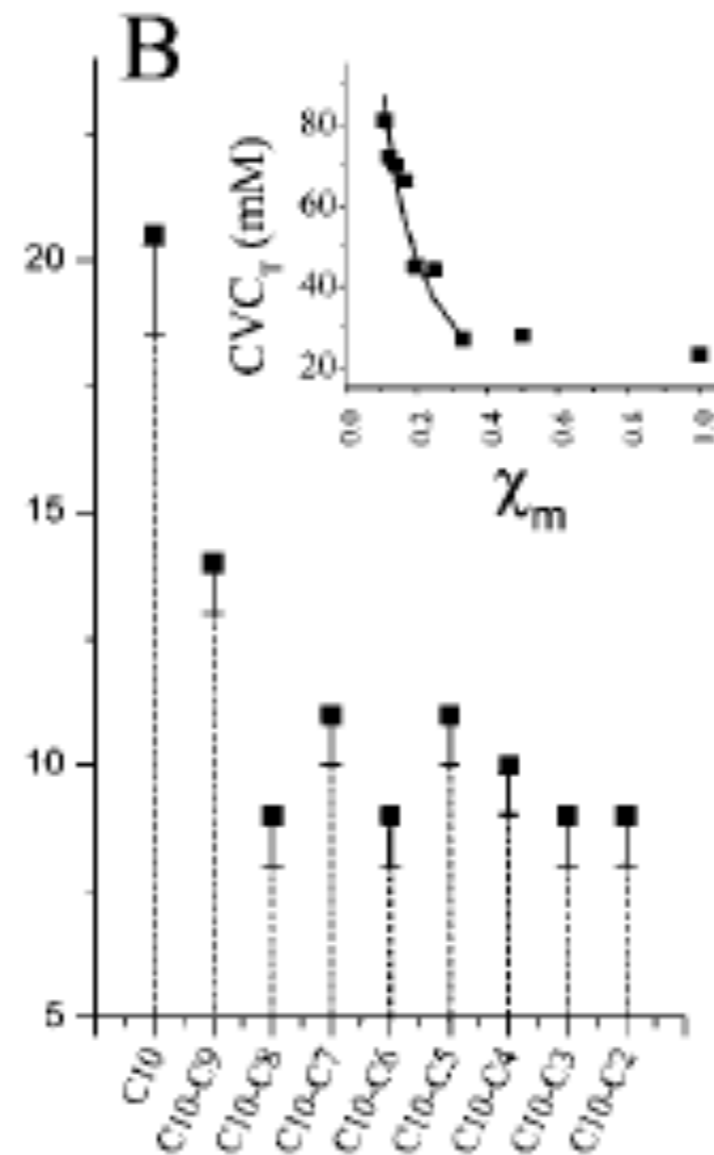
pH = 11.0 to 3.0

*T.E. Rasmussen*

# More complex mixtures: Prebiotically plausible mixture of carboxylic acids

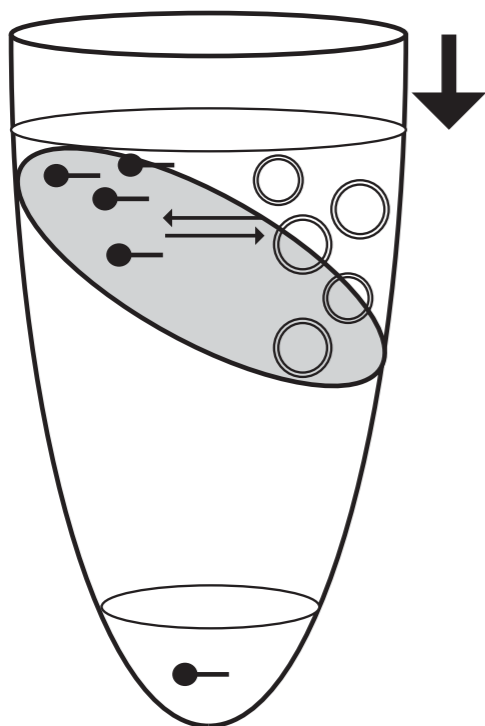
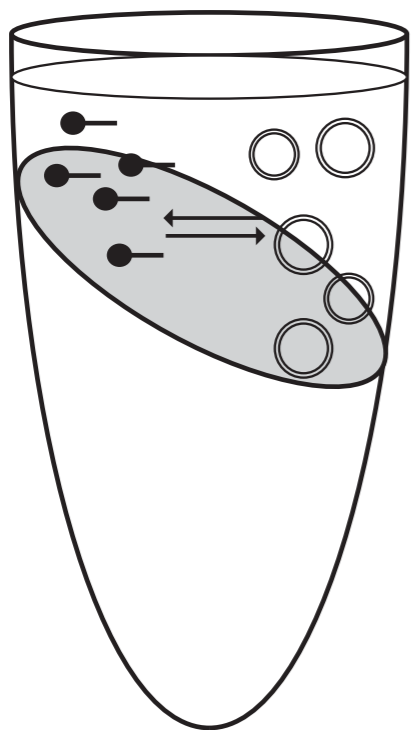


A)  $CVC_{DA}$  in mixtures of constant concentration of the other CAs



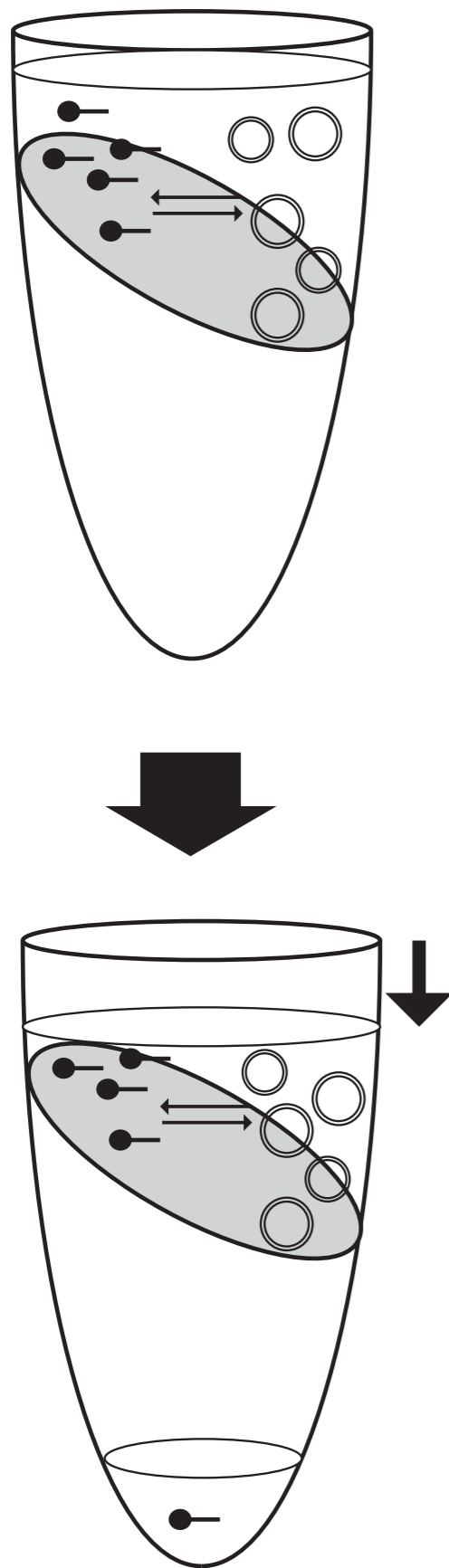
B)  $CVC_{DA}$  in mixtures where all CA concentration ratios are maintained



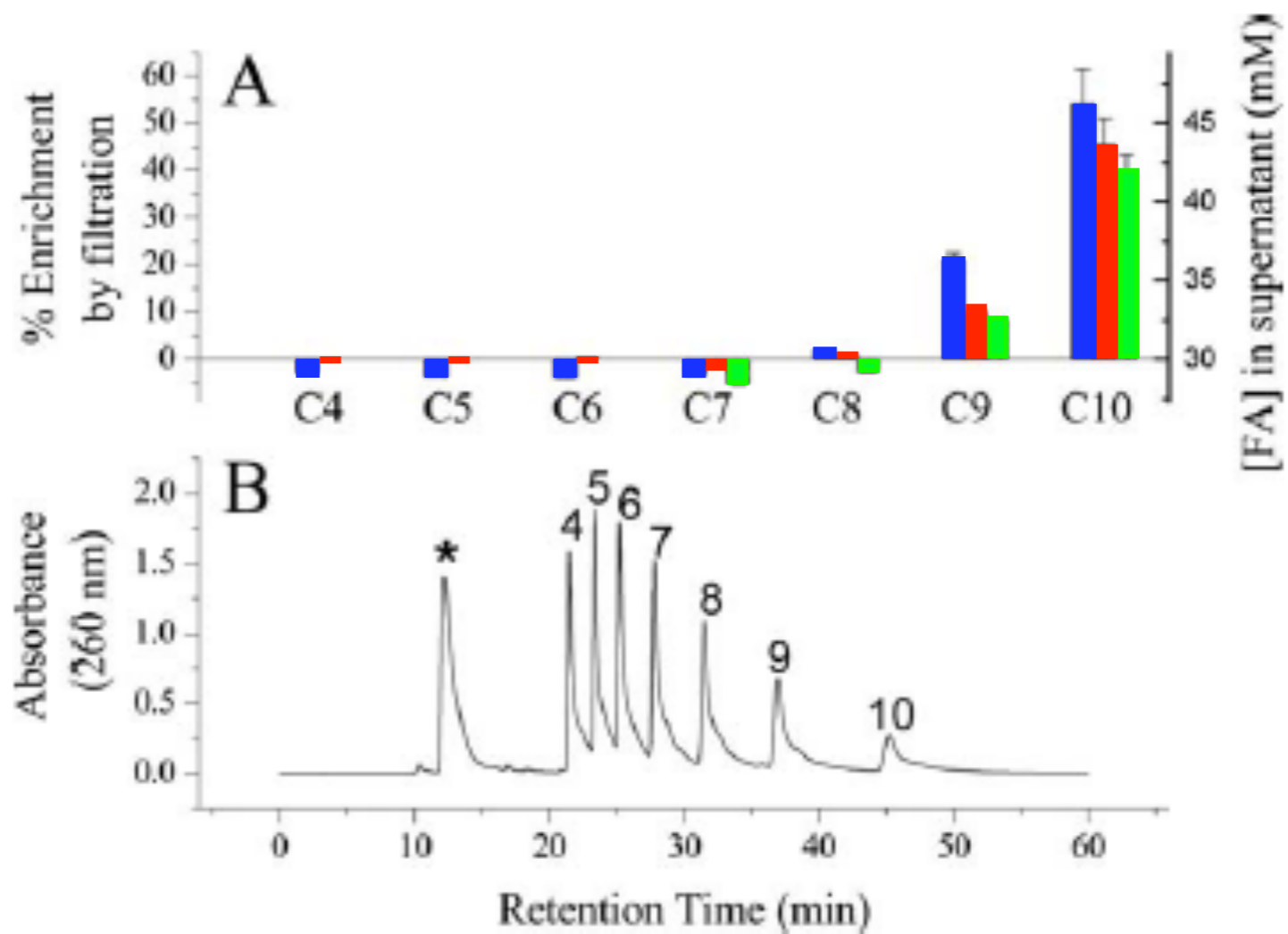


*Cape et al., 2011 Chem Sci*





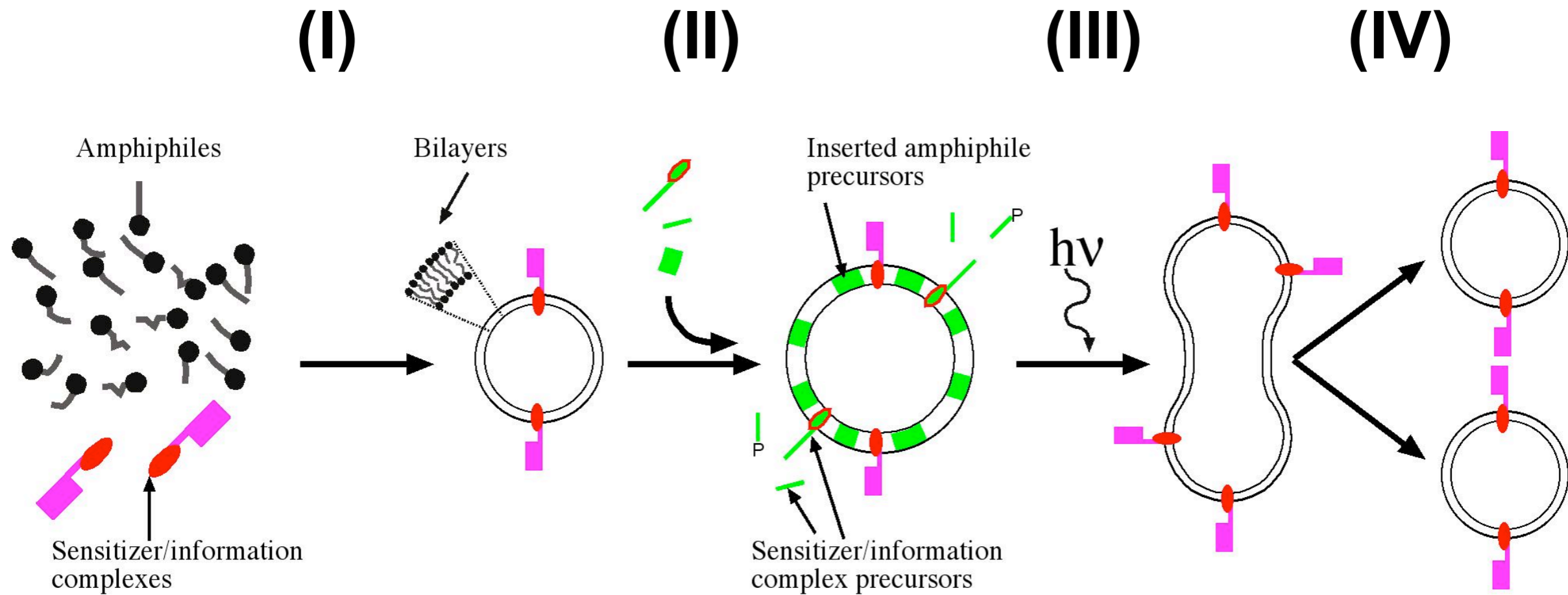
- Mixture containing C7 to C10
- Mixture containing C5 to C10
- Mixture containing C4 to C10



Cape et al., 2011 Chem Sci



# Procell Assembly: a bottom-up systemic approach



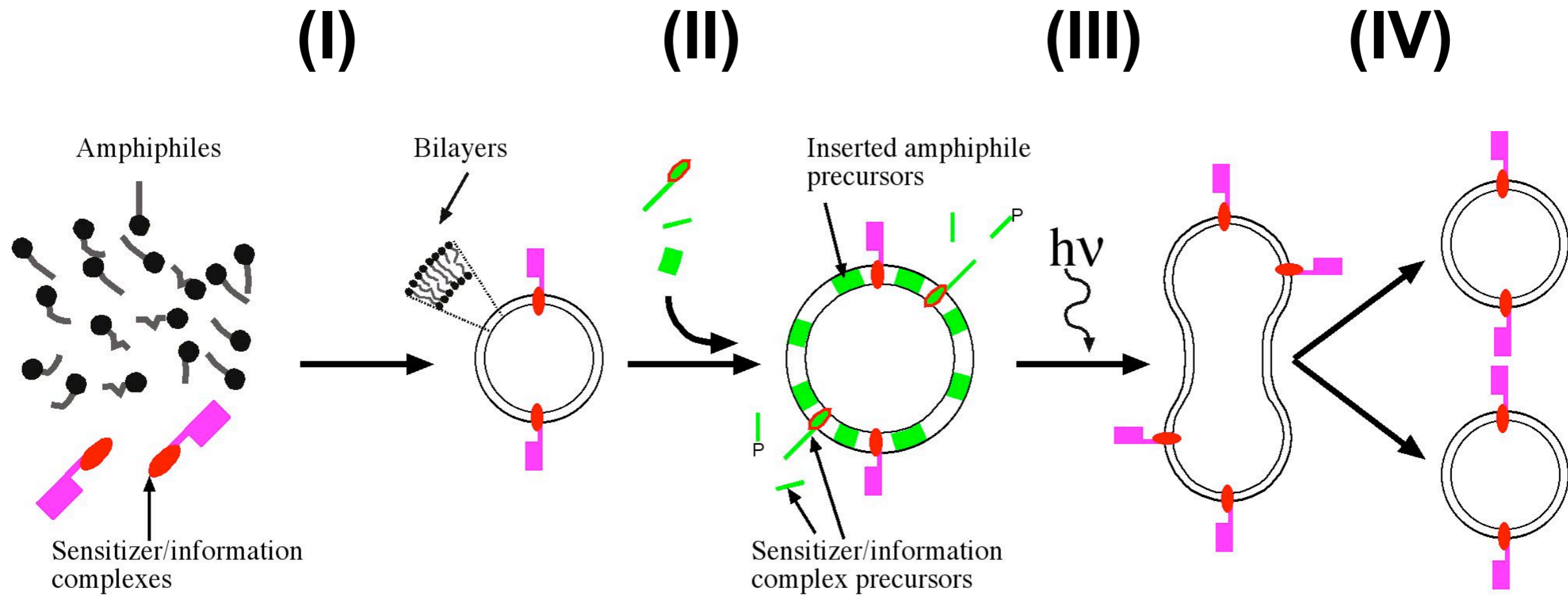
## Traits specific to this design

- Container is the amphiphile structure itself





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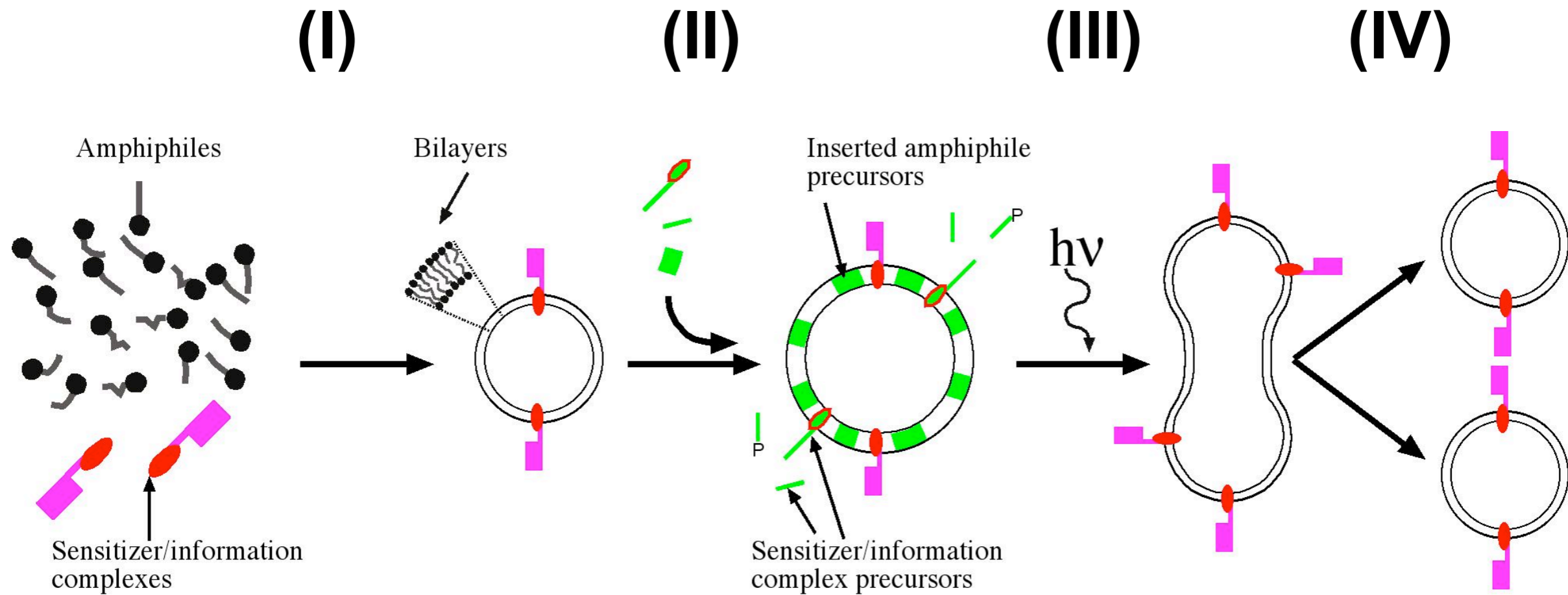


## Traits specific to this design

- Container is the amphiphile structure itself
- Total self-assembly of initial protocell



# Procell Assembly: a bottom-up systemic approach

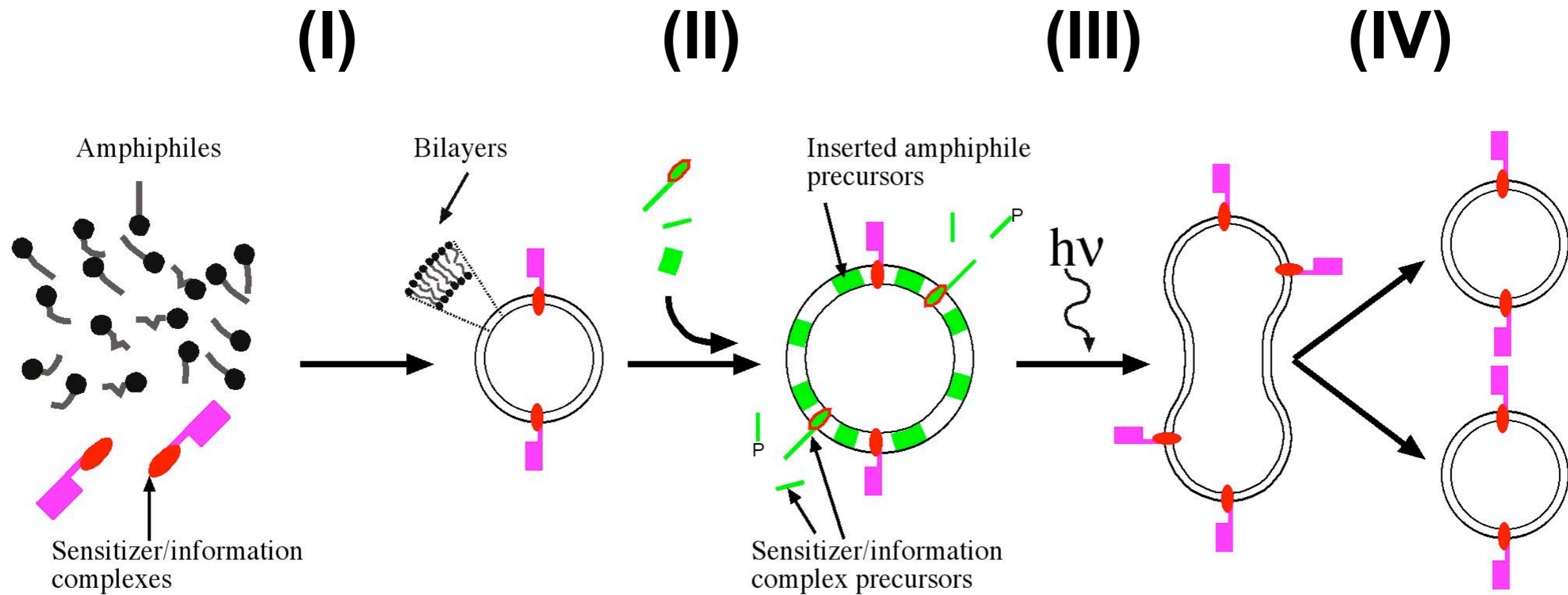


## Traits specific to this design

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- Direct access to “nutrients”



# Protocell Assembly: a bottom-up systemic approach

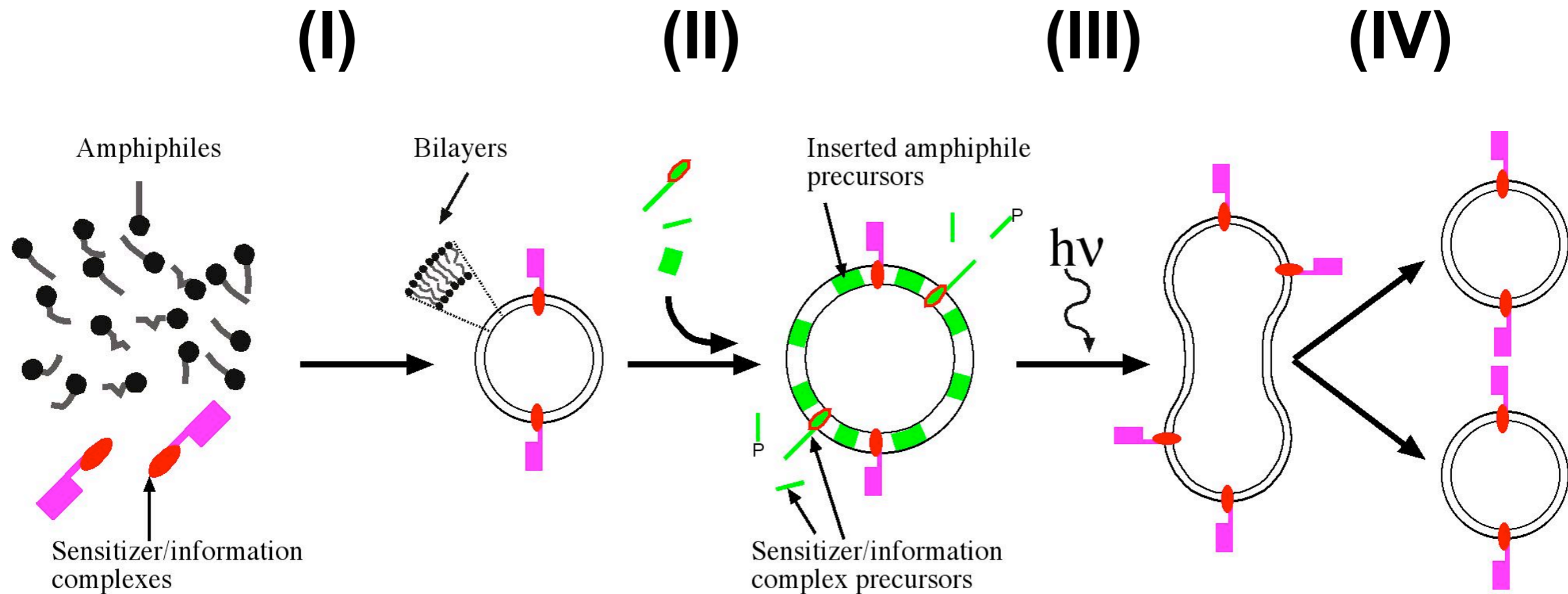


## Traits specific to this design

- Container is the amphiphile structure itself
- Total self-assembly of initial protocell
- Direct access to “nutrients
- Coupling of catalytic reaction to uptake of primary energy (light)



# Protocell Assembly: a bottom-up systemic approach



## Traits specific to this design

- Container is the amphiphile structure itself
- Total self-assembly of initial protocell
- Direct access to “nutrients”
- Coupling of catalytic reaction to uptake of primary energy (light)
- Direct control of the catalysis by the “information”



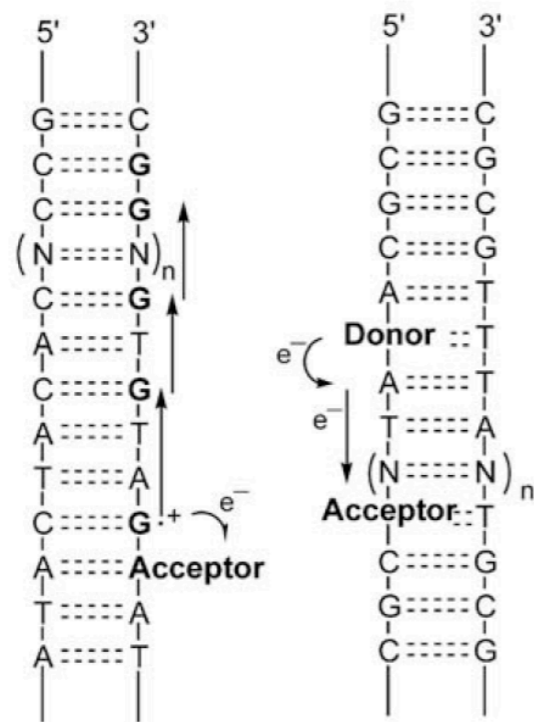
# Possible information control of metabolism



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## DNA as conducting polymer

Oxidative Hole Transfer    Reductive Excess Electron Transfer



**Fig. 1** Oxidative transfer of a radical cation through DNA and reductive transfer of an excess electron through DNA

*Behrens, C. et al 2004 Top. Curr. Chem*

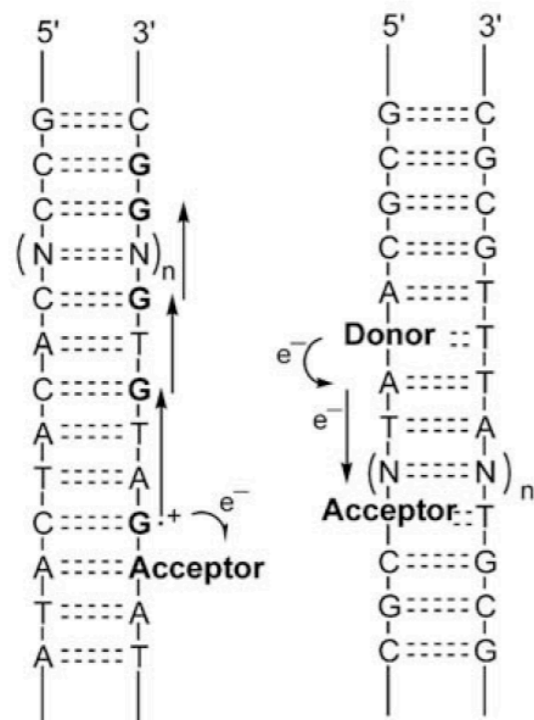
In Principle DNA could be an actuator for Redox or photochemical reaction even in small double stranded oligomeric systems



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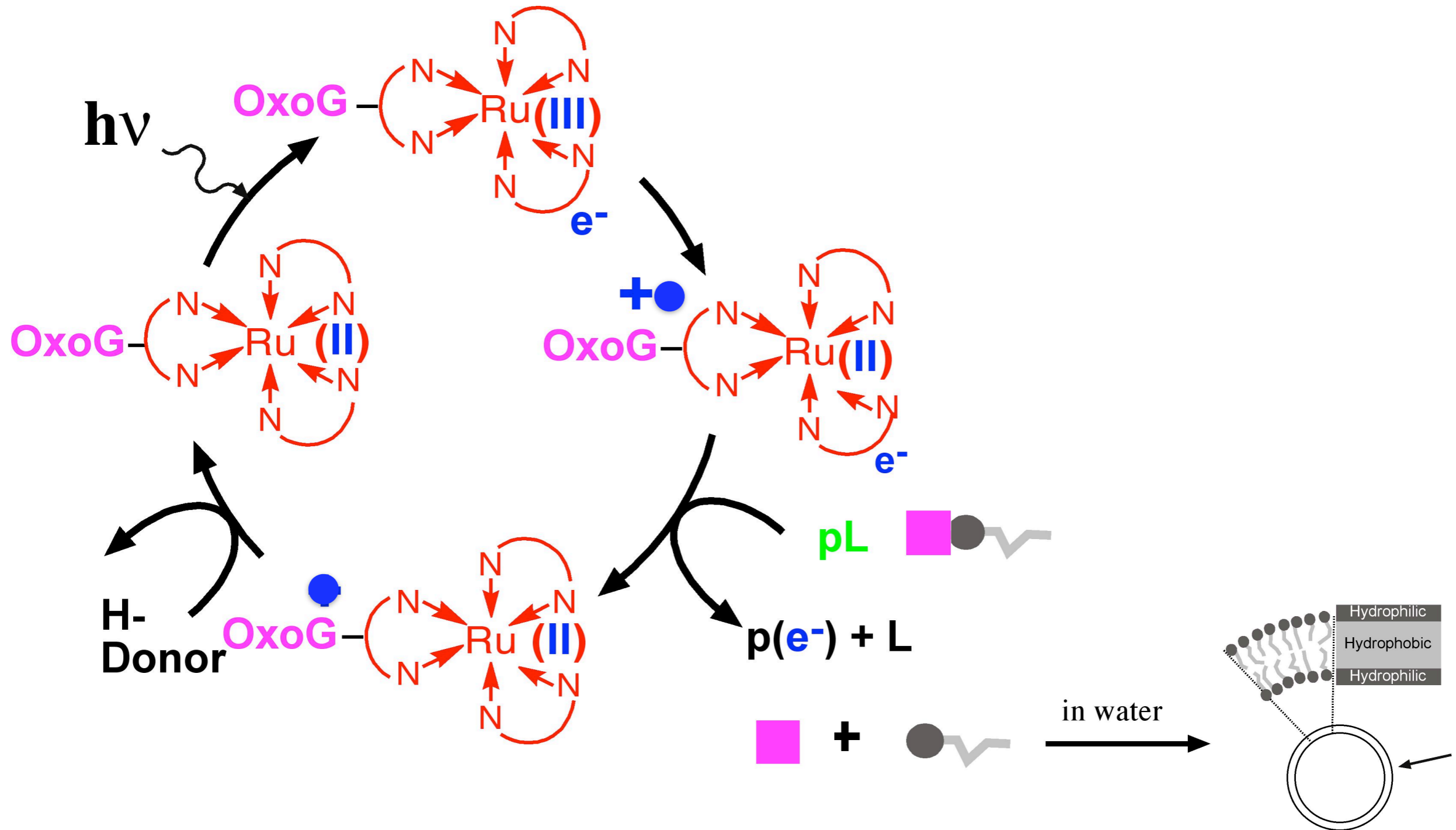
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two problems

- degradation of DNA
- electron relay must be recycled

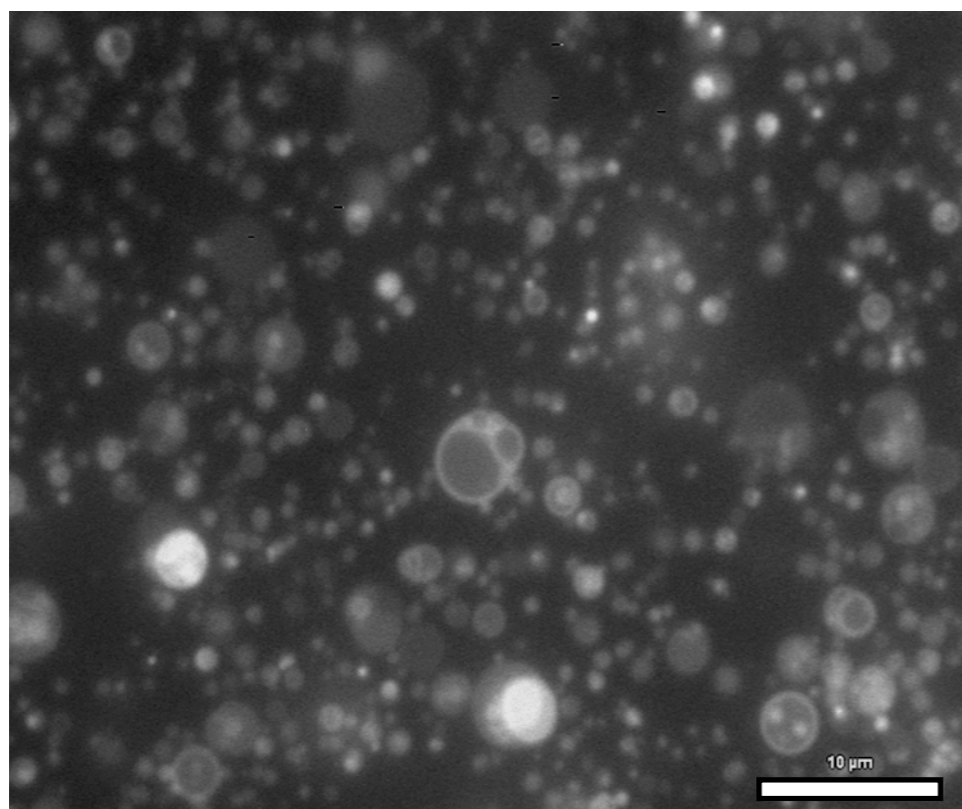
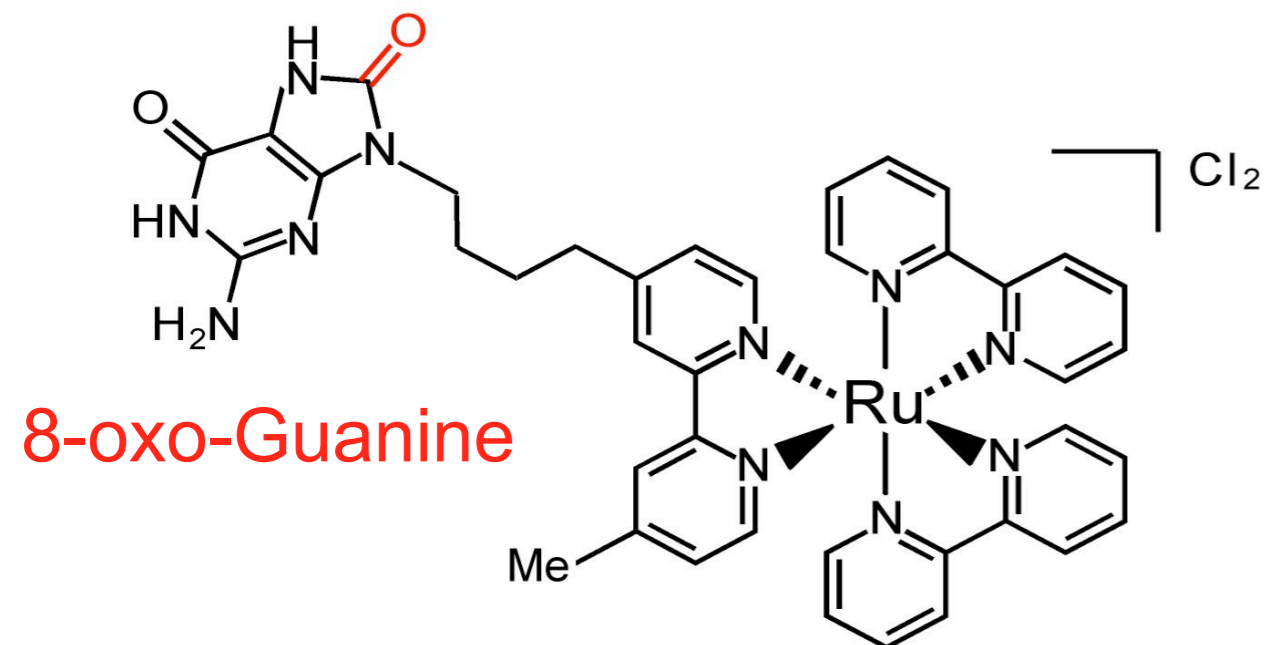


# Conclusion: Procell Assembly: "Info"-Metabolism-Container Interconnection





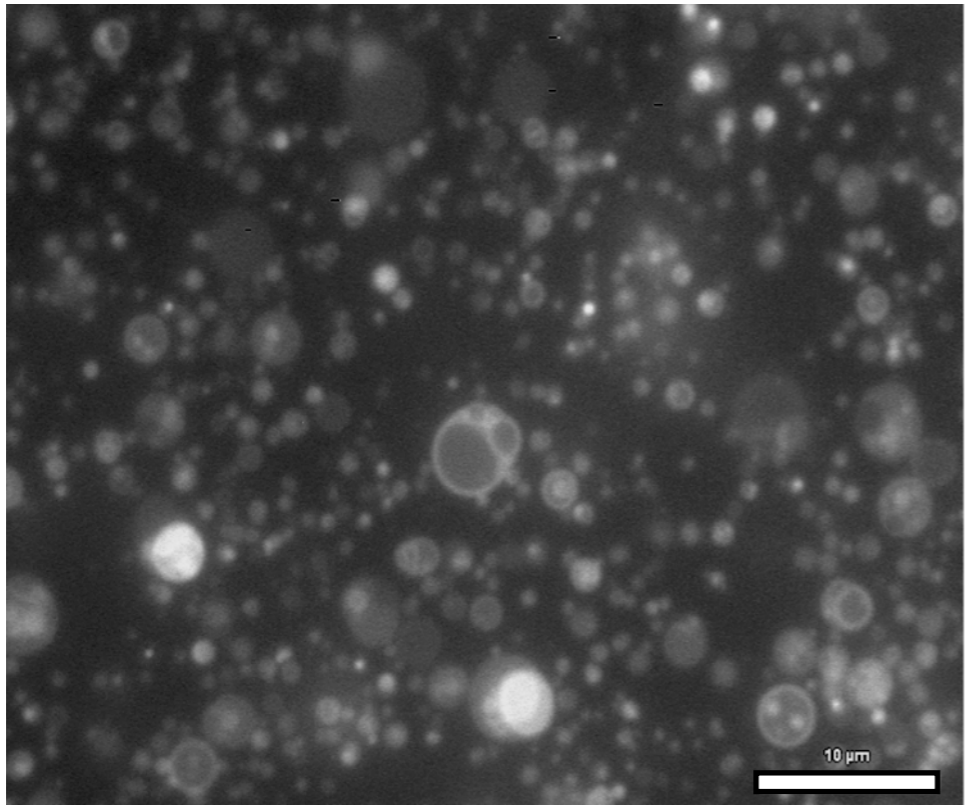
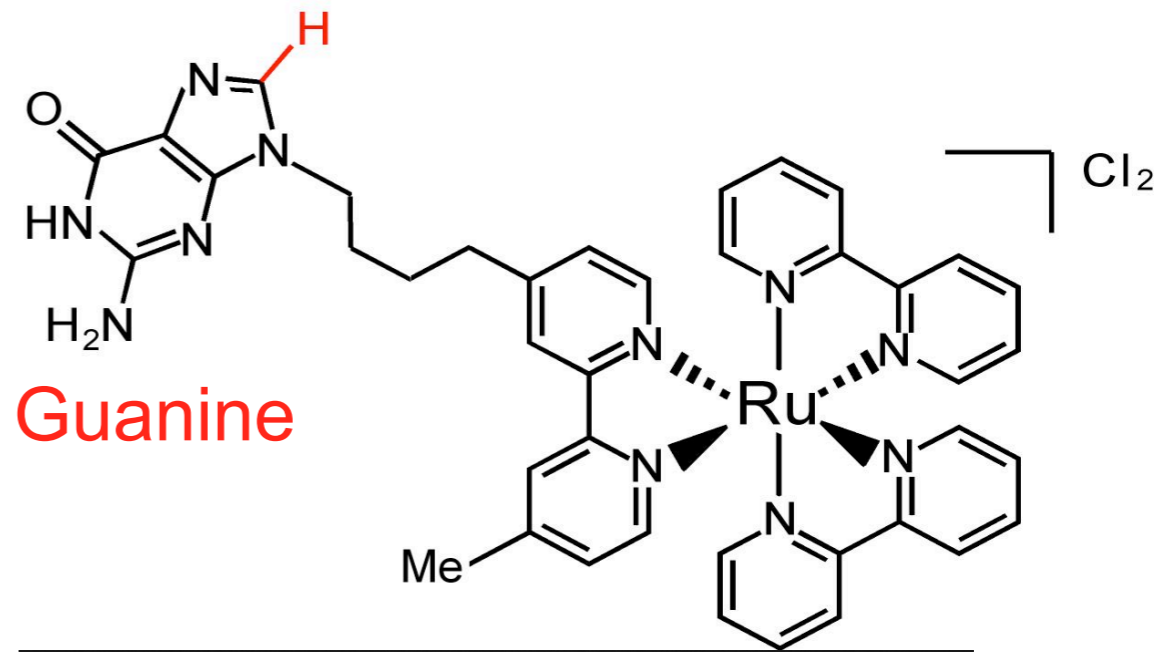
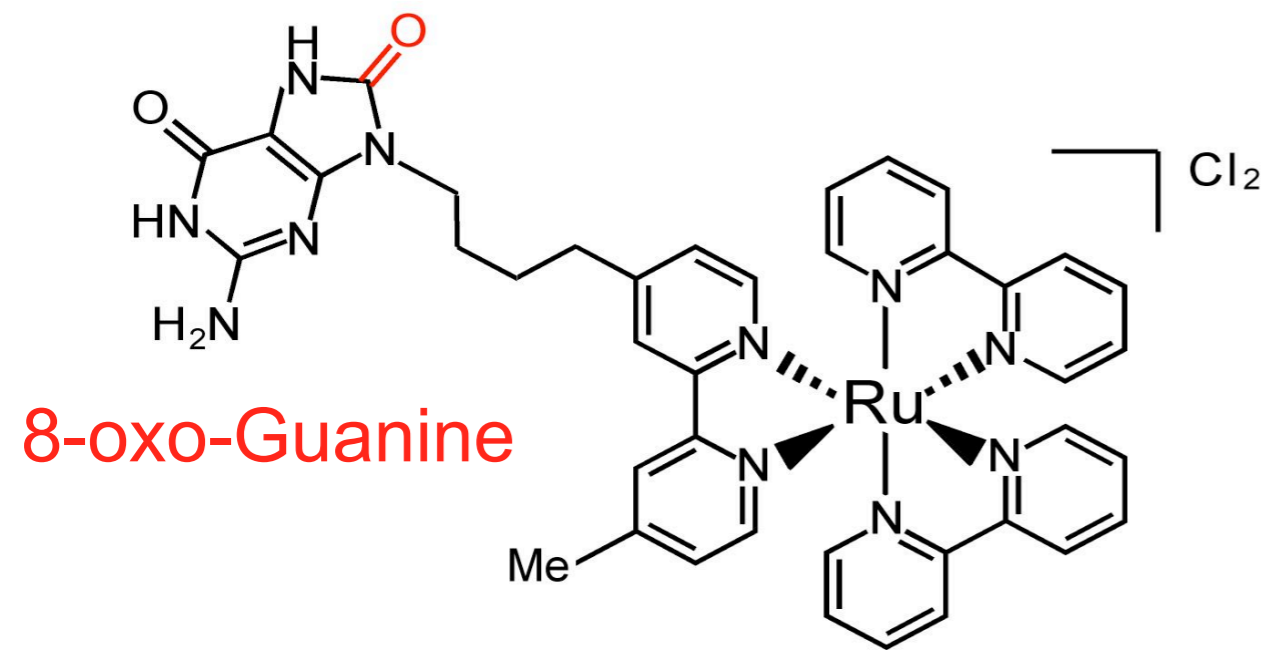
# Impact of the “information” molecule



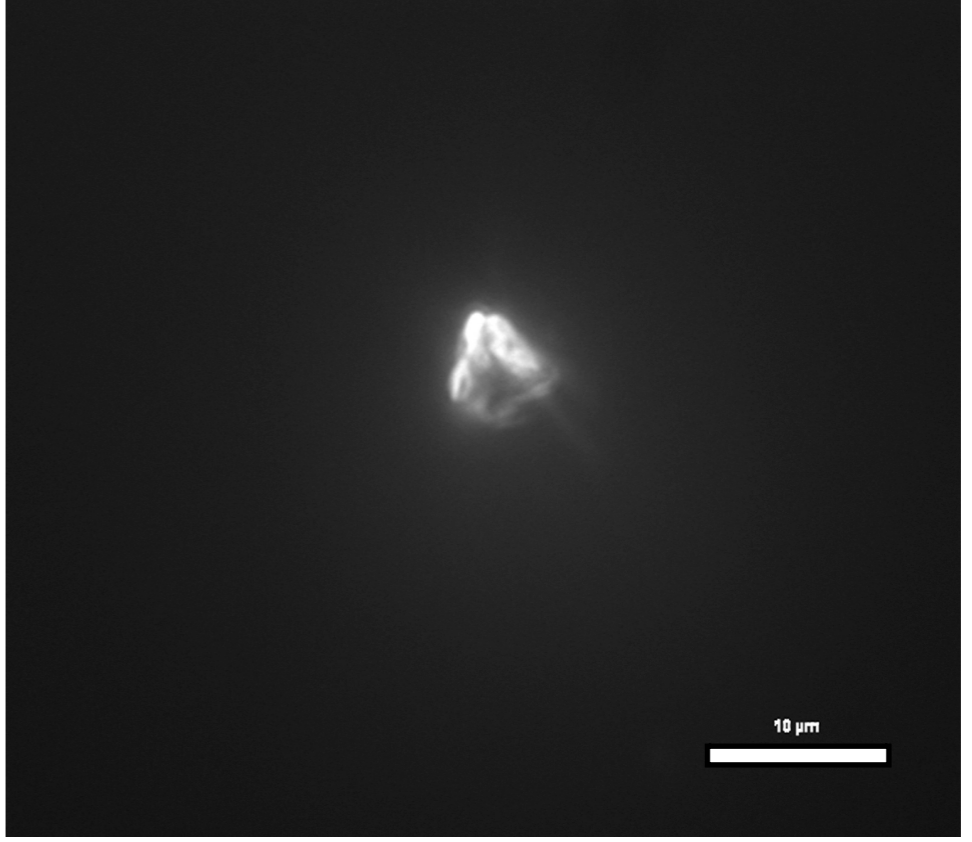
Membranous structures (100%)



# Impact of the "information" molecule



Membranous structures (100%)



Oil and crystals only (<10%)



# Influence of the reaction set-up on precursor conversion (initial rates)



Reaction: 0.1 mM catalyst, 5 mM of precursor, 15.75 mM H-source. With vesicles: 10 mM Decanoic acid

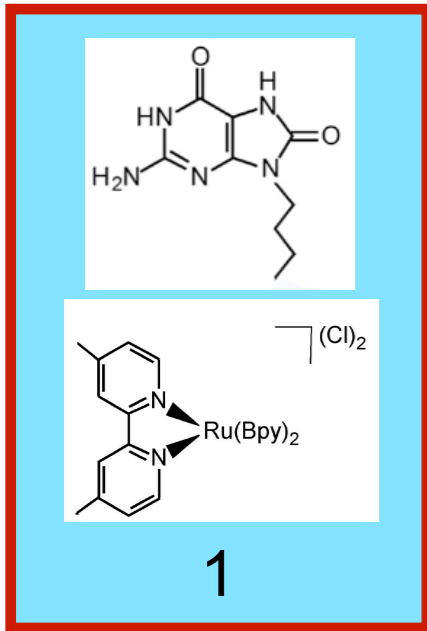
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*Maurer et al 2011 ChemPhysChem*

# Influence of the reaction set-up on precursor conversion (initial rates)

Intermolecular

aqueous



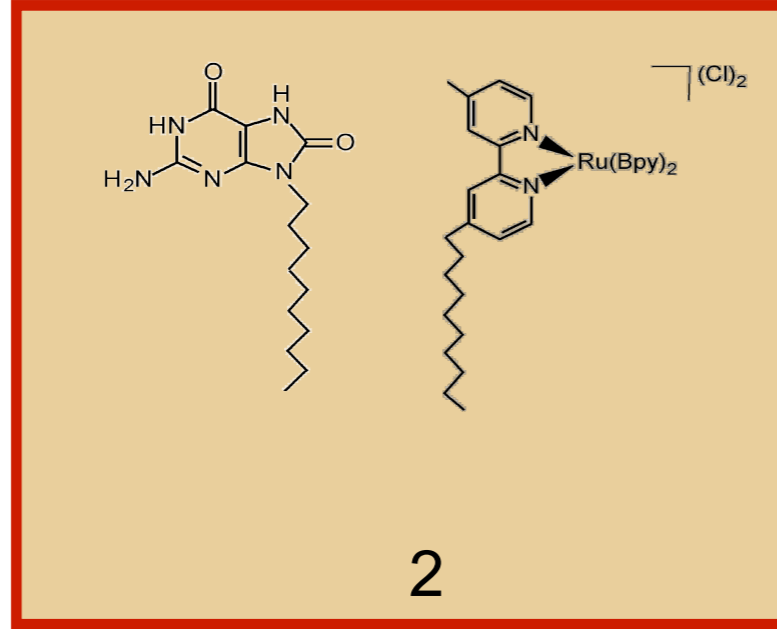
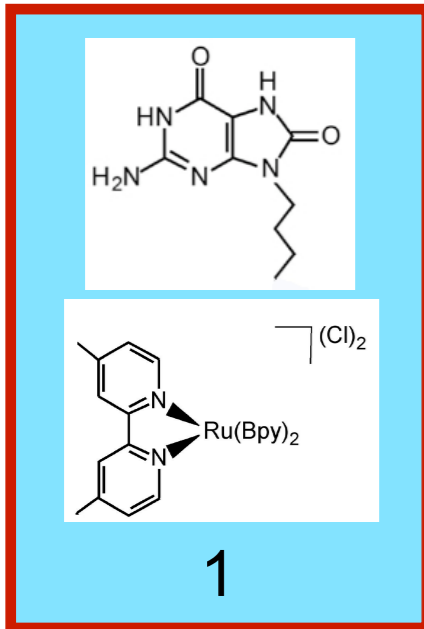
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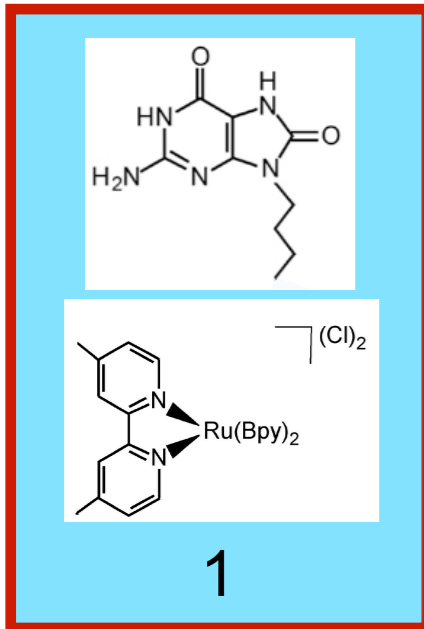
oil/bilayer linked



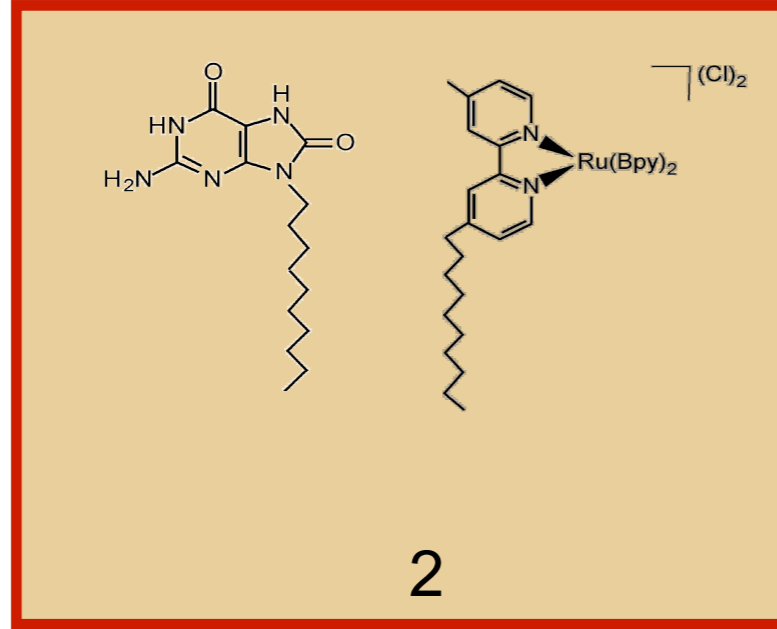
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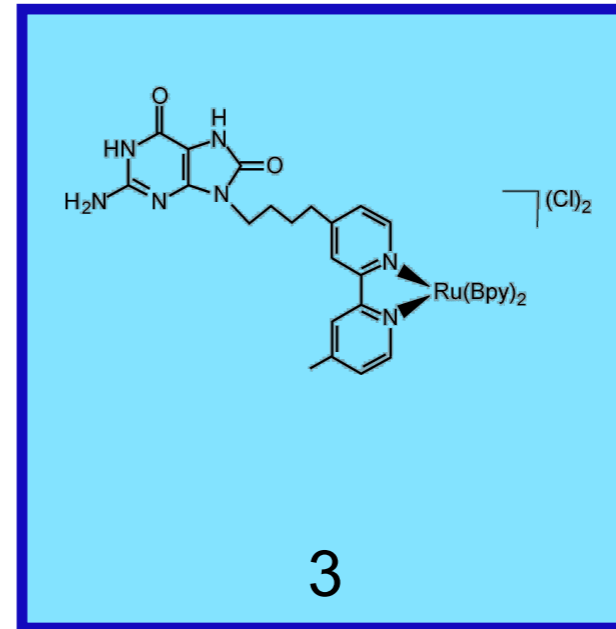
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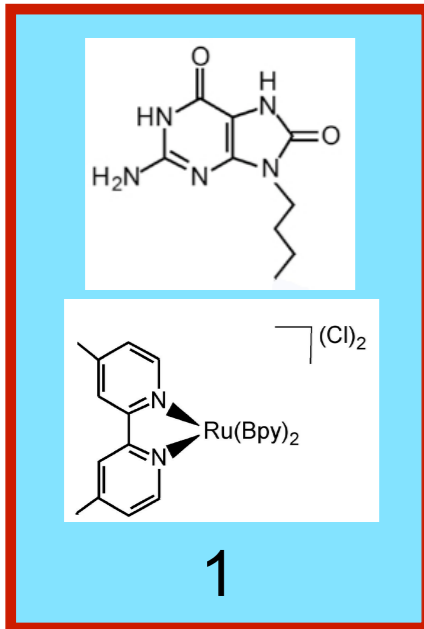
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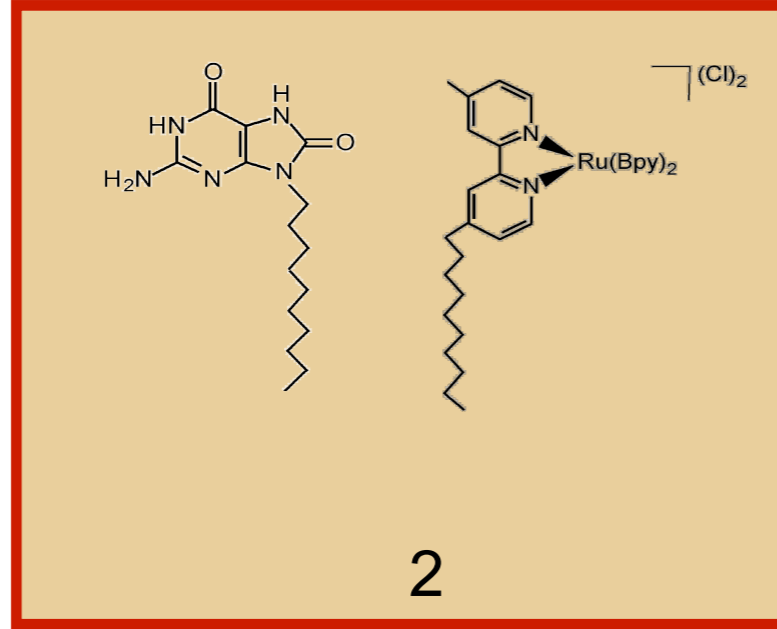
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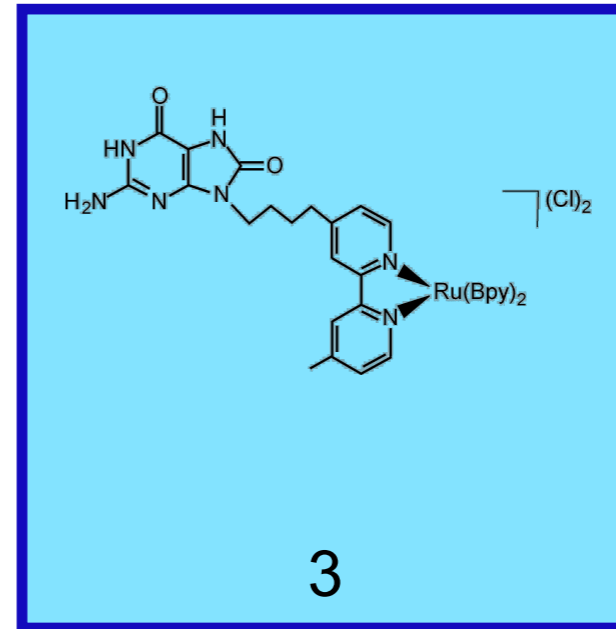
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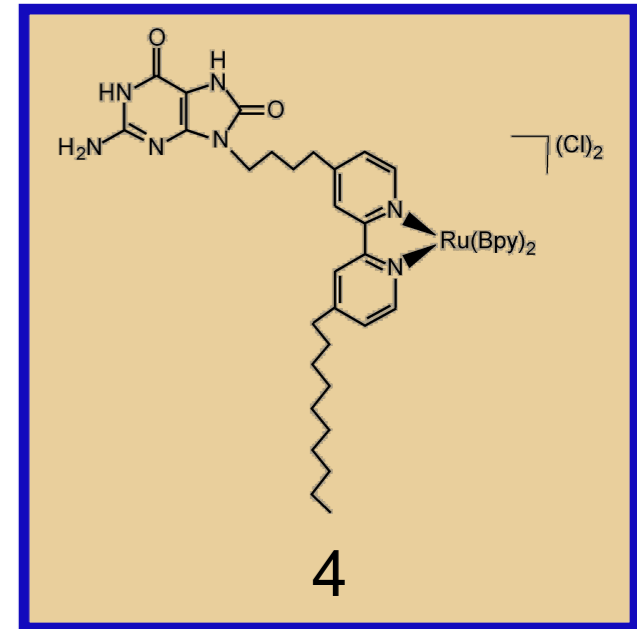
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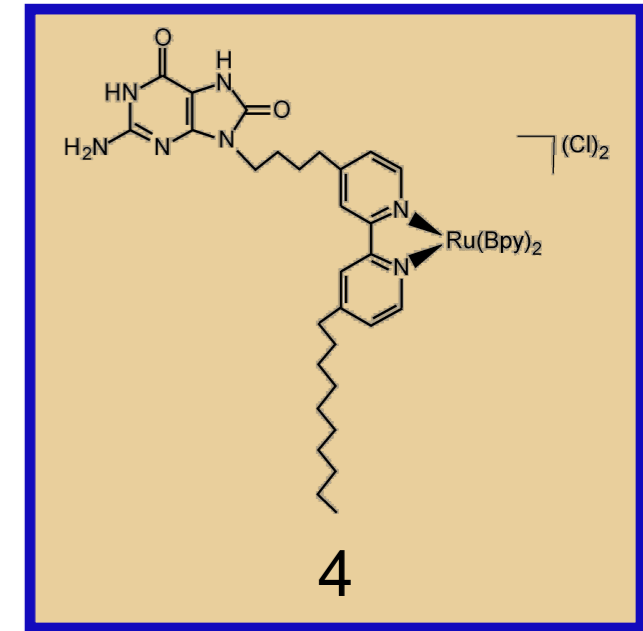
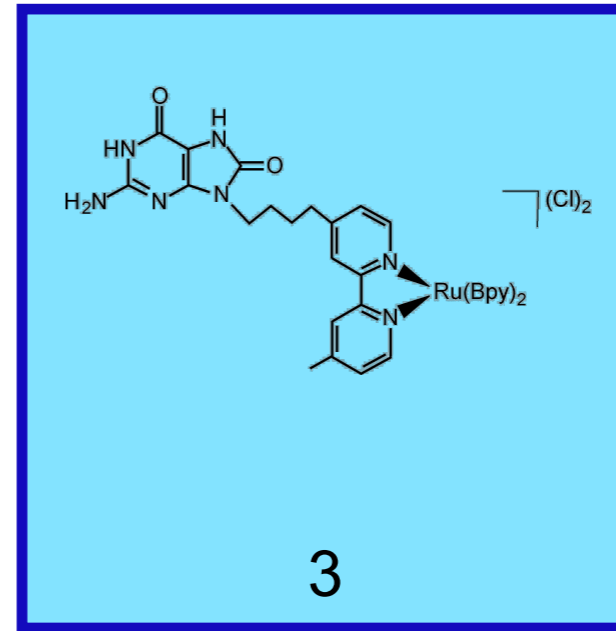
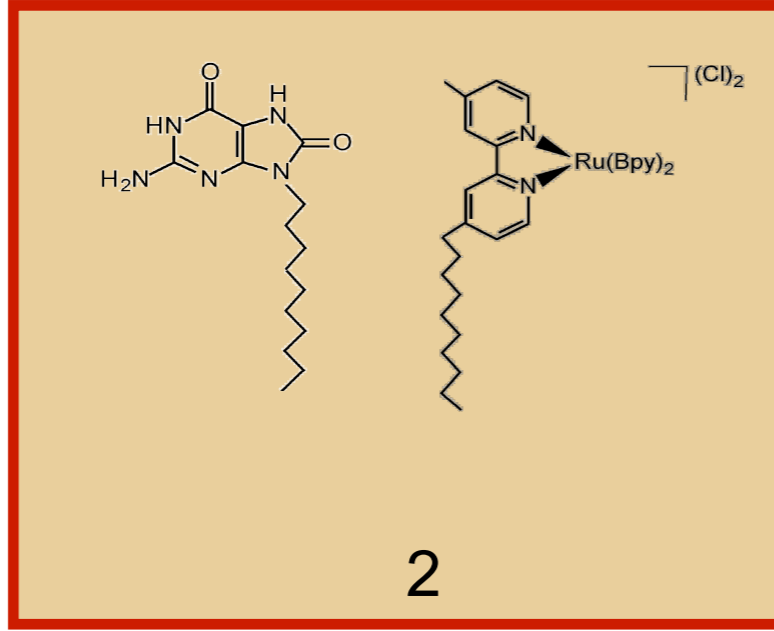
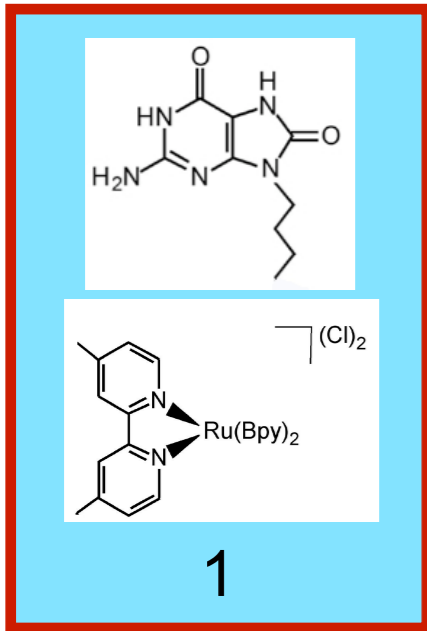
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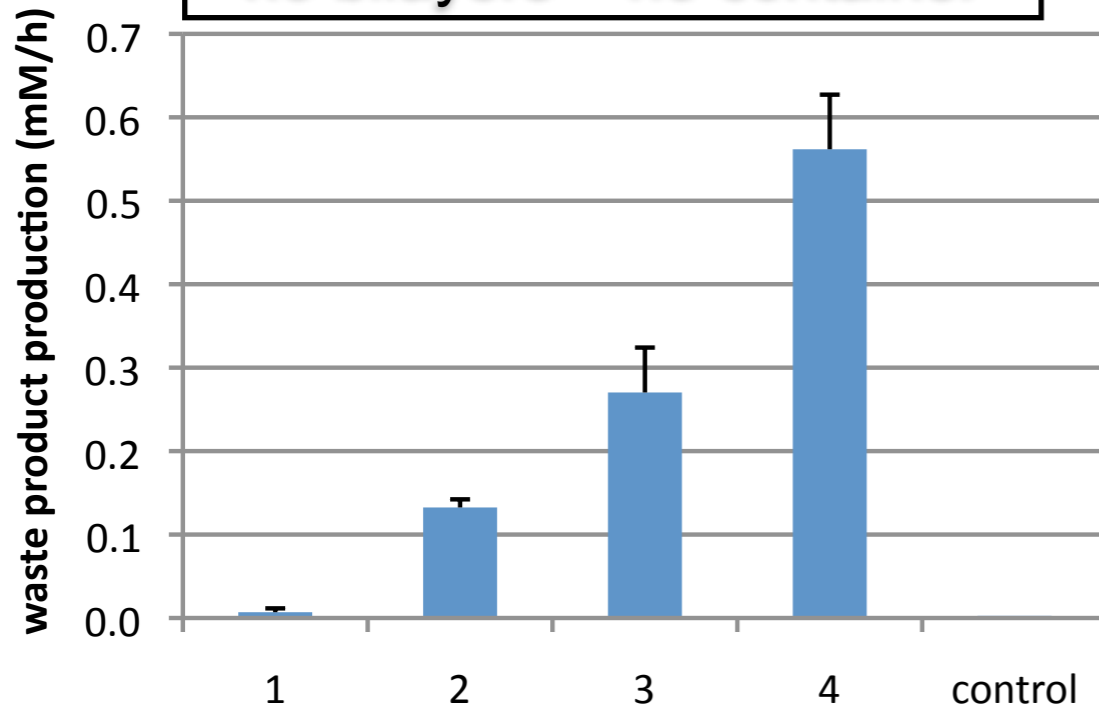
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no bilayers = no container



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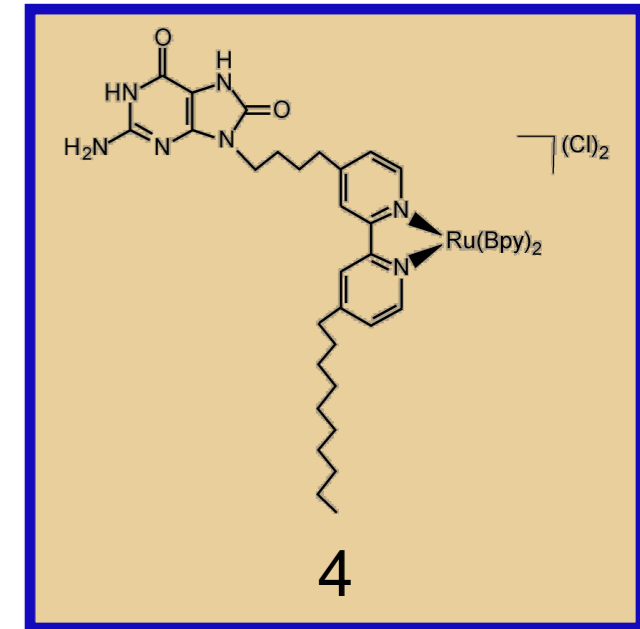
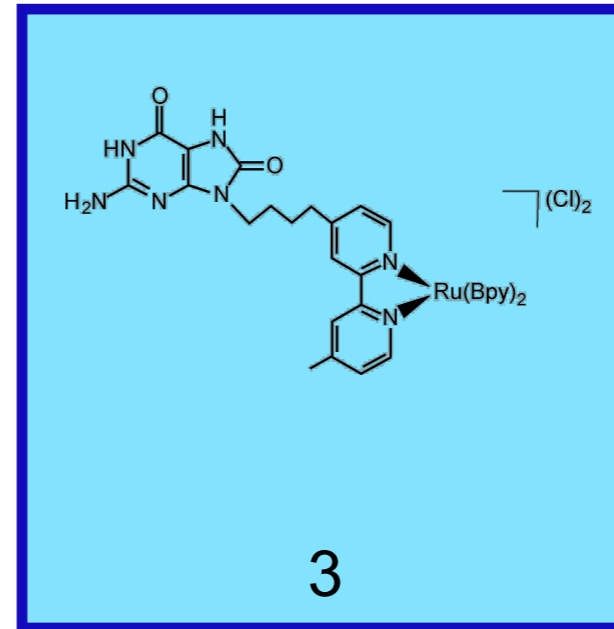
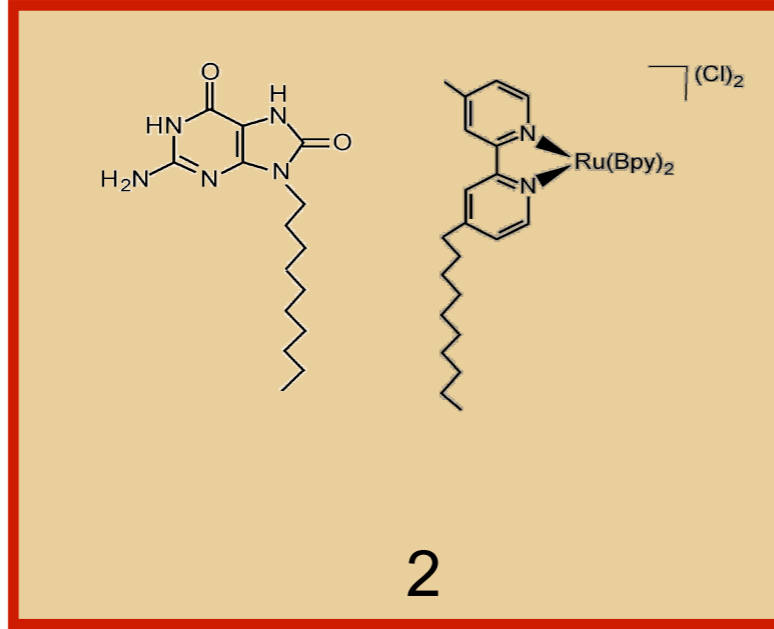
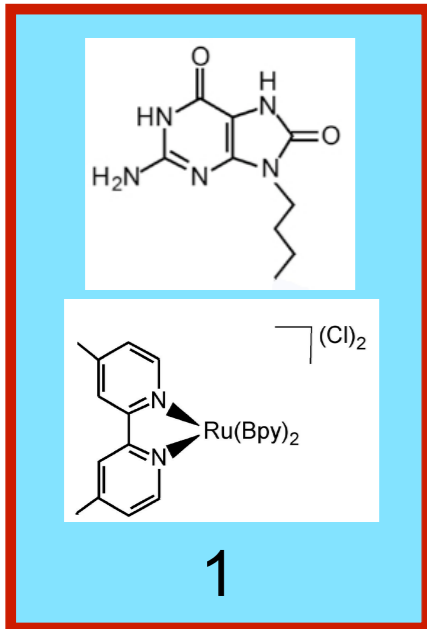
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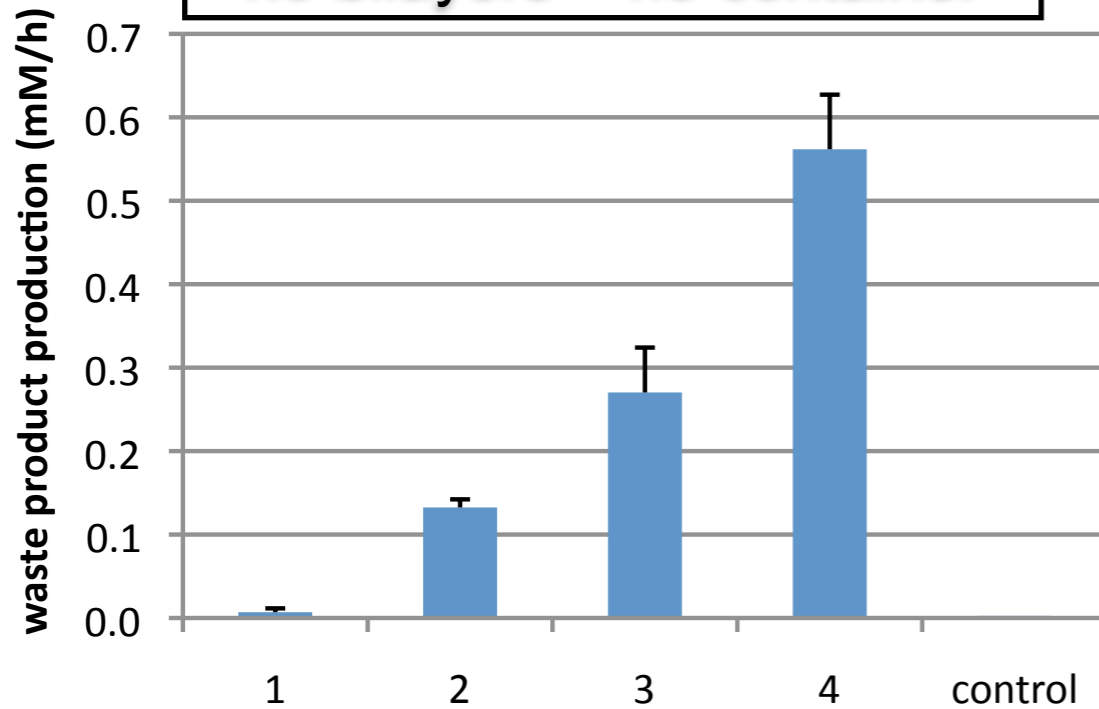
intramolecular

aqueous

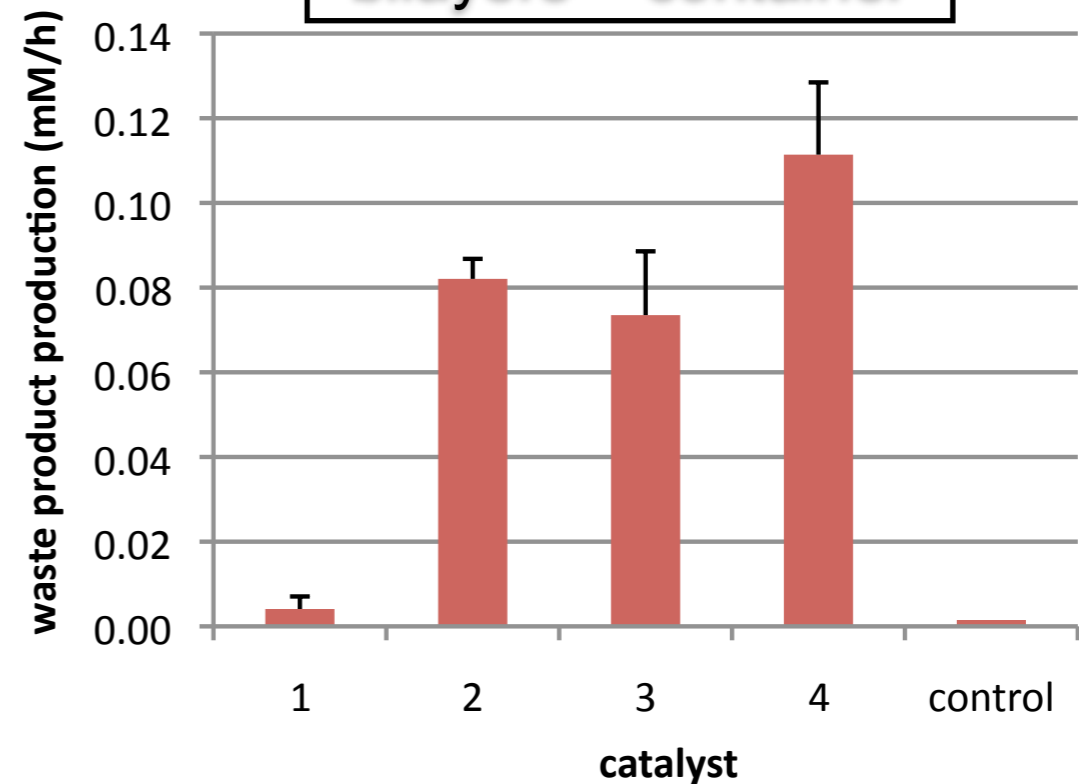
oil/bilayer linked



no bilayers = no container



bilayers = container



Reaction: 0.1 mM catalyst, 5 mM of precursor, 15.75 mM H-source. With vesicles: 10 mM Decanoic acid

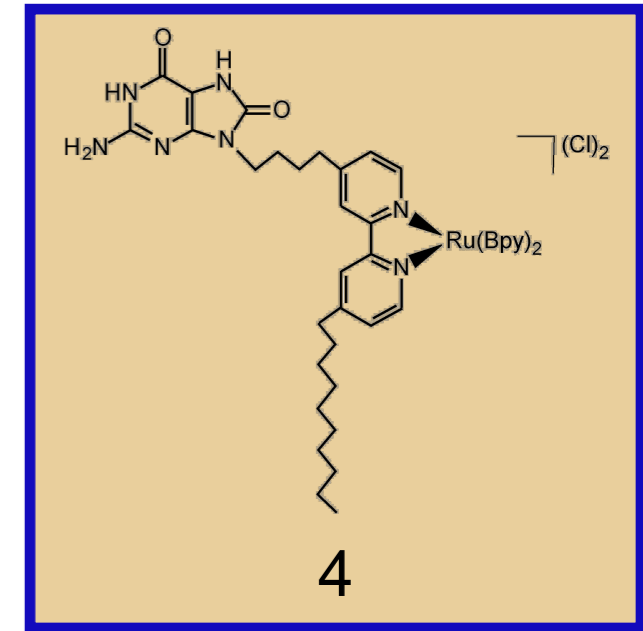
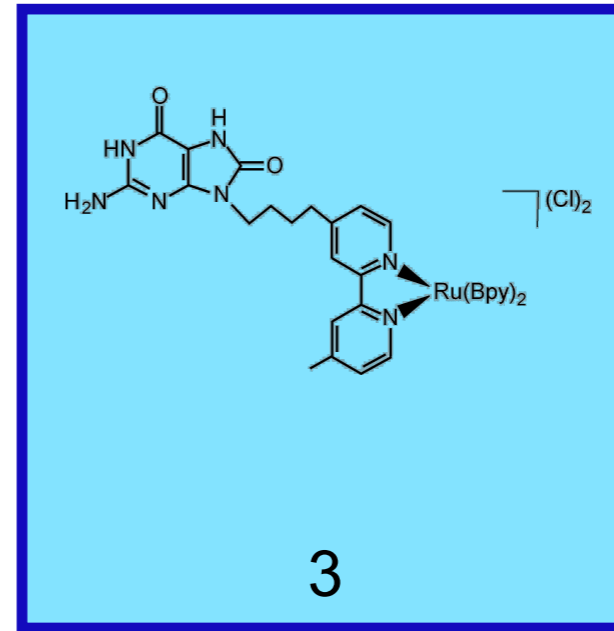
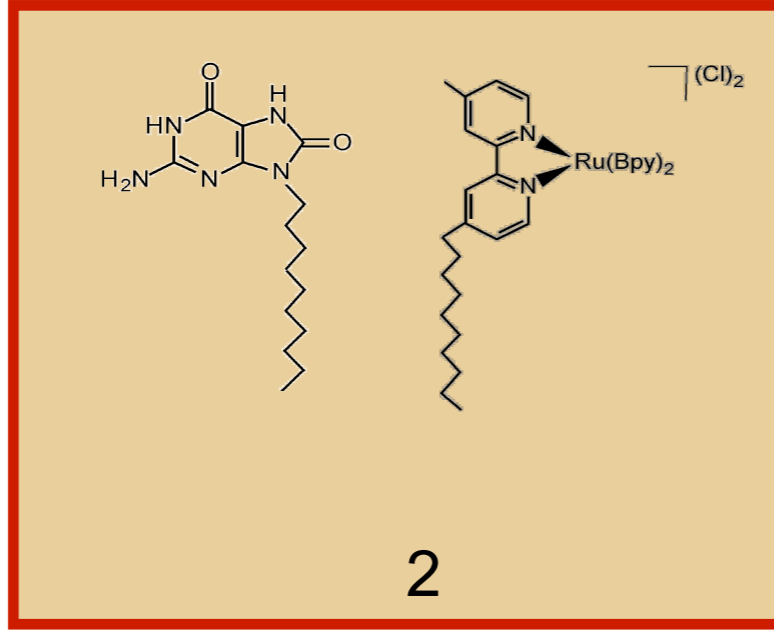
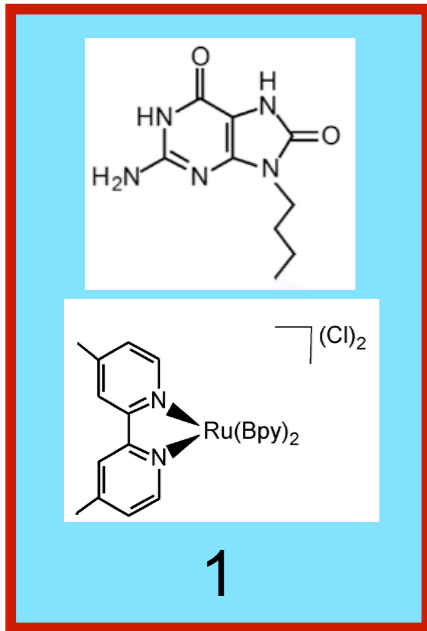
# Influence of the reaction set-up on precursor conversion (initial rates)

Intermolecular

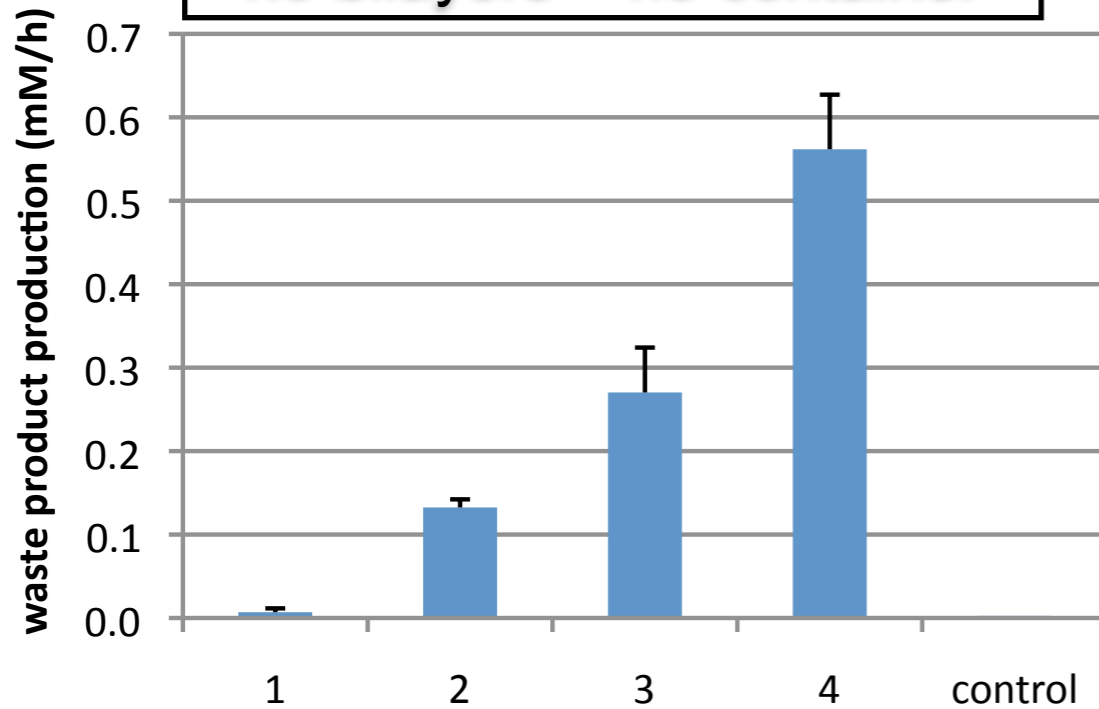
intramolecular

aqueous

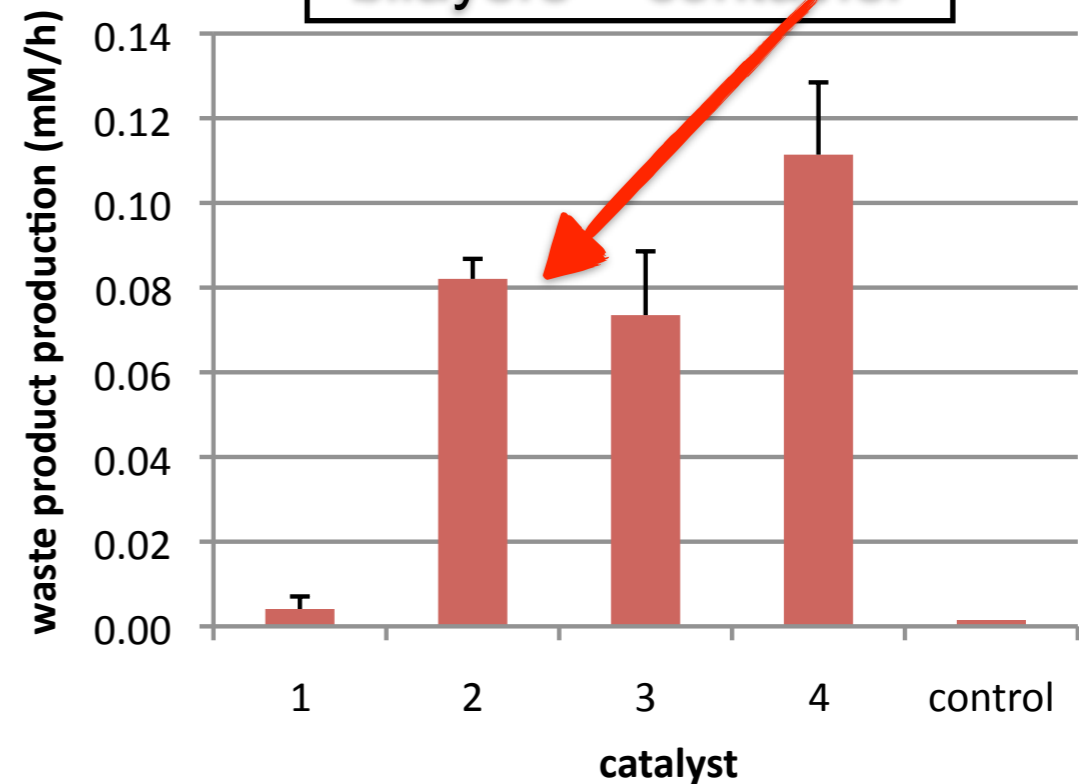
oil/bilayer linked



no bilayers = no container

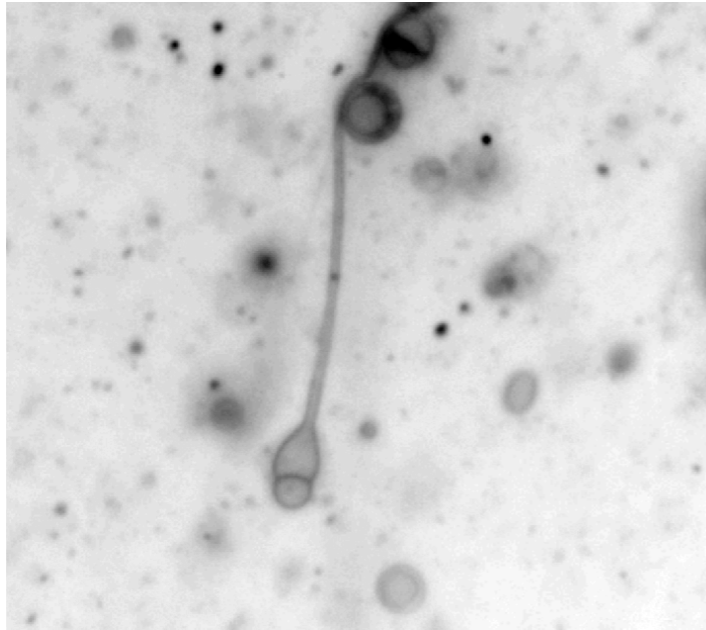


bilayers = container

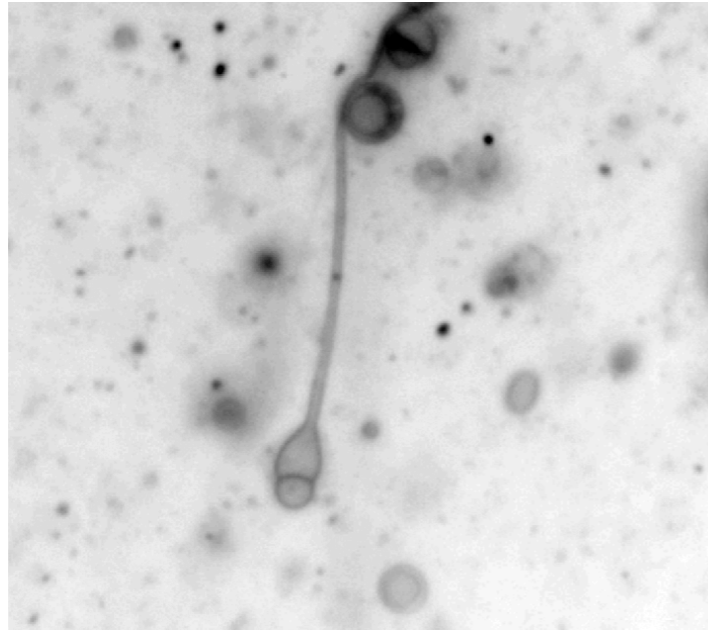


Reaction: 0.1 mM catalyst, 5 mM of precursor, 15.75 mM H-source. With vesicles: 10 mM Decanoic acid

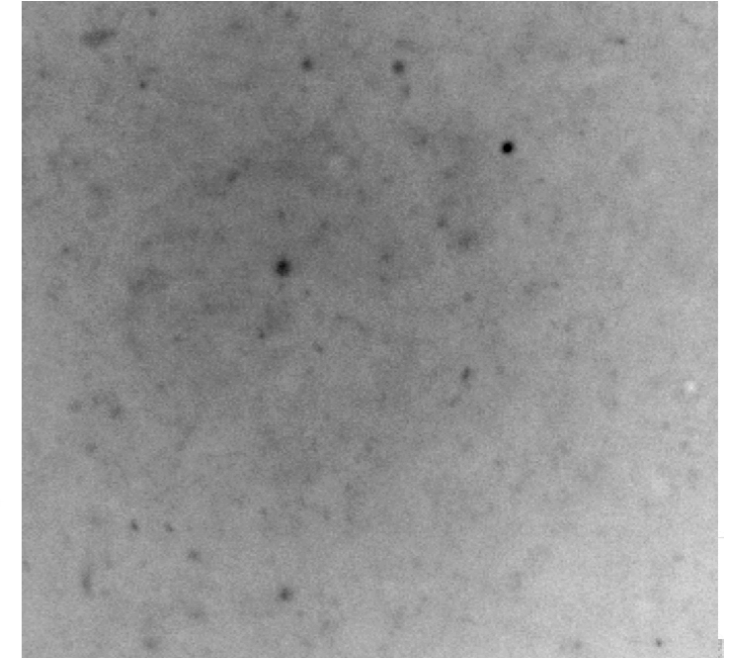
# Observation of a complete life cycle



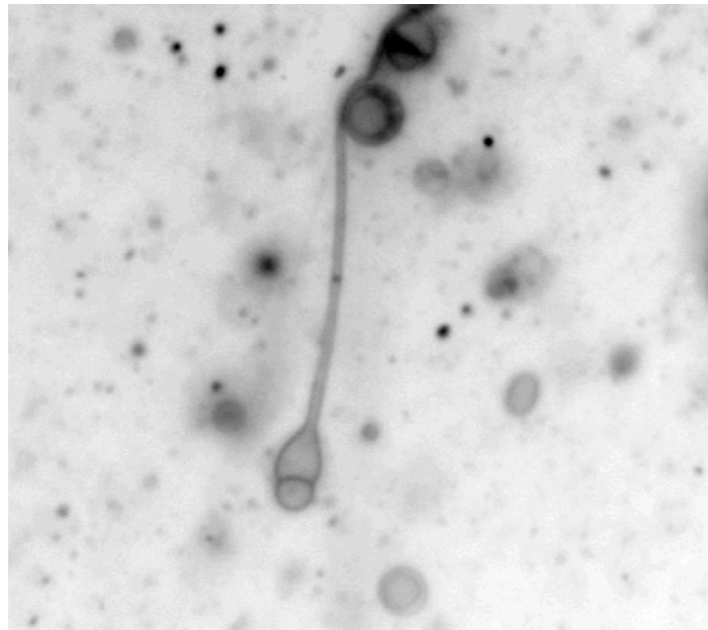
# Observation of a complete life cycle



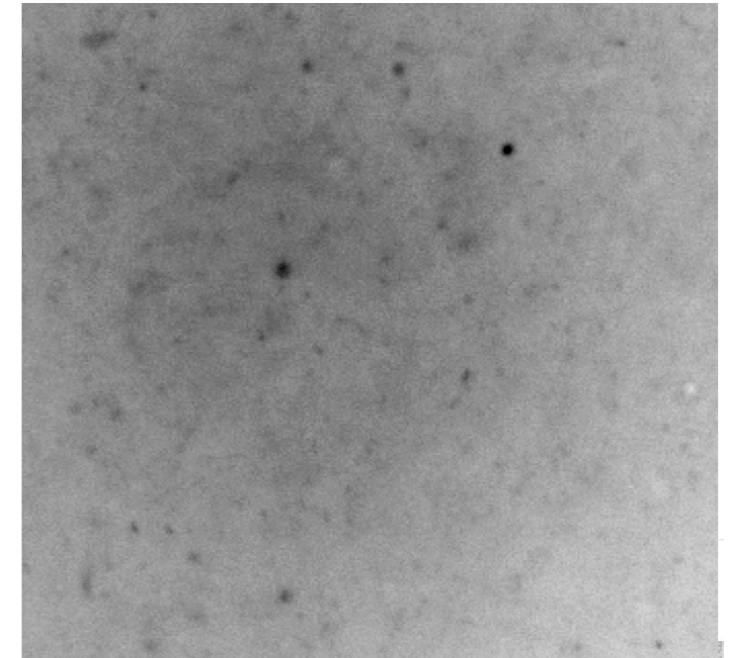
Homogenization



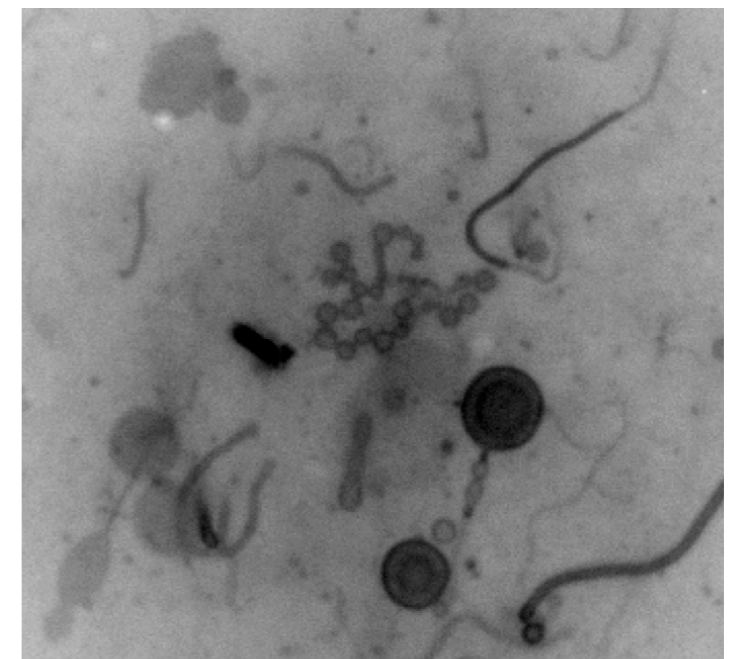
# Observation of a complete life cycle



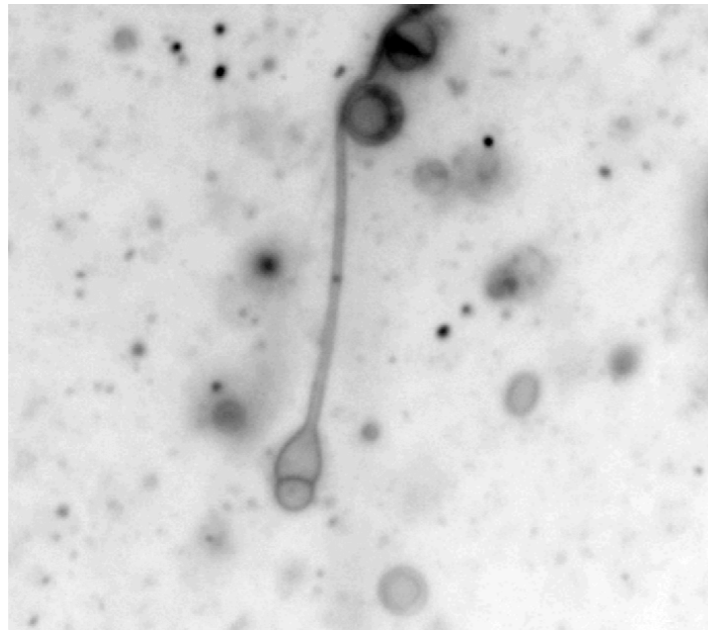
Homogenization



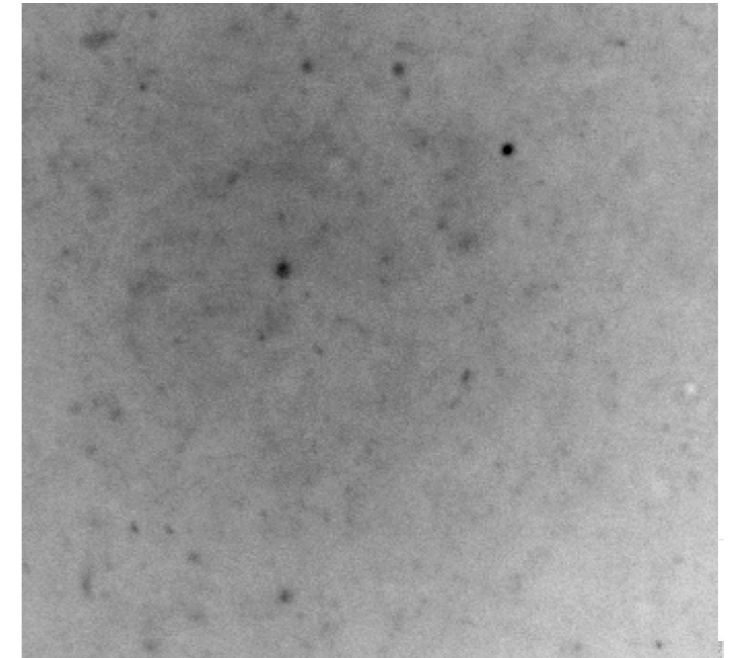
Nutrient uptake



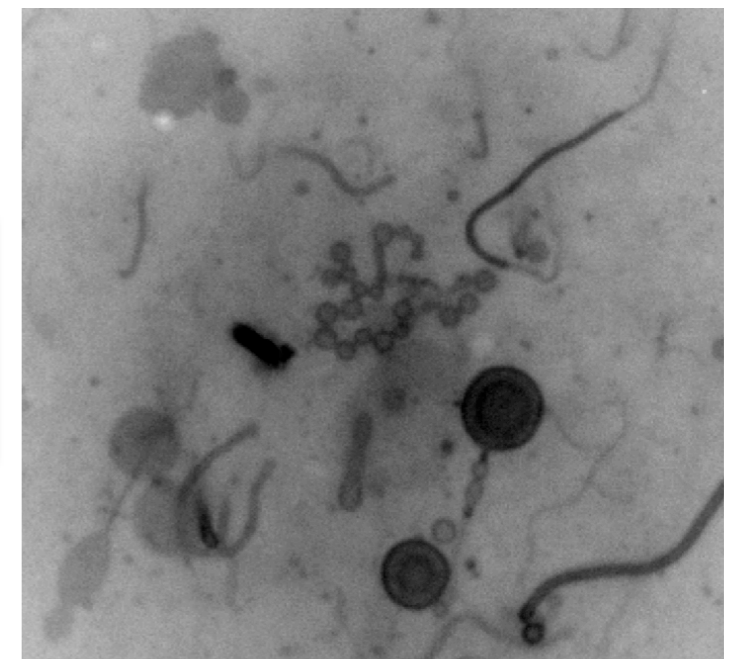
# Observation of a complete life cycle



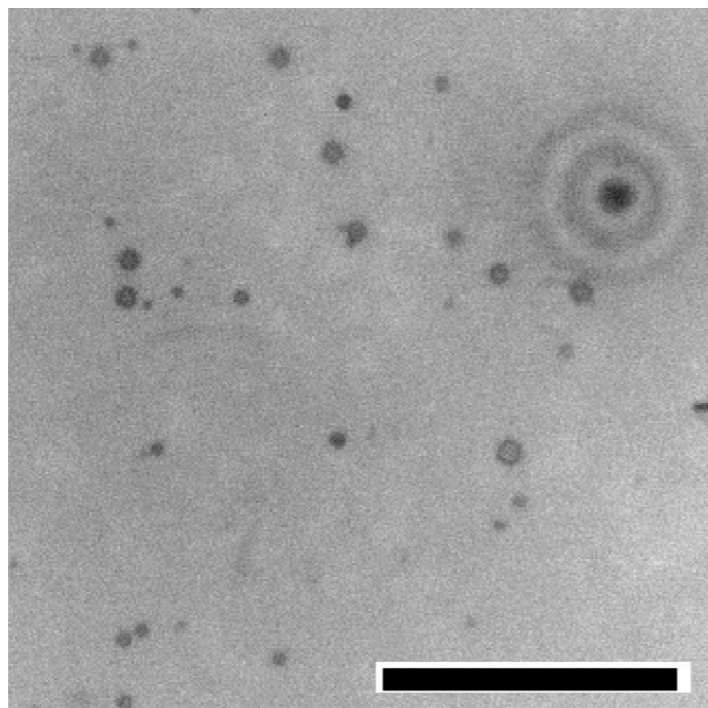
Homogenization



Nutrient uptake



Metabolic amphiphile production

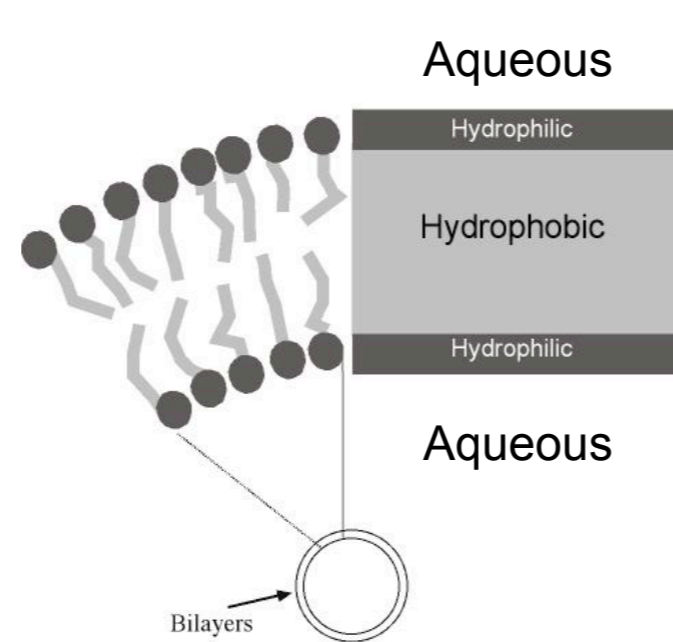


bar = 25  $\mu\text{m}$

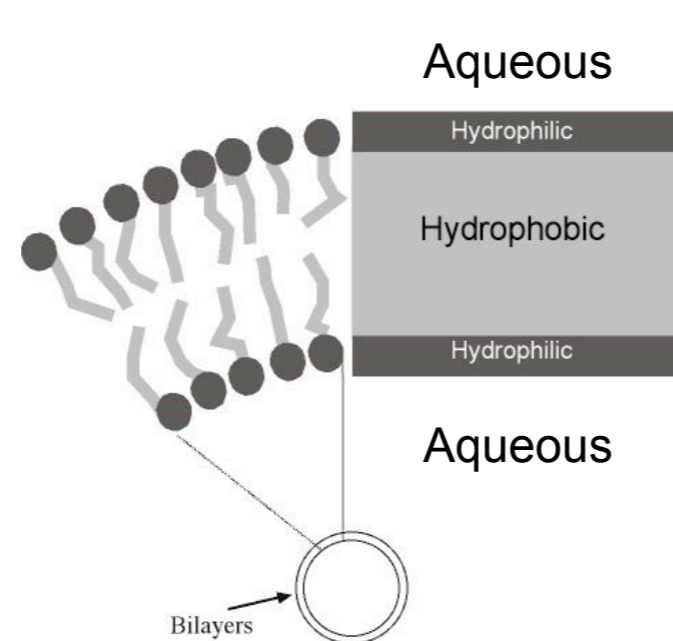
*Maurer & Albertsen*



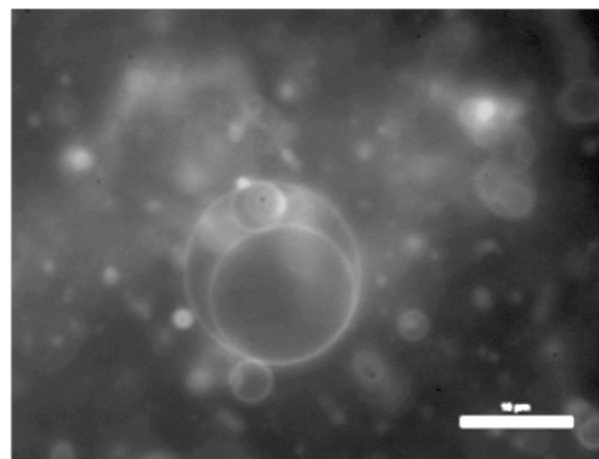
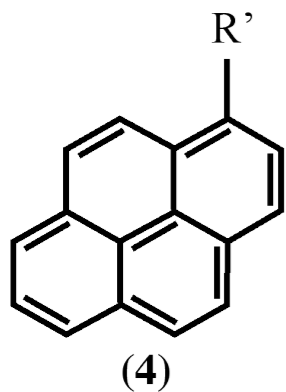
# Other possible functions of primitive membranes: Can primitive membranes promote reactions?



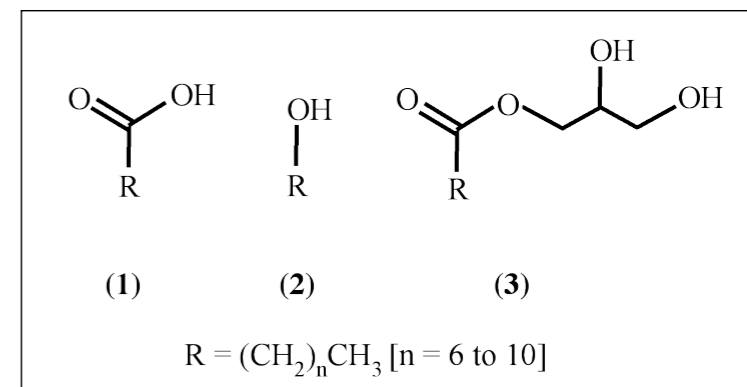
# Other possible functions of primitive membranes: Can primitive membranes promote reactions?



## Chemical autonomy by internalized production of chemicals using light harvesting systems



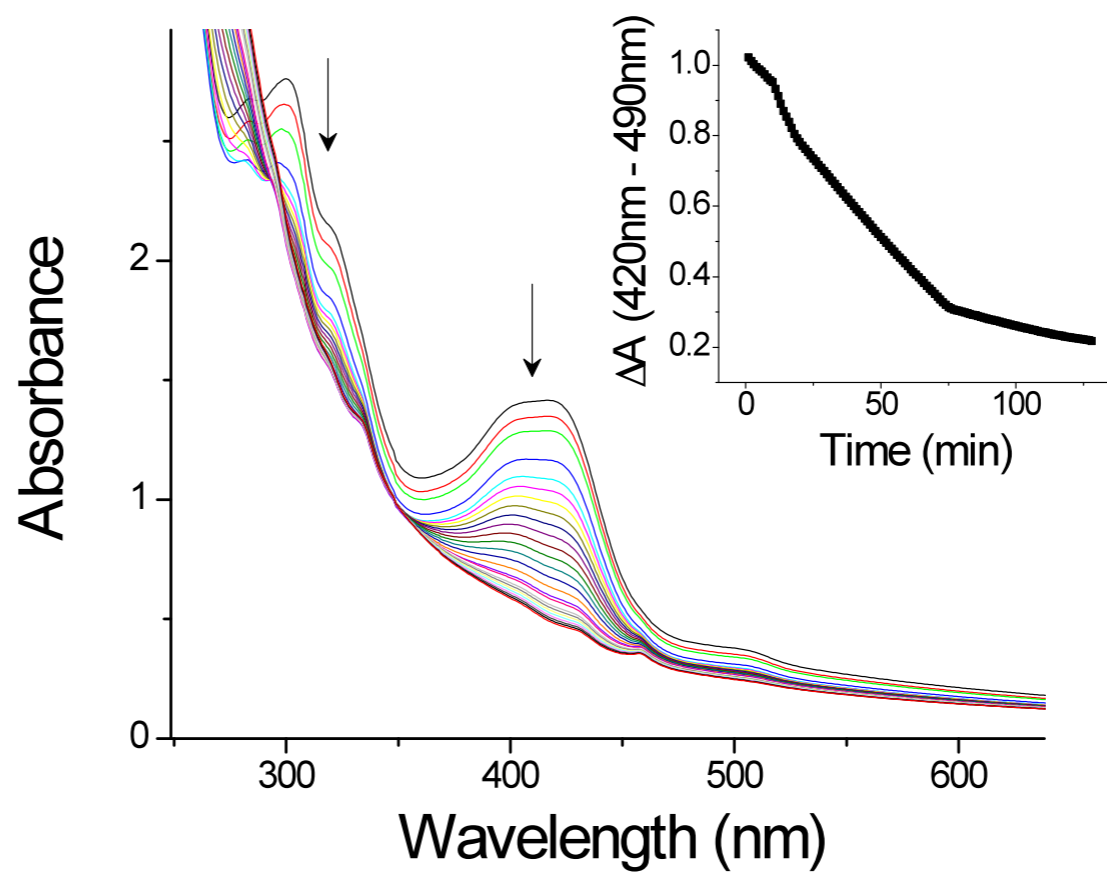
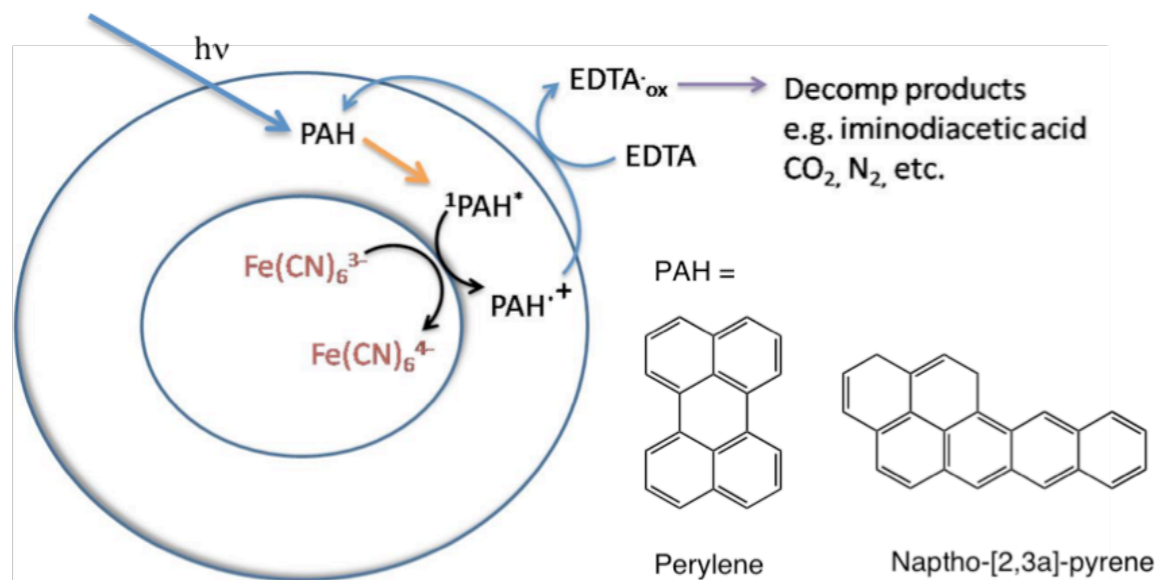
(4) Pyrene and its derivatives



Fatty (1) acids, (2) alcohols, (3) monoacylglycerol. Epifluorescence micrographs from suspensions of decanoic acid (1) with  $n$  equals 9. Bars = 10  $\mu\text{m}$ .







# Other Quasi-compartmentalization self-replication of early genetic/catalytic material

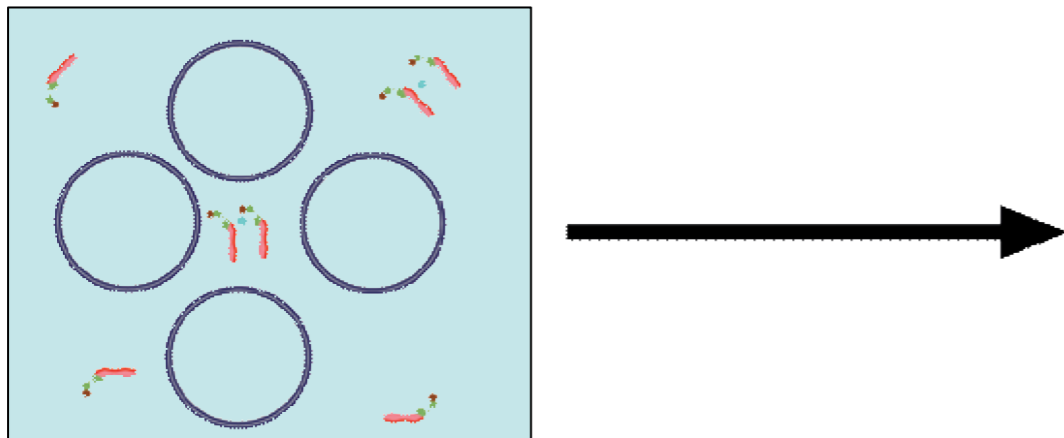
## Lipid matrices: Principle

*Courtesy of D. W. Deamer*



# Other Quasi-compartmentalization self-replication of early genetic/catalytic material

## Lipid matrices: Principle



Aqueous suspension

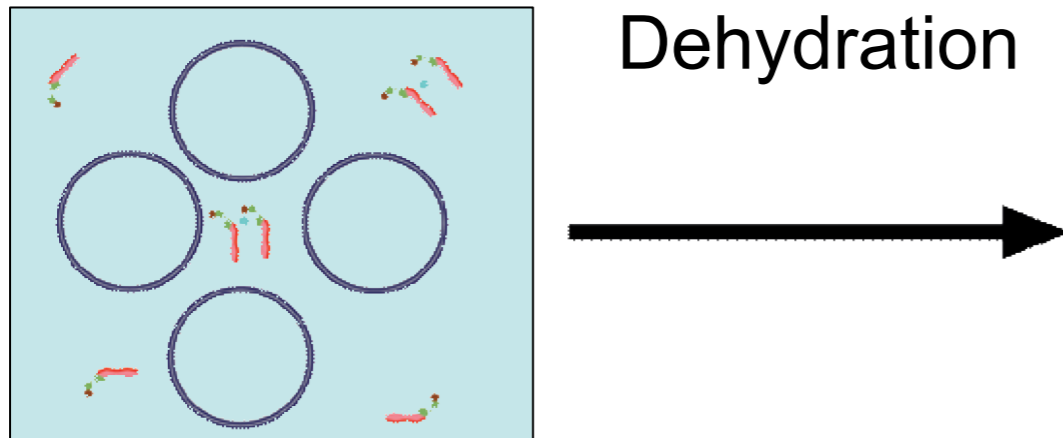
*Courtesy of D. W. Deamer*



UNIVERSITY OF SOUTHERN DENMARK

# Other Quasi-compartmentalization self-replication of early genetic/catalytic material

## Lipid matrices: Principle



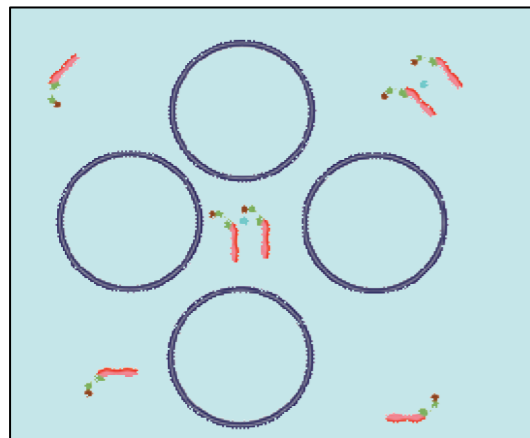
Aqueous suspension

*Courtesy of D. W. Deamer*



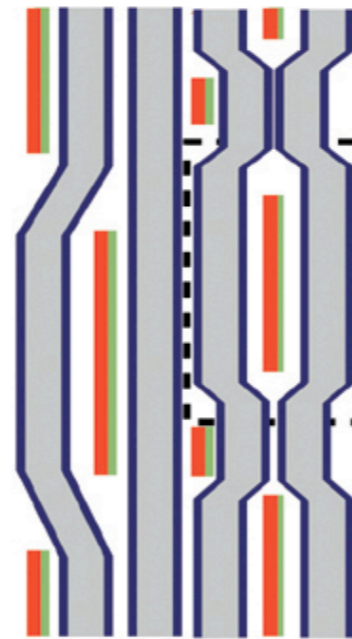
# Other Quasi-compartmentalization self-replication of early genetic/catalytic material

## Lipid matrices: Principle

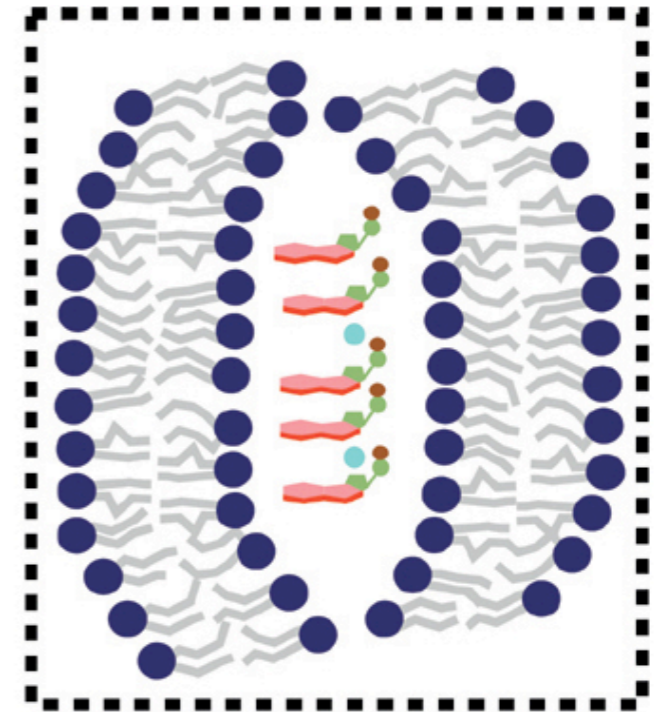


Aqueous suspension

Dehydration



Dried film

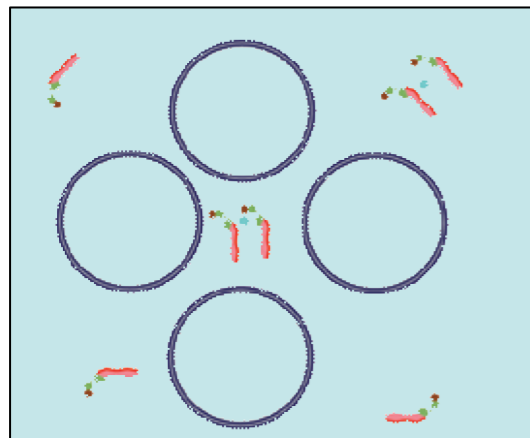


Liquid crystalline lipid phase



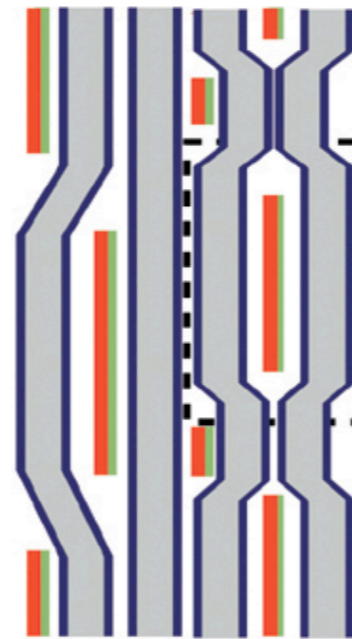
# Other Quasi-compartmentalization self-replication of early genetic/catalytic material

## Lipid matrices: Principle



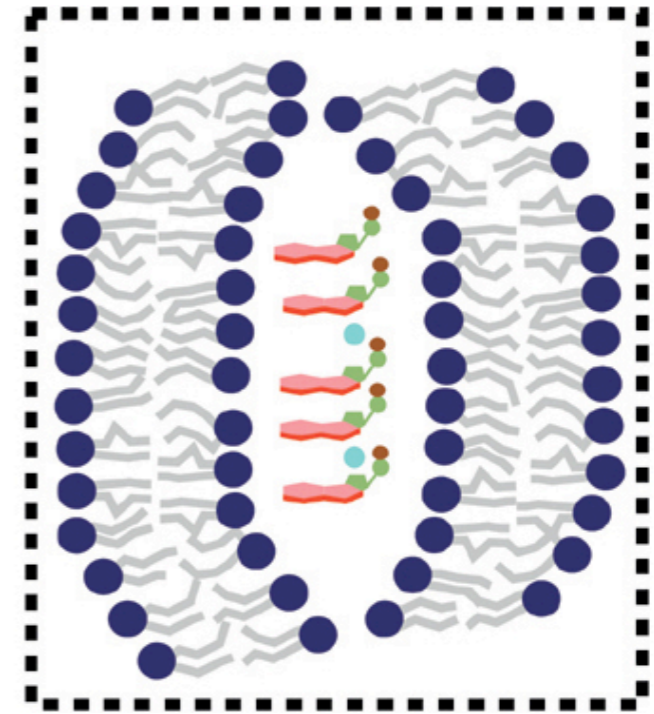
Aqueous suspension

Dehydration



Dried film

=



Liquid crystalline lipid phase

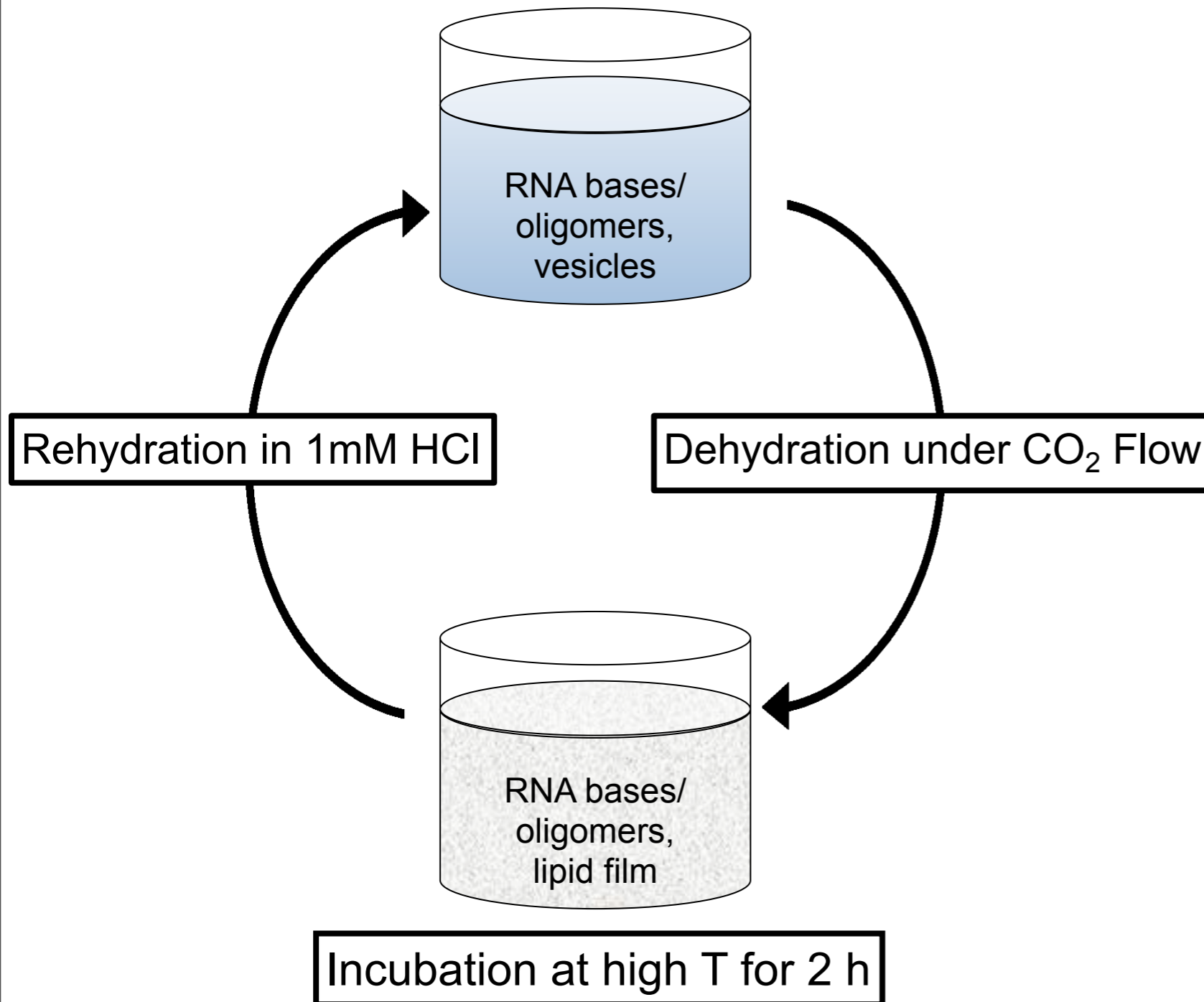


DOPC/DNA (2:1)  
ffEM: Pt,C shadowing

*Courtesy of D. W. Deamer*



# Dehydration-Rehydration cycles: Non-enzymatic RNA polymerization

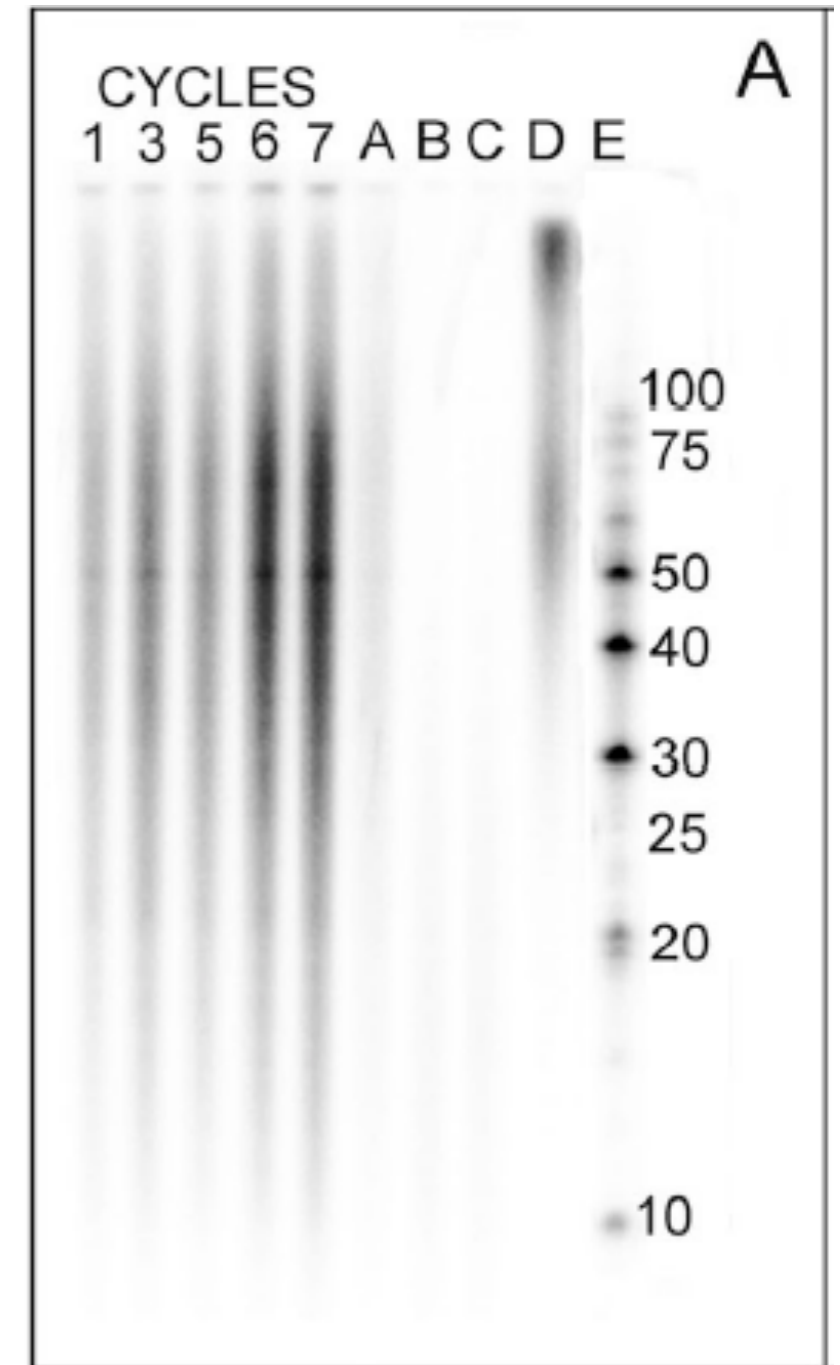
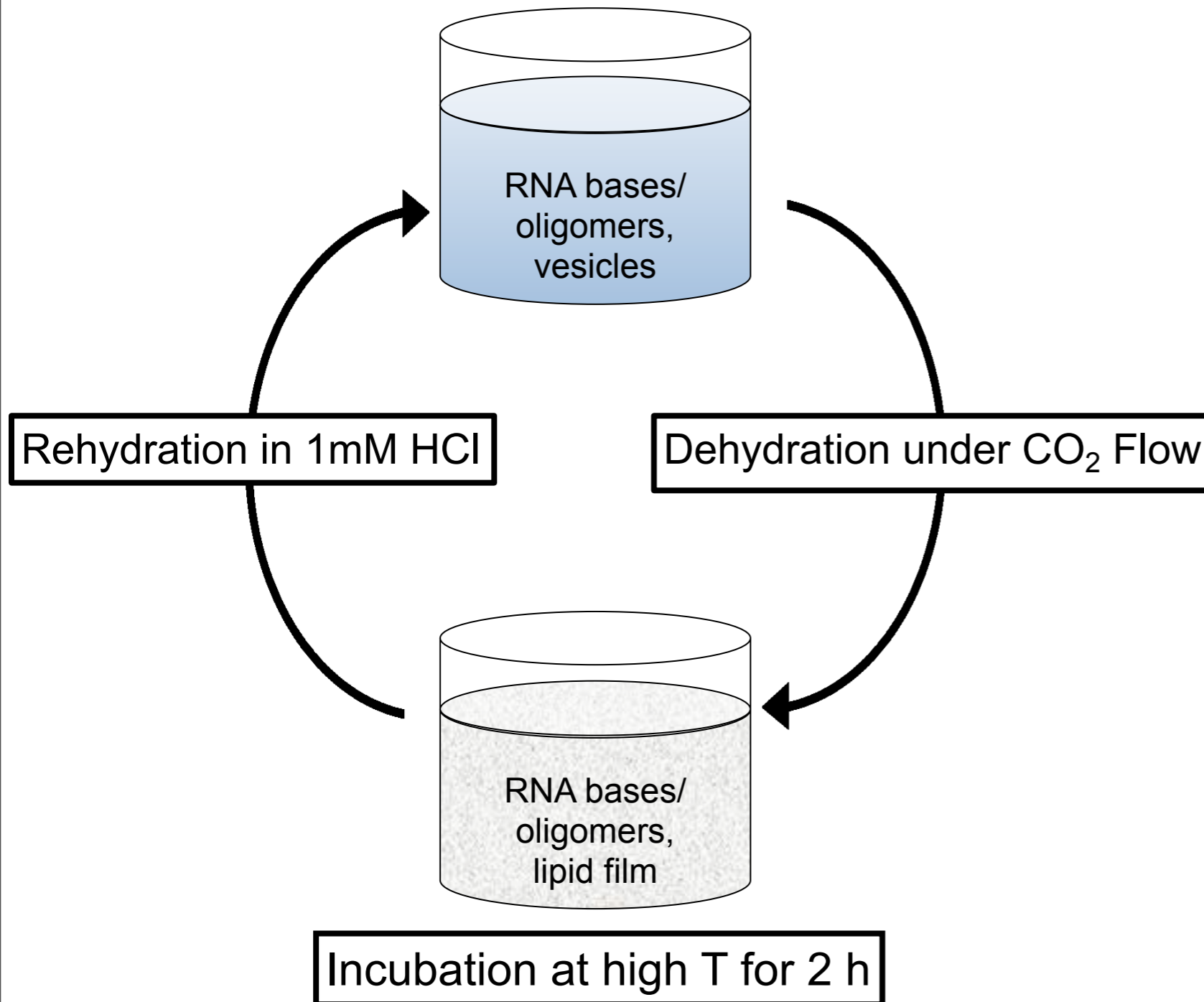


Rajamani, S., et al. *Orig Life Evol Biosph* (2008) 38:57-74



# Dehydration-Rehydration cycles: Non-enzymatic RNA polymerization

Phosphatidyl lipids and lysolipids

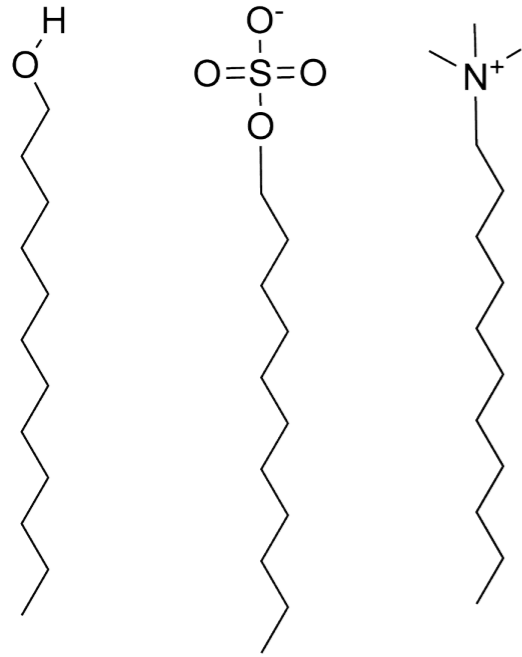


Rajamani, S., et al. *Orig Life Evol Biosph* (2008) 38:57-74





# Morphology of hydrated/dried “prebiotic” samples

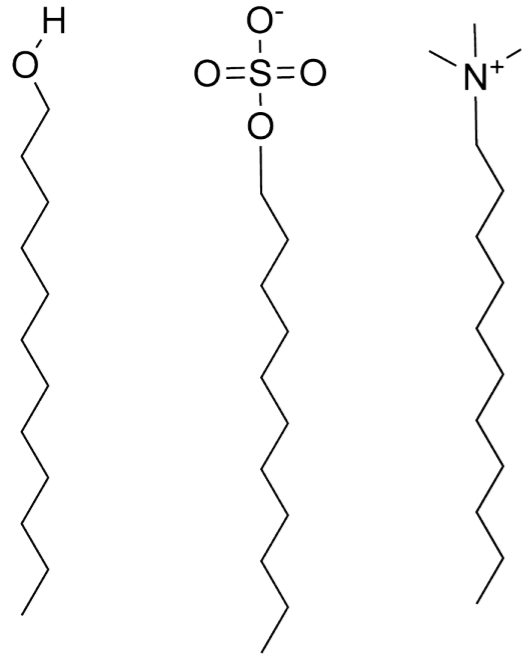


bar = 10  $\mu\text{m}$

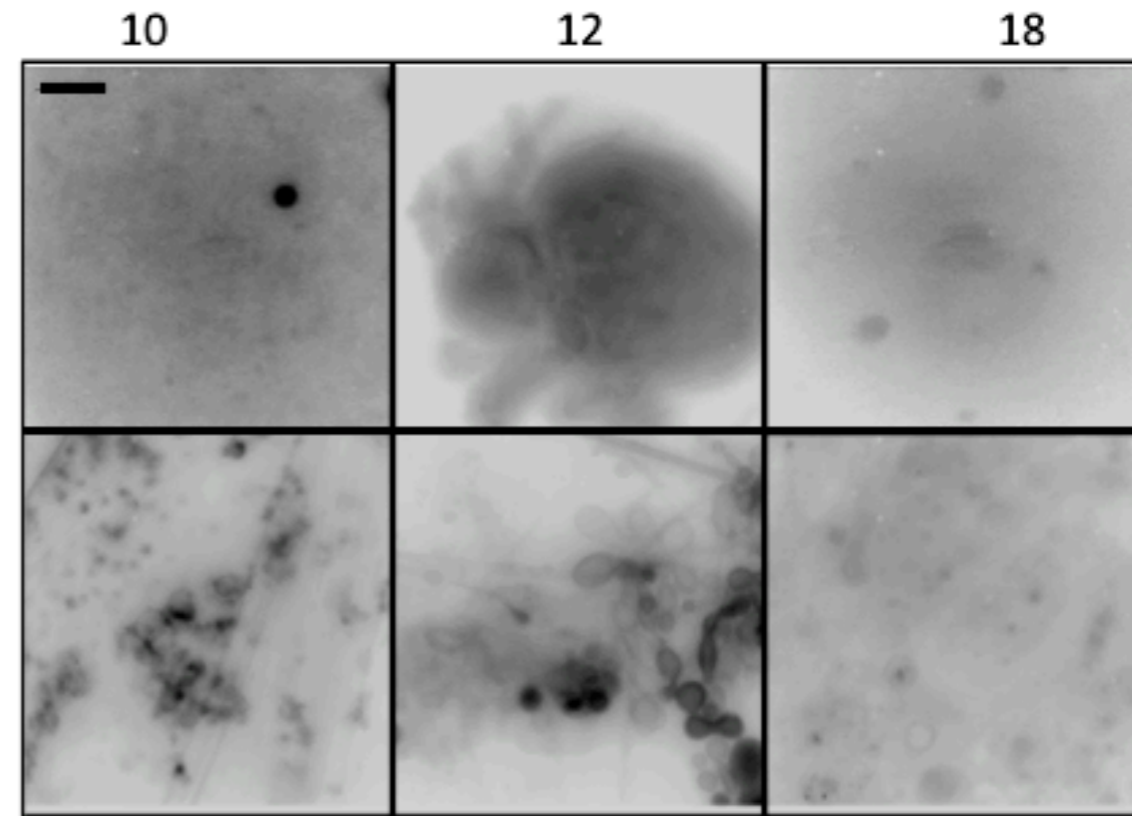


# Morphology of hydrated/dried “prebiotic” samples

Alkyl alcohols



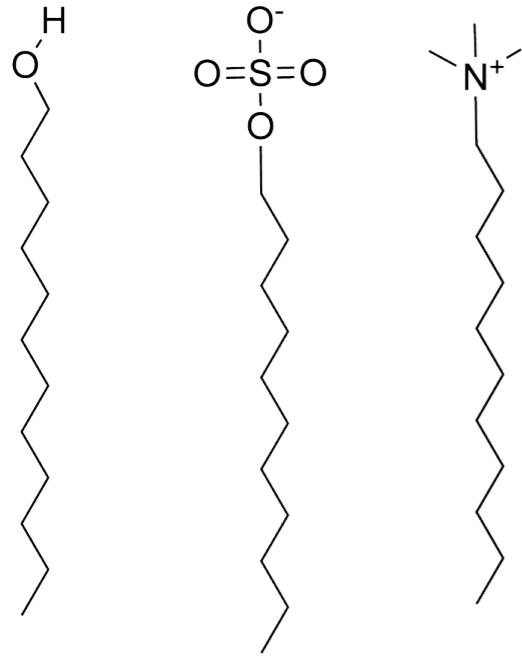
Alkyl trimethyl  
ammoniums



bar = 10  $\mu\text{m}$



# Morphology of hydrated/dried "prebiotic" samples

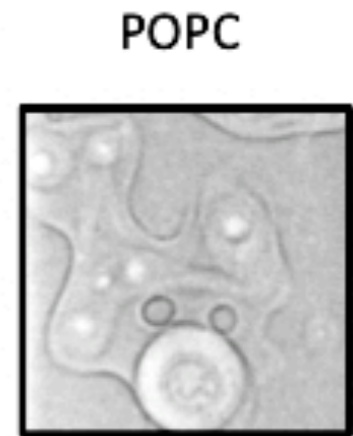
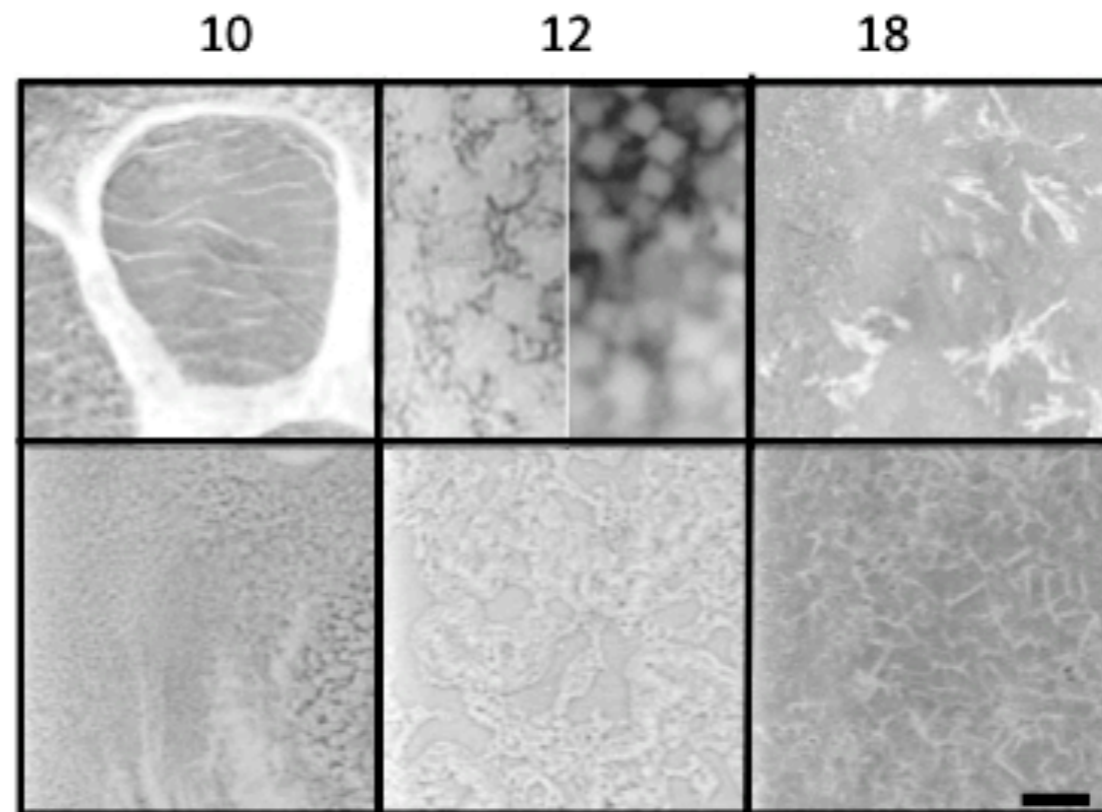
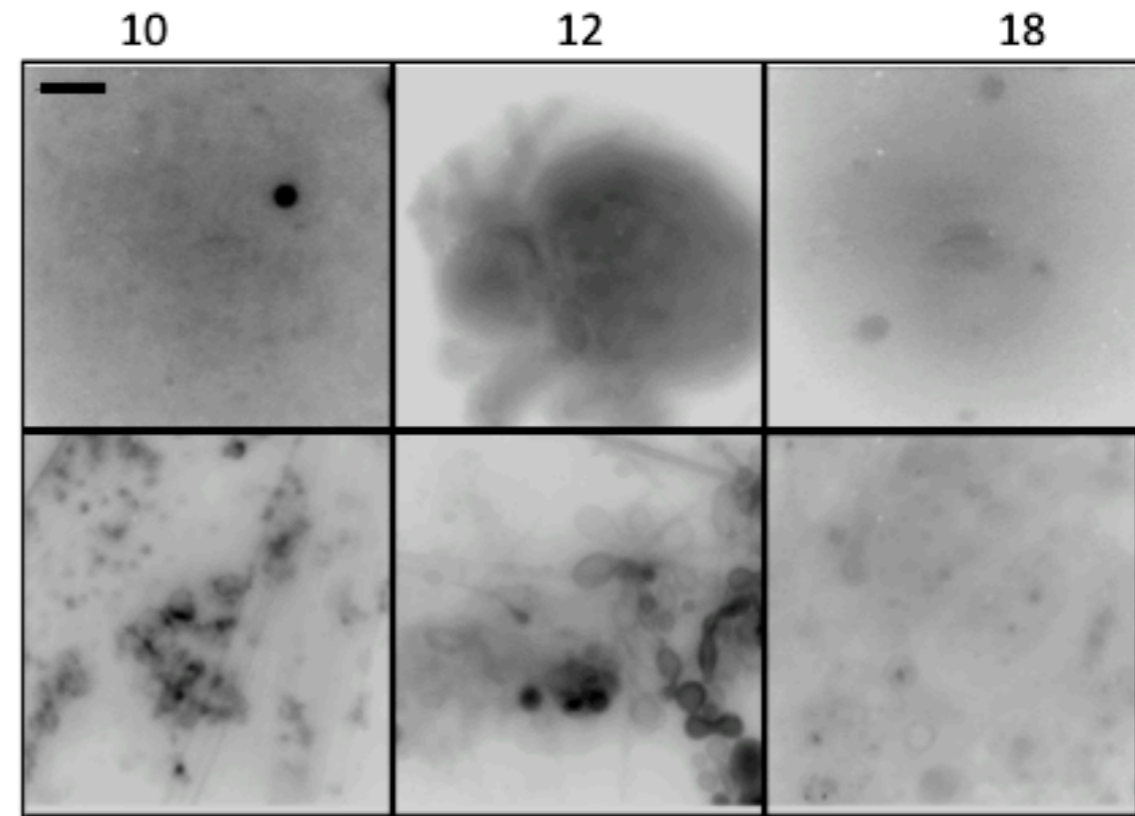


Alkyl alcohols

Alkyl trimethyl ammoniums

Alkyl alcohols

Alkyl trimethyl ammoniums

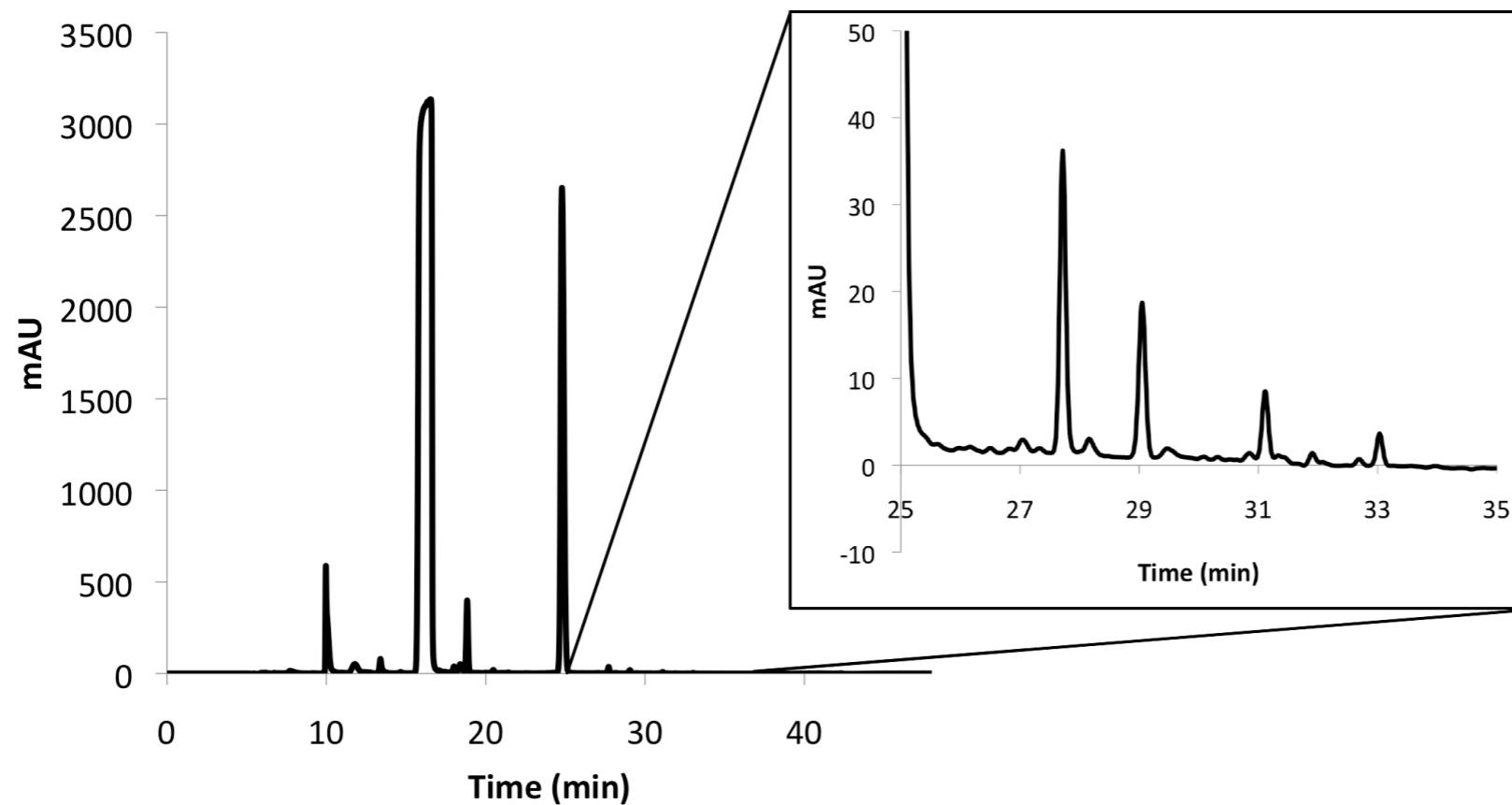


bar = 10  $\mu$ m



# Shorter product analysis

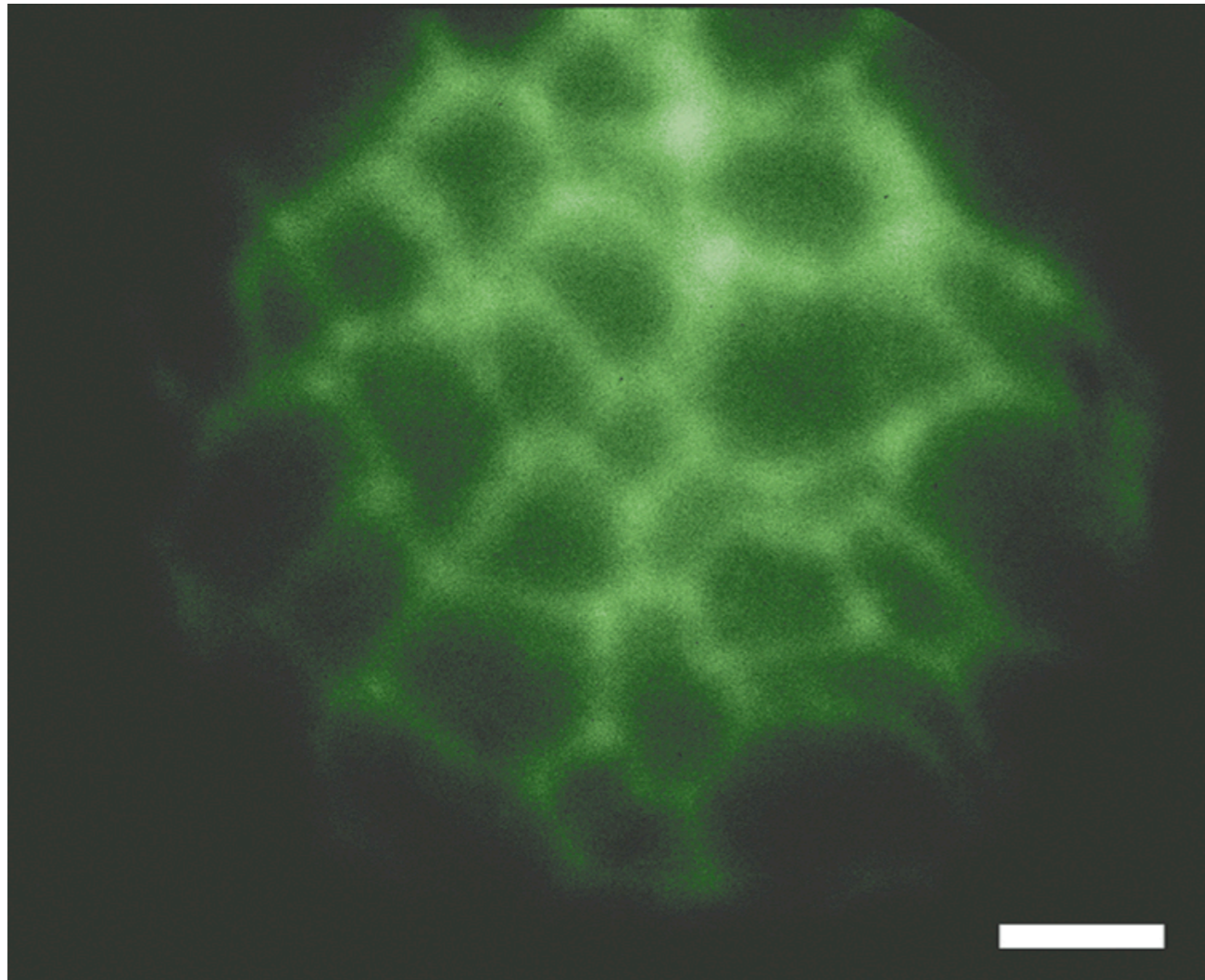
AMP + amphiphiles (mol ratio 1:2)



After reaction cycle 8 all samples showed formation of a product at 25 min.  
by RP HPLC



# Quasi-compartmentalization in EUTECTIC PHASE IN WATER-ICE



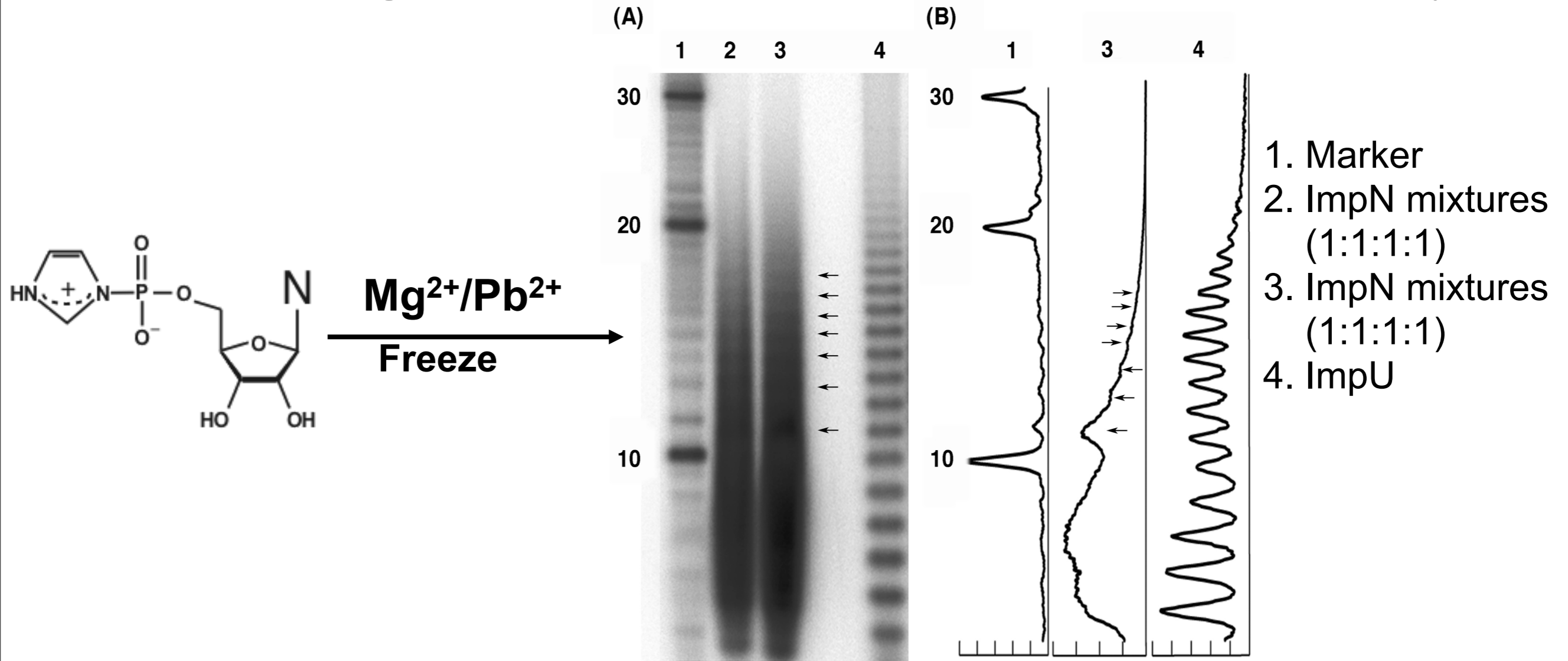
Upon the initiation of freezing, the concentration of the solutes increases which simultaneously lowers the freezing point of the residual solution.

Epifluorescence micrograph of monomer suspension used in self-condensation experiments. Acridine Orange was added to visualize the structures

# Self-condensation of monomers

Samples: < 5 mM activated monomers (ImpN), < 5.2 mM Mg(II), < 0.6 mM Pb(II)

Preparation: Mix @ 25 °C, freeze and maintain @ -18 °C for up to 40 days.



All mixtures tested same average yields (>80% incorporation, 50% ≥ 3-mer, up to 15- to 22-mer, equal incorporation of all nucleobases in mixed products, more 45% oligomers with 3'-5')

*Kanavarioti et al 2001 Astrobiology; Monnard et al 2003 J. Am. Chem. Soc*

# Non-enzymatic, template-directed polymerization: Steps towards self-replication

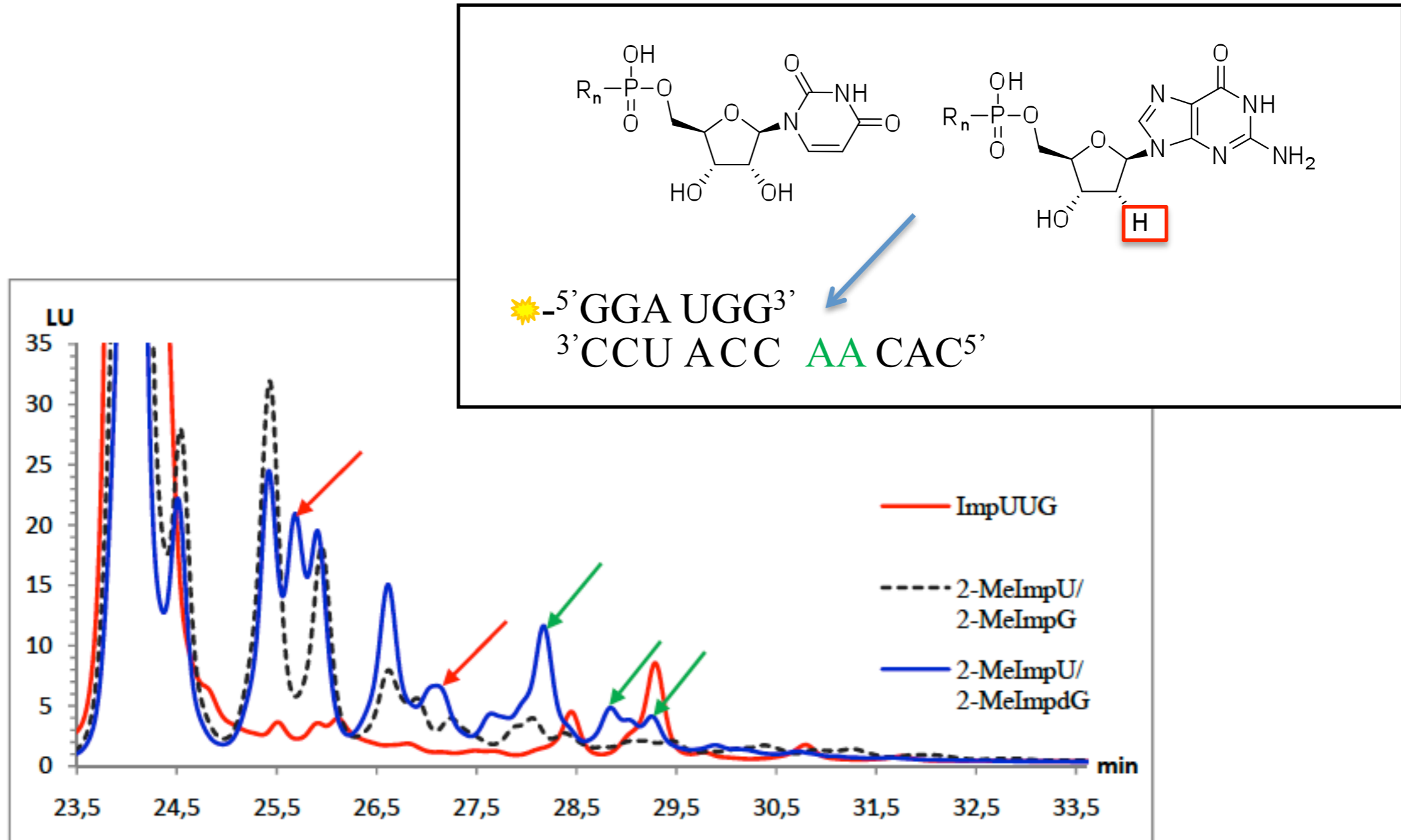
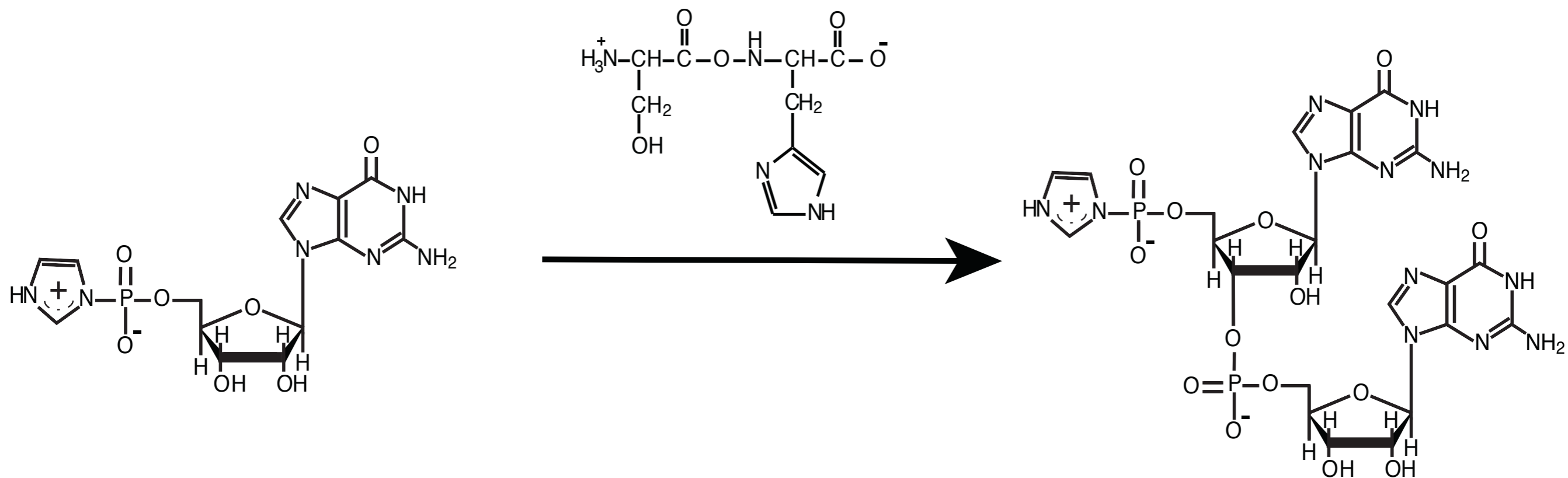


Figure 2.7-3: Experiment with 2-MeImpdG. An overlay of the chromatograms of the UUG ligation (PL17, red, 5d) with the experiment where dG was employed to “cap” the primer or its primer+(U)<sub>n</sub> elongation products. (PL18, blue). As a reference the analysis of the reaction with a ribonucleotide G is overlaid. (black, stipled). See text for the explanation of red and green arrows. Reactions were incubated at -18.4° C for 14d, using 1.75mM 2-MeImpU and 0.6mM 2-MeImpdG or 2-MeImpG in the presence of chim-P and t#U.



# Emergence of peptide catalysis



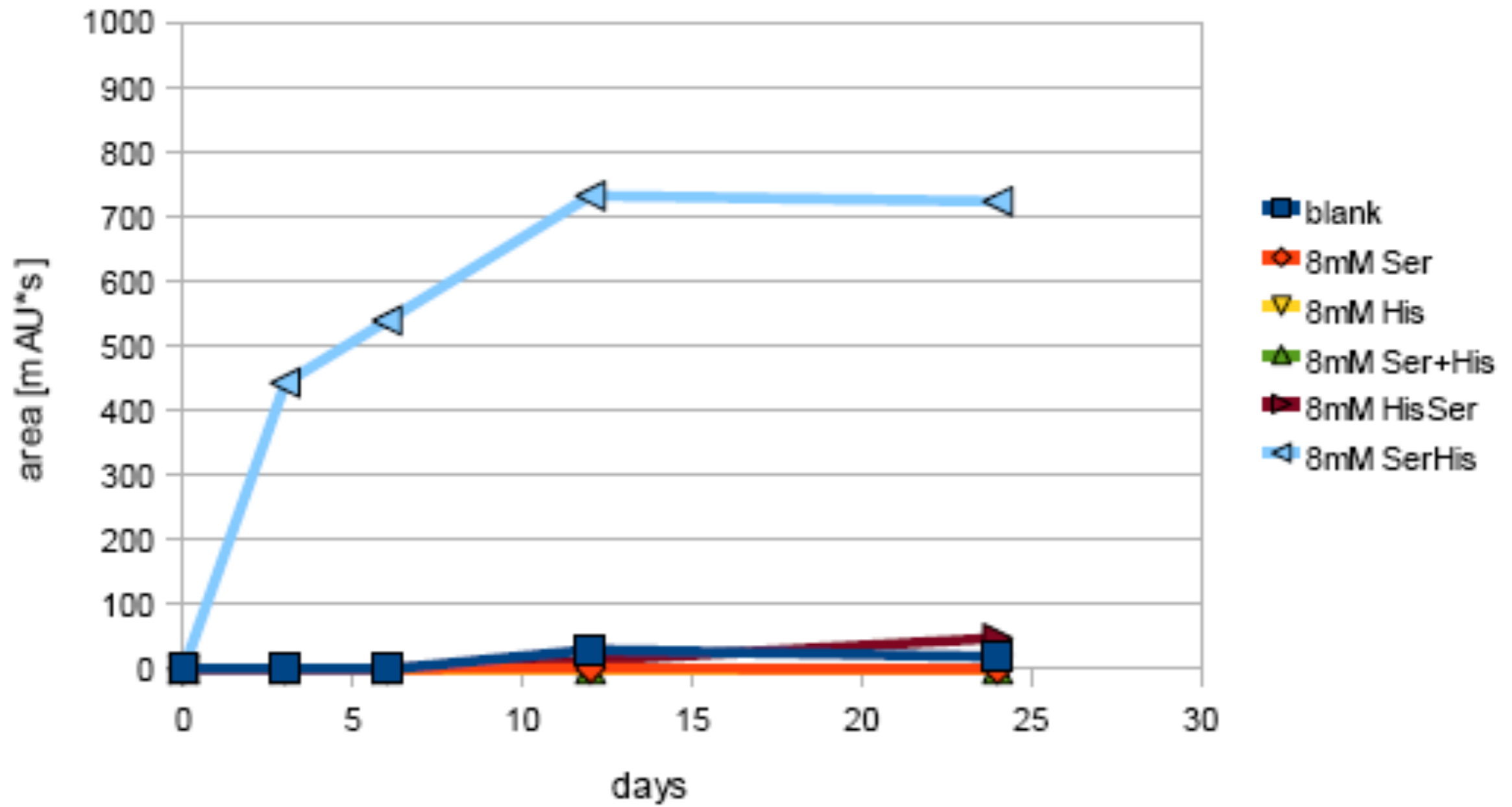
*Rafał Wieczorek*





# relative formation of n-mers

5 mM MES pH 6,5



# How to handle the research in the Origin of Life fields:

What do we still need to understand this emergence?

- Chemical composition
- Plausible pathways and reaction networks
- Sequence of events (i.e., the transitions)

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What do we still need to understand this emergence?

- Chemical composition
- Plausible pathways and reaction networks
- Sequence of events (i.e., the transitions)

Is it sufficient to study one aspect? (development of membranes, polymer based genetics and catalytic system...)

- Necessary to understand each aspect/component by itself but not sufficient
  - ➡ Systemic approach to the problem (system chemistry)
  - ➡ Less attachment to specific molecules and more considerations on classes of molecules and processes that are plausible with them.
  - ➡ Computing support (simulations, numerical analysis) to allow deconvolution of the complex interactions that exist in complex chemical systems.
  - ➡ Perhaps shift of paradigms is necessary: co-evolution of the various components of early living systems. Acceptance of sub-optimal yields (i.e. conditions for a given reaction) and perhaps unusual functions of some components (compared to today biology)

# Acknowledgements

## Collaborators

Dr. J. A. Bailey (LANL, scientist)  
**Dr. J.M. Boncella (LANL, scientist)**  
Dr. H.-J. Ziock (LANL, scientist)  
**Dr. S. Rasmussen (SDU, Prof.)**  
**Dr. D. W. Deamer (UCSC, Prof.)**  
**Dr. K. A. Nielsen (SDU, assistant Prof.)**  
**Dr. P.L. Luisi (Roma Tre, Prof.)**

## Post docs

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**Dr. J. Cape (LANL)**  
**Dr. M. Dörr (SDU)**  
**Dr. M. C. Wamberg (SDU)**  
**Dr R. Wieczorek (SDU)**

## Graduate students

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**W.H. Jørgensen (SDU, Ph.D.)**  
**P.L. Pedersen (SDU, Ph.D.)**  
**A. Albertsen (SDU, Ph.D.)**  
**P.M.G. Löffler (SDU, Ph.D.)**  
  
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**A. Zebitz Eriksen (SDU, BSc)**

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Danish National Science Foundation (current)  
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